



ISSN: 2230-9926

Available online at <http://www.journalijdr.com>

# IJDR

*International Journal of Development Research*

Vol. 11, Issue, 10, pp. 51203-51209, October, 2021

<https://doi.org/10.37118/ijdr.22900.10.2021>



RESEARCH ARTICLE

OPEN ACCESS

## PUTATIVE ONCOGENIC VIRUSES: SOME DATA ON BRAZILIAN SUBJECTS

Bernardo G. Tenório<sup>1</sup>, Isadora S. Bretas<sup>1</sup>, João P. R. A. Bernardes<sup>1</sup>, Munnah N. J. Mansour<sup>1</sup>, Sarah M. S. Napoleão<sup>1</sup>, Sheyla A. S. Rocha<sup>1</sup>, Thatiane L. Sampaio<sup>2</sup>, Elisiário C. V. Leitão<sup>3</sup>, Joaquim X. Silva<sup>3</sup>, Marcus V. S. Coimbra<sup>1</sup>, Edson J. M. Bello<sup>4</sup>, Elida C. G. M. Kanzaki<sup>1</sup> and I. Kanzaki<sup>1\*</sup>

<sup>1</sup>Laboratory of Bioprospection, University of Brasilia<sup>2</sup>Federal Institute of Brasilia/DF, Brazil;<sup>3</sup>Asa Norte Regional Hospital, Brasilia/DF, Brazil;<sup>4</sup>Central Laboratory of Public Health/LACEN, Brasilia/DF, Brazil.

### ARTICLE INFO

#### Article History:

Received 14<sup>th</sup> August, 2021

Received in revised form

28<sup>th</sup> September, 2021

Accepted 02<sup>nd</sup> October, 2021

Published online 30<sup>th</sup> October, 2021

#### Key Words:

HTLV-1/2, HPV, MMTV, HERVs, HCV, Cancer, Brazilians.

#### \*Corresponding author:

I. Kanzaki

### ABSTRACT

The concepts of cancer etiology have changed over the years, mainly based on molecular epidemiology studies and bioinformatics approaches. Until relatively recently, the most accepted theory of cancer etiology has dealt with the accumulation of gene mutations and the consequent cognate proteins dysfunction, but now some authors have argued against the proposed theory. The additional role of noncellular genes in the cause of malignancy, associated to environmental factors and host genetic background, has been proposed and mostly accepted by the scientific community. Some of our data from human populations in Brazil concerning cancer epidemiology, molecular and serological surveys, were conducted looking for the detection of putative oncogenic viruses, as the Human T-cell Lymphotropic virus/HTLV-1/2, Human Papillomavirus/HPV, the Mouse or Human Mammary Tumor Virus/MMTV, the Human Endogenous Retrovirus/HERVs and the Hepatitis C virus/HCV, in healthy and malignized human tissues. Generally, research work around the world suggests that 10 to 20 % of all human cancers are etiologically linked to oncogenic viruses, so if the presence of exogenous or endogenous virus sequences in the human DNA has any significance in the cancer etiology, it deserves further and continuous research work and discussion, always taking in account the human populations in their interacting surrounding environment.

Copyright © 2021, Bernardo G. Tenório et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Bernardo G. Tenório, Isadora S. Bretas, João P. R. A. Bernardes, Munnah N. J. Mansour, Sarah M. S. Napoleão et al. "Putative oncogenic viruses: some data on brazilian subjects", *International Journal of Development Research*, 11, (10), 51203-51209.

## INTRODUCTION

Despite some controversies, it is the common sense that cancer is a multifactorial event mainly determined by aging, as the cellular biochemical machinery, along the years, gathers nucleotide sequences coding for altered or non-functional proteins playing pivotal roles at different stages of the cell cycle (Wu *et al.*, 2018). Dysregulated cell function displays, naturally, a dynamic process intrinsically linked to DNA replication and consequent cell division, with important contributions of endogenous and exogenous factors (Lewandowska *et al.*, 2019). Living organisms are continuously under the direct or indirect influence of physical, chemical and biological agents and, the closest interaction with these environmental determinants, ultimately trigger mechanisms of evolution or life termination. Cancer seems to be an unsuccessful event for the host, that winds up with abnormal cells metabolism and growth, in a chaotic genomic and tissue

organization (Lewandowska *et al.*, 2019; Ben-David *et al.*, 2019; Davis *et al.*, 2019; Dahiya *et al.*, 2019). It is understandable and well-known that isolated events do not play any significative role in cancer etiology and evolution but, integrated and orchestrated molecular events, even in distorted physiological processes, have been well-characterized and proved to participate in the tumorigenesis and cancerization. Particularly here, we explore the eventual participation of the so-called oncogenic viruses in the etiologic processes of cell transformation and malignization (Akram *et al.*, 2017), by the detection of gene sequences of oncogenic viruses and humoral response to these agents. How do proteins coded by virus genomes could participate or contribute to the genesis of the genomic chaos in the host cell?

**The Human T-cell lymphotropic virus (HTLV):** The Human T-cell lymphotropic virus, also known as the Human T-cell leukemia virus, represents a group of viruses taxonomically positioned in the

retroviridae family, *deltaretrovirus* genus. The Primate T-cell lymphotropic viruses (PTLVs) group include deltaretroviruses infecting human and nonhuman primates, the HTLVs and STLVs, respectively. There are, presently, 4 types of HTLV (HTLV-1, HTLV-2, HTLV-3 and HTLV-4) and the corresponding 4 STLV types (STLV-1, STLV-2, STLV-3 and STLV-4). The Simian T-cell lymphotropic viruses (STLVs) are closely related to the HTLVs as phylogenetic studies point out that humans were initially infected by STLVs, in Africa and Asia, and these viruses infecting human hosts evolved to HTLVs. It is claimed that hunting of simians by human beings, and feeding on them, as also cohabitation of these two hosts species, such like pet animals or in zoos, have favored deltaretroviruses zoonotic transmission (Hron *et al.*, 2019; Narat *et al.*, 2018; LeBreton *et al.*, 2014). Among the PTLVs, the HTLV-1 is the most studied virus as it is etiologically linked to neuromyelopathy, the HTLV-1 associated myelopathy (HAM), also known as Tropical Spastic Paraparesis (TSP), and a leukemogenic process, the Adult T-cell leukemia/lymphoma (ATLL). HTLV-1, as its designation stands for, has tropism for T lymphocytes, being transmitted by biological fluids containing these cells, as mucosal fluids, breast milk and blood, of infected subjects, therefore, the virus is acquired by sexual intercourse, breast milk feeding and blood transfusion (Rocamonde *et al.*, 2019). Neuromyelopathy is characterized by the spinal cord chronic inflammation or myelitis, caused by strong immune response to HTLV-1 infected cells, in the central nervous system milieu (Enose-Akahata *et al.*, 2019), while leukemogenic processes arise from TCD4<sup>+</sup> abnormal proliferation, after cell physiological dysregulation in the event of HTLV-1 infection (Wong *et al.*, 2020). ATLL, a CD4<sup>+</sup> T cell malignancy, is rare, aggressive and a hard-to-treat hematological malignancy, differing its onset in Japanese subjects around 70 years, and 50 years among Americans and Europeans (Cook *et al.*, 2020), and not well-determined among other Asians and African populations. About 4 out of 100 HTLV-1 infected subjects slowly progress to ATLL, with this proportion mainly determined by ethnicity and geographical distribution.

It is worthwhile to mention that, as ruled for all malignancies, the sole HTLV-1 infection is not enough to cause ATLL, but comorbidities, the host genetic background and environmental factors synergize for the pathogenesis development, despite the clonal integration of HTLV-1 proviral DNA in all patients' ATLL cells (Iwanaga, 2020). One of the proteins encoded by HTLV-1, the p40 tax oncoprotein, plays a pivotal role activating the canonical and non-canonical forms of the host nuclear transcription factor  $\kappa$ B (NF- $\kappa$ B), leading to the expression of numerous viral genes involved in inflammation processes. The hijacking and aberrant activation of the cell ring finger protein 8 (RNF8) exert a critical role in the cell DNA repair mechanisms or cellular DNA damage response (DDR), that in later instances generates host cell genomic instability, as DNA lesions are not repaired (Zhi *et al.*, 2020). According to Ameer *et al.* (2020), it is not well-understood the molecular mechanisms of HTLV-1 induced splicing modifications and/or the interplay between transcription and splicing mechanisms. Physiological processes that determine cells' fate comprehends an intrinsic biochemical interaction of endogenous and exogenous factors affecting RNA processing in the transcriptome and proteome plasticity, ultimately characterizing the cells' phenotype (Toyoda *et al.*, 2019; Williams and Gilmore, 2020). Besides p40tax, the negative-stranded virus encoded helix-basic-zipper-protein (HBZ) expressed in HTLV-1 chronically infected cells contrasts to the low levels of Tax and Gag proteins production. Tax and HBZ have antagonistic roles in the cell signaling pathways, as tax activates NF- $\kappa$ B, NFAT and AP-1, while HBZ inhibits them. Also, HBZ activates the TGF- $\beta$  /Smad pathway and tax inhibits it. Therefore, to rescue HTLV-1 infected cells of tax deleterious effects, HBZ promotes cell survival and proliferation, as an essential factor for leukemogenesis and the inflammation process in HAM/TSP, and minor HTLV-1 related pathologies (Toyoda *et al.*, 2019). Another accessory protein, the p30 (tof protein), encoded by the doubly spliced Tax-ORF II mRNA, puts in place transcriptional and post-transcriptional activities blocking virus replication and preventing interferon synthesis in HTLV-1 infected myeloid cells, consequently impairing innate and adaptive host immunity. Some studies point out

that the tof protein participates in the mechanisms of gene expression, cell cycle progression and DNA damage response, raising the possibility of its role in T cell transformation, being also essential for virus infection and persistence (Matsuoka and Mesnard, 2020; Moles *et al.*, 2019).

**The Human Papillomavirus (HPV):** The members of the *Papillomaviridae* family are non-enveloped small virions of 50-60 nm in diameter. Its capsid encloses a circular double stranded DNA that extends approximately for 8 kb. Up to date, 32 genera have been described in this family, distributed in the vertebrate classes, except amphibians. Human papillomavirus (HPV) is restricted to the genera Gamma, Mupa and Nupapapillomavirus, incorporating more than 200 different HPV types (Wong *et al.*, 2020; Ata *et al.*, 2021; Day *et al.*, 2019; Durzynska *et al.*, 2017). Just about 4-5 % of all human cancers have HPV etiologic contribution, despite the majority of HPV infections shows as benign growth expressed mainly as warts of epithelial origin, as also present in healthy skin, even though actinic keratosis, epidermal cysts, lichen sclerosus, psoriatic plaques, seborrheic keratosis and skin tags have also been claimed as related to HPV infection. There are evidences that the fate of HPV infection that culminates in cancer or other less serious skin condition depends on the type of HPV and epithelial cells' genes responses to the infection (Rector and Van Ranst, 2013; Prati *et al.*, 2018; Ljubojevic and Skerlev, 2014). Papillomavirus has tropism for epithelial cells, in both cutaneous and mucosal environments, including the oral cavity and anogenital tract. The virus gains space through tissues' microlesions, and specifically infects the host's dividing basal stratified epithelia, spreading and remaining the viral progeny persistently and latently interacting with the host cells' biochemical machinery for years (Al-Eitan *et al.*, 2020; Spurgeon *et al.*, 2019). Depending on the environmental and host factors, most of the persistent papillomavirus infection could be cleared by the host immune system or triggers, more or less, benign skin lesions. In the worst-case scenario, it is constantly proposed the concomitant role played by the mentioned factors as the host exposure to environmental UV irradiation, tobacco smoking, alcohol consumption, HIV infection and host hormonal disbalance, particularly among women, concerning estrogen metabolites, in the evolution of papillomavirus persistent infection to neoplasia development (McBride, 2017; Gheit, 2019; Ding, 2019). Besides the interplay among these factors, papillomaviruses are classified in the high-risk and low-risk groups according to the carcinogenic potential. Therefore, the HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58 and 59 are considered the high-risk papillomaviruses, and the others, the low-risk. Independently of high or low-risk papillomavirus infection, there are assumptions that any persistent papillomavirus infection could lead to malignization (Ramírez-López *et al.*, 2019; de Sanjosé, 2018). Papillomavirus infection is circumscribed to the wounded basal stratified epithelial layer, through initial interaction of L1 and L2 viral capsid proteins with epithelial cell membrane heparan sulfate proteoglycans and host cell factors generated by the microlesions as growth factors and cytokines (Egawa and Doorbar, 2017). The E6 and E7 papillomavirus proteins, highly produced by infected cells, are the major determinants of associated diseases, from relatively benign warts to aggressive malignization processes in the anogenital to head and neck body areas (Buck *et al.*, 2013).

**The Mouse/Human mammary tumor virus (MMTV/HMTV):** Human breast cancer is the second cause of mortality among women. Its etiology is multidiverse as the cancer per se, but mainly determined by hormonal factors. Apart from environmental and genetic factors, infectious agents have been investigated for their causative role in the cancerization process, e.g., the Herpes simplex virus and the Human papillomavirus (Vonsky *et al.*, 2019; Fahad, 2019; Lehrer and Rheinstein, 2019). Nevertheless, a sole viral agent, the Mouse Mammary Tumor Virus (MMTV), transmitted from lactating mice to their offsprings, also named Bittner Milk Agent, in honor to J.J. Bittner since his initial studies in 1936, proved to be etiologically linked to mouse breast cancer (Bittner, 1936). At that time, there were already reports concerning the above-mentioned factors involved in the etiology of breast cancer, mainly utilizing mice

as a model (Lehrer and Rheinstein, 2019). A MMTV homologue, tentatively denominated Human Mammary Tumor Virus (HMTV) has been detected in human breast tissues, as viral particles by electron microscopy, and the viral RNA, even though it is a hot topic the role played by this virus in the etiology of human breast cancer (Noon *et al.*, 1975; Perzova *et al.*, 2017; Al Dossary *et al.*, 2018). A controversial data obtained in Poiesz laboratory (Perzova *et al.*, 2017) showed that human DNA samples, from healthy and breast cancer tissues, amplified for MMTV *env* primers, as also mouse DNA of several species, but the authors evaluated the number of copies obtained, and concluded that the amplified DNA samples were from laboratory environment contamination, arguing also that the low number amplification copies would not explain the possible etiologic role played by MMTV in human breast cancer, according to one of the proposed mechanisms of insertional mutagenesis, activating neighboring proto-oncogenes, as the fibroblast growth factor receptor 2/*Fgfr2* and the wingless/*Wnt-1*, in the MMTV proviral integration site (Nartey *et al.*, 2017; Li *et al.*, 2000). Nevertheless, a recent study by Lessi *et al.* (2020) confirmed the pioneering studies of Bogo laboratory (Nartey *et al.*, 2014), of MMTV-like virus in human breast cancer, when finding MMTV-like sequences in the DNA of human mandibula fossils, dated back between the Copper age and the 17<sup>th</sup> century, therefore it is estimated that MMTV jumped species, from mouse to man, around 4,500 years ago (Lessi *et al.* 2020). One of the 7 MMTV genes, *Sag*, commonly described in the genome of certain viruses and bacteria, codes for a superantigen, a protein that strongly and excessively stimulates T cells polyclonal activation by an uncommon mechanism. This viral protein is claimed to be linked to MMTV infection and carcinogenesis (Us, 2016; Lawson and Glenn, 2021). Cells, of the immune system and of the breast epithelia, express receptors for MMTV infection, of both endogenous and exogenous origin, at least in mice, leading the retroviral integrated zygote chromosome to exhibit viral antigens which generate immunologic tolerance during T lymphocytes clonal selection in the thymus, allowing exogenous MMTV infection, host cell chromosome integration and insertional mutagenesis, that ultimately activate proto-oncogenes promoting the carcinogenic process, certainly modulated by endogenous and epigenetic factors, as also T cell polyclonal activation by *Sag* super antigen, which consequentially induces the expression of inflammatory cytokines. According to reports of different research groups, these mechanisms occur slowly, taking years to progress to breast cancer (Fahad, 2019; Nartey *et al.*, 2017; Us, 2016; Lawson and Glenn, 2021; Dudley *et al.*, 2016).

**Human Endogenous retroviruses (HERVs):** Genomes of all vertebrate species have homologous sequences to genes of exogenous retroviruses. It is commonly accepted that ancient infection of multicellular organisms by exogenous viruses, particularly retroviruses, followed by its genome integration into the host cells' germ line chromosome, perpetuated the acquisition of viral nucleotide sequences by host organisms. Katsura and Asai (2019) stated that "accumulation of viral sequences has created the current human genome". About 5-8% of the human genome is composed of endogenous retroviral nucleotide sequences, exerting a plethora of functions by the cognate proteins expressed, some of them fundamental for the host reproduction and survival, but also, it is argued the pathological role played by these endogenous retroviral sequences, mainly in autoimmune diseases and cancer. According to Gao *et al.* (2021), HERVs expression acts as accessory factors in the carcinogenic process activating oncogenic signaling pathways and inhibiting tumor suppressor genes. The taxonomy of the Endogenous Retroviruses is not yet defined, the present time nomenclature utilized is based on distinct features, anyway efforts to establish a rational classification is undertaken as proposed by Gifford *et al.* (2018). For example, one of the most important HERVs, the HERV-K, has its designation based on the usage of lysine tRNA, but the applied criteria will not cover other Endogenous Retrovirus representatives. Also, HERV-K is a recently acquired group of HERVs profusely spread in the human genome. Some of these HERV-K integrated copies displays intact open reading frames capable of transcription and translation, linking its activity to indispensable role in the embryogenesis mechanisms. Among members of HERV-K group,

HML-2 is the most actively transcribed and its aberrant expression has been associated to disease pathophysiology as certain types of cancer and neurodegenerative processes (Garcia-Montojo *et al.*, 2018). The HML-2 abnormal *env* gene expression produces two viral oncogenic polypeptides, Np9 and Rec, besides their role in autoimmune diseases. Experimental therapeutic approaches target the *env* gene products with monoclonal antibodies for multiple sclerosis treatment (Grandi and Tramontano, 2018). Another HERV member, HERV-W, encodes a syncytin-1 protein, essential in the trophoblast formation, therefore playing a major role in the fetal physiology. The fine-tuning regulatory mechanisms in the expression of HERVs genes is determinant for a physiological or pathological performance in human organisms, as similarly for ERVs in other eucaryotic pluricellular species, mainly vertebrates (Kristensen and Christensen, 2021).

**The Hepatitis C Virus (HCV):** Hepatitis of viral etiology encompasses agents of 4 different families (*Hepadnaviridae*, *Picornaviridae*, *Hepeviridae* and *Flaviviridae*) and still one of an unsigned family (Delta virus genus), but not all claimed to be oncogenic (Ebrahim, 2011). Molecular and immunovirological studies characterize the Hepatitis C Virus infection in primates (*Homo sapiens* and *Pan troglodytes*) and also, reassure its link to liver pathologies, including cancer, conclusively performed by Houghton's group, but not successfully without many years of non-A non-B hepatitis viruses' investigation by other research groups (Houghton, 2009; Choo *et al.*, 1989). Classified in the *Flaviviridae* family, the enveloped HCV particles range from 30 nm to 140 nm in diameter, presenting a single-stranded RNA genome of positive polarity with 9600 nucleotides, coding for a long polyprotein cleaved into structural and replicative polypeptides (Sagan *et al.*, 2015; Piver *et al.*, 2017). It is estimated that 0.91 % of the entire world population is chronically infected by HCV, which progress to cirrhosis leading to the most common associated pathologies as hepatocellular carcinoma, cholangiocarcinoma and B-cell non-Hodgkin lymphoma (Navas *et al.*, 2019; Khatun *et al.*, 2021; Spearman *et al.*, 2019). The continuous expression of HCV proteins resulting in simultaneous disturbed cell signaling processes, oxidative stress and chronic liver inflammation reinforced by the host and environmental factors, plays an orchestrated and long sustained activity to progression of cancerous pathologies and other life-risking diseases (Virzi *et al.*, 2018). Oxidative stress is the main biochemical cell erosion process ubiquitous in disease mechanisms, triggered by many agents including HCV, mostly implicated in cirrhosis progression to cancer, as the hepatocellular carcinoma, the hallmark of HCV persistence. Elevated levels of superoxide and hydrogen peroxide in hepatocytes triggered by phagocytes' production of NADPH oxidase responding to HCV infection, and glutathione reduction, promote cell death and Hepatitis C virions progeny replication. Counterbalance mechanisms in the redox microenvironment contributes to persistent infection which dysregulate the MDM2-p53 axis, or MDM2 overactivation and p53 inactivation, as a feedback mechanism, besides virus transformed hepatocytes protection from cellular immune response by the depletion of Kupffer cells, stellate cells and CD4<sup>+</sup>T cells (Anticoli *et al.*, 2019; Cao *et al.*, 2020).

## DISCUSSION

Our previous work on HTLV-1/2 epidemiology, in northern Brazil, demonstrated virus circulation in both healthy and cancer patients of different ethnicities, including Amazonian Amerinds, even though our initial collaborative research work, in early years of 1985, with Maruyama's laboratory in Japan, did not distinguish HTLV-1 from HTLV-2 (Nakauchi *et al.*, 1990; Nakauchi *et al.*, 1992; Kanzaki *et al.*, 1997), but later on, we were aware that HTLV-2 was responsible for the high prevalence among Amazonia Amerinds, except our findings of HTLV-1 positivity among Waiãpi amerinds inhabiting forest areas in the border of the Brazilian Amãpa state and French Guyana (Barros Kanzaki and Casseb, 2007). Cervix uterine cancer patients exhibited relatively elevated HTLV-1 prevalence (Nakauchi *et al.*, 1990; Kanzaki *et al.*, 1997), but scarce data confirmed our results, as

**Table 1. Synopsis of HTLV-1/2, HPV, HCV, MMTV and HERV-K analysis on human groups in Central/Northern region of Brazil**

Viral Agent	Method	Primer/gene region	Number of Subjects	Positive
HTLV-1/2 <sup>1</sup>	Immuno Assay PCR	N.A. <sup>7</sup> A/B/C/D/E/F/G/H <sup>8</sup>	156 HIV/AIDS <sup>11</sup> ; 35 breast cancer <sup>11</sup> 35 breast cancer <sup>11</sup>	6 HIV 0
HPV <sup>2</sup>	RT/PCR <sup>6</sup>	HPV L1 genotype-specific fragments	38 head and neck cancer <sup>11</sup>	1 (HPV 70)
HCV <sup>3</sup>	Immuno Assay	N.A. <sup>7</sup>	288 healthy <sup>12</sup> ; 156 HIV <sup>11</sup>	3 healthy; 15 HIV
MMTV <sup>4</sup>	PCR	brt1/brt2/brt3/brt4( <i>env</i> ) <sup>9</sup>	20 healthy <sup>11</sup> /15 breast cancer <sup>11</sup>	0
HERV-K <sup>5</sup>	PCR	fwGAGherv/rwGAGherv( <i>gag</i> ) <sup>10</sup>	35 head and neck cancer <sup>11</sup>	31

<sup>1</sup>Human T-cell Lymphotropic Virus Type 1/2; <sup>2</sup>Human Papillomavirus; <sup>3</sup>Hepatitis C Virus; <sup>4</sup>Mouse Mammary Tumor Virus; <sup>5</sup>Human Endogenous Retrovirus-K; <sup>6</sup>Real Time Polymerase Chain Reaction; <sup>7</sup>Not Applicable; <sup>8</sup>A(5'-CTCCTTCCCCACCCAGAGA-3')/B(5'-GGGTGGGTCCCATGTATCCATT-3')/C(5'-CTCCTTCCCCACCCAGAGA-3')/D(5'-GTTGGTTCAGGCATCCATT-3')/E(5'-AGAAGTACCCGCACCTCAA-3')/F(5'-GGTGAGCTCGAGGC AATTGTTTC-3')/ G(5'-GCAAGAAAGTGCTCGGTG-3')/ H(5'-CTACTCAGTGTGGCAAAGGTG-3'); <sup>9</sup>brt1(5'-CCTCACTGCCAGATC-3')/brt2 (5'-TACATCTGC CTGCCTGTGTAC-3')/ brt3(5'-ATCTGTG GCATACCT-3')/brt4(5'-GAATCGCTTGGCTCG-3'); <sup>10</sup>fwGAGherv(5'-GGGGCCATCAGA GTCTAAACC-3')/rwGAGherv(5'-TGATAGGCTACTTGGCGGTGG-3'); <sup>11</sup>Subjects from Central region of Brazil (Brasília); <sup>12</sup>Subjects from Northern Brazil (Marajó Islands).

published by Du *et al.* (2019) assessing the potential association of HTLV-1 to endometrial carcinoma; in the Yucatan peninsula, Góngorra-Bianchi *et al.* (1997) showed that Mexican Mayan descendants with high incidence of cervix uterine cancer, yielded HTLV-2 positivity, in both cancer and healthy women, as expected; as well, in Japan and Jamaica, HTLV-1 markers were detected among cervix uterine cancer patients (Taguchi *et al.*, 1988; Strickler *et al.*, 1995). HTLV-1/2 etiologic involvement in cervix uterine cancer is elusive despite its high statistical probability. As commonly substantiated, cervix uterine cancer is etiologically linked to HPV persistent infection, being both, HTLV-1/2 and HPV, mainly sexually transmitted, and mostly by promiscuous behavior, despite HTLV-1/2 infection be usually circumscribed to nuclear families. In addition, radiotherapy procedures represent an important variable among cancer uterine patients that could contribute to HTLV-1/2 expression (Kanzaki *et al.*, 1997; Tanaka *et al.*, 2019) furthermore, we still reported HTLV-1 positivity among health care workers exposed to ionizing radiation as already discussed elsewhere (Kanzaki, 2018). Based on these findings, of HTLV-1 and HTLV-2 expression, it could be reinforced the hypothesis of ionizing radiation protagonism in the activation of host cell integrated HTLV-1/2 genes (Barros Kanzaki, 2006).

In vitro and ex vivo studies aim to explain in vivo phenomena which accounts for Koch postulates (Hosainzadegan *et al.*, 2020) that is under review, so it would infer, for example, that virus-associated cytopathology and related dysregulated cell signaling, not necessarily and definitively prove the etiologic role played by some viruses in cancer etiology. As already stated, environmental factors and host genetic background are associated with the genesis of cancer and, why not, also, contributing to viral genes expression? Recent serological survey performed by us (Table 1), did not detect HTLV-1/2 among patients attended in hospitals at the Central region of Brazil, with different histological types of neck, head and breast cancer (Mansour and Kanzaki, 2018), but some HIV infected subjects of this same area showed HTLV-1 positive serology (unpublished data), possibly explained by the direct and strong immunosuppression exerted by HIV infection and, the usual mode of virus transmission (Mühle *et al.*, 2016; Montano *et al.*, 2005). Carriers, of HTLV-1/2 and of many HPV genotypes, are largely distributed in different geographical locations and among distinct ethnicities, besides the host-genetic background that somehow influences virus type as happens among Amerindian populations, more prone to express HTLV-2 viral genes (Nakauchi *et al.*, 1990; Nakauchi *et al.*, 1992; Kanzaki *et al.*, 1997). In the Central region of Brazil, screening head, neck and breast cancer patients, we detected HPV type 70 (low-risk type) by the Real Time PCR (Table 1), in a patient with oral cancer (head and neck cancer group), that previously yielded negative result by the hybrid capture assay (Bretas and Kanzaki, 2018; Leitão *et al.*, 2015). As stated before, findings of any viral gene sequence, claimed to be enrolled in the cancer etiology, is not a definitive proof, as evidences reported by Ciccarese *et al.* (2021) of HPV positivity of 51% and 43% among subjects with genital lesions and apparently healthy subjects, respectively; Sontakke *et al.* (2019) found 44.23 % and 5.76 % positivity for HPV type 16 and HPV type 18, respectively, in a group of asymptomatic women with normal cervix,

type 18 positivity, respectively, and 62.5 % and 22.5% HPV type 16 and 18 positivity, respectively, in a group of women with cervical malignancy. As observed, HPV positivity ratio is elevated among cancer patients, anyway healthy subjects harbor HPV genome. Some authors argue that the host competent immune system clears the virus genome, and the failure to achieve it, conducts to HPV viral persistence, probabilistically linked to virus infection, progressing to malignization, not solely by the virus action but synergistically with other factors as host-derived and environmental, as previously discussed. It could not also be discarded the possibility that HPV composes the normal human virome; a diversified number of host genetic and environmental factors play a fundamental role in the host homeostasis such as that any dysregulation in the fine balance of these factors could trigger pathological phenomena culminating in cancer.

Various studies credit the role of E6 and E7 oncoproteins of high-risk HPV types in the steady state progression of malignization of uterine cervix by counteracting the host pro-apoptotic tumor suppressor p53 and pRb, addressing these anti-tumor proteins to ubiquitination and proteasome degradation (Ci *et al.*, 2020; Chávez-López *et al.*, 2020; Javanmard *et al.*, 2019; Li *et al.*, 2019). Neutralization of the host cell anti-tumor proteins is not exclusively exerted by HPV oncoproteins, but by other viral and non-viral factors. Also, depending on the HPV genera and type, there are conflicting data suggesting the protective role of common skin existing HPV genus and types, including protection against UV induced cancer or even as an adjuvant factor in the oncogenic mechanism (Tahseen *et al.*, 2021). Most of the patients, mainly those diagnosed with oral squamous cell carcinoma, in the group of head and neck cancer, attending hospitals in the Central region of Brazil, displayed amplified bands corresponding to the *gag* gene by PCR of HERV-K nucleotide sequences (Table 1). Previously, these samples had negative results for low-risk and high-risk HPV and MMTV gene sequences by the hybrid capture assays and PCR, respectively (Kanzaki *et al.*, 2017). Concerning our preliminary data on HCV epidemiology (Table 1) (unpublished data), healthy subjects of a small village, in the Marajó island in northern Brazil, had low prevalence, of 1.04 % (3/288), comparing to a high prevalence in a group of HIV/AIDS patients in the Central region of Brazil, of 9,6% (15/156). Interesting to note the relatively high HTLV-1 prevalence in this small Marajó village, of 2.8 %(8/287), even though we could not detect dually HTLV/HCV infected subjects (Mata *et al.*, 2018).

## CONCLUSION

Our data contributes to the epidemiology of these putative oncogenic viruses, particularly in populations of diverse ethnicity in Brazil as in the Central region of Brazil inhabited by people of all regions of the country, as also in northern Brazil, specially mixed ethnicities' descendants of Amerinds, black Africans and mainly European Portuguese. Further assays and analysis in enlarged number of samples, including representants of other ethnicities and regions of the country, will be necessary to offer more robust scientific information.

## Acknowledgments

Research works and undergraduate students were supported with funds from FAP/DF, Brazil. We are grateful to the administrative and medical staff of hospitals in the Central and Northern region of Brazil. This work is specially dedicated to Dr. Koshi Maruyama, former Chiba Cancer Research Center oncovirus pathologist, in Japan.

**Disclosure statements:** None of the authors have conflict of interests.

## REFERENCES

- Akram N, Imran M, Noreen M, Ahmed F, Atif M, Fatima Z, Bilal Waqar A. Oncogenic Role of Tumor Viruses in Humans. *Viral Immunol.* 2017; 30(1):20-27. doi: 10.1089/vim.2016.0109.
- Al Dossary R, Alkharsah KR, Kussaibi H. Prevalence of Mouse Mammary Tumor Virus (MMTV)-like sequences in human breast cancer tissues and adjacent normal breast tissues in Saudi Arabia. *BMC Cancer.* 2018;18(1):170. doi: 10.1186/s12885-018-4074-6.
- Al-Eitan LN, Tarkhan AH, Alghamdi MA, Al-Qarqaz FA, Al-Kofahi HS. Transcriptome analysis of HPV-induced warts and healthy skin in humans. *BMC Med Genomics.* 2020;13(1):35. doi: 10.1186/s12920-020-0700-7.
- Ameur LB, Marie P, Thenoz M, Giraud G, Combe E, Claude JB, et al. Intragenic recruitment of NF- $\kappa$ B drives splicing modifications upon activation by the oncogene Tax of HTLV-1. *Nat Commun.* 2020;11(1):3045. doi: 10.1038/s41467-020-16853-x.
- Anticoli S, Amatore D, Matarrese P, De Angelis M, Palamara AT, Nencioni L, Ruggieri A. Counteraction of HCV-Induced Oxidative Stress Concur to Establish Chronic Infection in Liver Cell Cultures. *Oxid Med Cell Longev.* 2019; 2019:6452390. doi: 10.1155/2019/6452390. Erratum in: *Oxid Med Cell Longev.* 2019;2019:3712969.
- Ata EB, Allam AM, Elbayoumy MK, Mahmoud MAE. Electron microscopy and phylogenetic analysis of Bovine papillomavirus infection in cattle from four Egyptian governorates. *Trop Anim Health Prod.* 2021;53(1):160. doi: 10.1007/s11250-021-02607-4.
- Barros Kanzaki LI, Casseb J. Unusual finding of HTLV-I infection among Amazonian Amerindians. *Arch Med Res.* 2007;38(8):897-900. doi: 10.1016/j.arcmed.2007.05.002.
- Barros Kanzaki LI. Hypothetical HTLV-I induction by ionizing radiation. *Med Hypotheses.* 2006;67(1):177-82. doi: 10.1016/j.mehy.2006.01.017.
- Ben-David U, Beroukhim R, Golub TR. Genomic evolution of cancer models: perils and opportunities. *Nat Rev Cancer.* 2019;19(2):97-109. doi: 10.1038/s41568-018-0095-3.
- Bittner JJ. SOME POSSIBLE EFFECTS OF NURSING ON THE MAMMARY GLAND TUMOR INCIDENCE IN MICE. *Science.* 1936 Aug 14;84(2172):162. doi: 10.1126/science.84.2172.162.
- Bretas IS, Kanzaki LIB. Pesquisa de Sequências gênicas do Vírus do Papiloma Humano em Pacientes com Câncer de Cabeça e Pescoço no Hospital de Base do Distrito Federal. [Screening of Human Papillomavirus Gene Sequences in Head and Neck Cancer Patients in the Base Hospital of Federal District]. Poster presented at 24<sup>o</sup> Congresso de Iniciação Científica da UnB e 15<sup>o</sup> do DF [24<sup>th</sup> Congress of Scientific Initiation at the University of Brasilia and 15<sup>th</sup> of Federal District]; 2018 November 30; Brasilia, DF/Brazil.
- Buck CB, Day PM, Trus BL. The papillomavirus major capsid protein L1. *Virology.* 2013;445(1-2):169-74. doi: 10.1016/j.virol.2013.05.038.
- Cao H, Chen X, Wang Z, Wang L, Xia Q, Zhang W. The role of MDM2-p53 axis dysfunction in the hepatocellular carcinoma transformation. *Cell Death Discov.* 2020;6:53. doi: 10.1038/s41420-020-0287-y.
- Chávez-López MG, Zúñiga-García V, Castro-Magdonel BE, Vera E, Garrido E, Sánchez-Ramos J, et al. Eagl Gene and Protein Expression in Human Retinoblastoma Tumors and its Regulation by pRb in HeLa Cells. *Genes (Basel).* 2020;11(2):119. doi: 10.3390/genes11020119.
- Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M. Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science.* 1989;244(4902):359-62. doi: 10.1126/science.2523562.
- Ci X, Zhao Y, Tang W, Tu Q, Jiang P, Xue X, et al. HPV16 E7-impaired keratinocyte differentiation leads to tumorigenesis via cell cycle/pRb/invulocrin/spectrin/adducin cascade. *Appl Microbiol Biotechnol.* 2020;104(10):4417-4433. doi: 10.1007/s00253-020-10492-4.
- Ciccarese G, Herzum A, Pastorino A, Dezzana M, Casazza S, Mavilia MG, et al. Prevalence of genital HPV infection in STI and healthy populations and risk factors for viral persistence. *Eur J Clin Microbiol Infect Dis.* 2021;40(4):885-888. doi: 10.1007/s10096-020-04073-6.
- Cook LBM, Rowan AG. CD28 fusions: an opportunity for young ATL? *Blood.* 2020;135(17):1415-1416. doi: 10.1182/blood.2020004958.
- Dahiya R, Naqvi AAT, Mohammad T, Alajmi MF, Rehman MT, Hussain A, Hassan MI. Investigating the structural features of chromodomain proteins in the human genome and predictive impacts of their mutations in cancers. *Int J Biol Macromol.* 2019;131:1101-1116. doi: 10.1016/j.ijbiomac.2019.03.162.
- Davis K, Roden LC, Leaner VD, van der Watt PJ. The tumour suppressing role of the circadian clock. *IUBMB Life.* 2019;71(7):771-780. doi: 10.1002/iub.2005.
- Day PM, Weisberg AS, Thompson CD, Hughes MM, Pang YY, Lowy DR, Schiller JT. Human Papillomavirus 16 Capsids Mediate Nuclear Entry during Infection. *J Virol.* 2019;93(15):e00454-19. doi: 10.1128/JVI.00454-19.
- de Sanjosé S, Brotons M, Pavón MA. The natural history of human papillomavirus infection. *Best Pract Res Clin Obstet Gynaecol.* 2018;47:2-13. doi: 10.1016/j.bpobgyn.2017.08.015.
- Ding L, Liu C, Zhou Q, Feng M, Wang J. Association of estradiol and HPV/HPV16 infection with the occurrence of cervical squamous cell carcinoma. *Oncol Lett.* 2019;17(3):3548-3554. doi: 10.3892/ol.2019.10005.
- Du G, Zhang W, Zhang Z, Zeng M, Wang Y. HTLV-1-associated genes as potential biomarkers for endometrial cancer. *Oncol Lett.* 2019;18(1):699-705. doi: 10.3892/ol.2019.10389.
- Dudley JP, Golovkina TV, Ross SR. Lessons Learned from Mouse Mammary Tumor Virus in Animal Models. *ILAR J.* 2016;57(1):12-23. doi: 10.1093/ilar/ilv044.
- Durzynska J, Lesniewicz K, Poreba E. Human papillomaviruses in epigenetic regulations. *Mutat Res Rev Mutat Res.* 2017;772:36-50. doi: 10.1016/j.mrrev.2016.09.006.
- Ebrahim GJ. The five hepatitis viruses. *J Trop Pediatr.* 2011;57(6):401-4. doi: 10.1093/tropej/fmr099.
- Egawa N, Doorbar J. The low-risk papillomaviruses. *Virus Res.* 2017;231:119-127. doi: 10.1016/j.virusres.2016.12.017.
- Enose-Akahata Y, Jacobson S. Immunovirological markers in HTLV-1-associated myelopathy/tropical spastic paraparesis (HAM/TSP). *Retrovirology.* 2019;16(1):35. doi: 10.1186/s12977-019-0499-5.
- Fahad Ullah M. Breast Cancer: Current Perspectives on the Disease Status. *Adv Exp Med Biol.* 2019;1152:51-64. doi: 10.1007/978-3-030-20301-6\_4.
- Gao Y, Yu XF, Chen T. Human endogenous retroviruses in cancer: Expression, regulation and function. *Oncol Lett.* 2021;21(2):121. doi: 10.3892/ol.2020.12382.
- Garcia-Montojo M, Doucet-O'Hare T, Henderson L, Nath A. Human endogenous retrovirus-K (HML-2): a comprehensive review. *Crit Rev Microbiol.* 2018;44(6):715-738. doi: 10.1080/1040841X.2018.1501345.
- Gheit T. Mucosal and Cutaneous Human Papillomavirus Infections and Cancer Biology. *Front Oncol.* 2019;9:355. doi: 10.3389/fonc.2019.00355.

- Gifford RJ, Blomberg J, Coffin JM, Fan H, Heidmann T, Mayer J, Stoye J, Tristem M, Johnson WE. Nomenclature for endogenous retrovirus (ERV) loci. *Retrovirology*. 2018;15(1):59. doi: 10.1186/s12977-018-0442-1.
- Góngora-Biachi RA, González-Martínez P, Castro-Sansores C, Bastarrachea-Ortiz J. Infección por virus linfotrópico de células T humanstipo I/II enpacientes con cáncercervicouterino de la península de Yucatán, México [Infection with HTLV virus type I-II in patients with cervico-uterine cancer in the Yucatan peninsula, Mexico]. *GinecolObstet Mex*. 1997;65:141-4.
- Grandi N, Tramontano E. HERV Envelope Proteins: Physiological Role and Pathogenic Potential in Cancer and Autoimmunity. *Front Microbiol*. 2018;9:462. doi: 10.3389/fmicb.2018.00462.
- Hosainzadegan H, Khalilov R, Gholizadeh P. The necessity to revise Koch's postulates and its application to infectious and non-infectious diseases: a mini-review. *Eur J Clin Microbiol Infect Dis*. 2020;39(2):215-218. doi: 10.1007/s10096-019-03681-1.
- Houghton M. Discovery of the hepatitis C virus. *Liver Int*. 2009;29 Suppl 1:82-8. doi: 10.1111/j.1478-3231.2008.01925.x.
- Hron T, Elleder D, Gifford RJ. Deltaretroviruses have circulated since at least the Paleogene and infected a broad range of mammalian species. *Retrovirology*. 2019;16(1):33. doi: 10.1186/s12977-019-0495-9.
- Iwanaga M. Epidemiology of HTLV-1 Infection and ATL in Japan: An Update. *Front Microbiol*. 2020;11:1124. doi: 10.3389/fmicb.2020.01124.
- Javanmard D, Moein M, Esghaei M, Naseripour M, Monavari SH, Bokharaei-Salim F, et al. Molecular evidence of human papillomaviruses in the retinoblastoma tumor. *Virusdisease*. 2019;30(3):360-366. doi: 10.1007/s13337-019-00540-7.
- Kanzaki L, Maruyama K, Fukushima T, Tamezguerra R, Trejoavila L, Casseb J, et al. Markers of human T lymphotropic virus type I in patients with cancer of uterine cervix in Amazon, Brazil. *Int J Oncol*. 1997;10(5):1021-4. doi: 10.3892/ijo.10.5.1021.
- Kanzaki L, Maruyama K, Mochizuki S, Miyauchi M, Koshikawa N, Kawamura K. Comparison of DNA sequences of the long terminal repeat of human T lymphotropic virus type I in Japanese and Brazilian Amazonian samples. *Oncol Rep*. 1997;4(6):1187-8. doi: 10.3892/or.4.6.1187.
- Kanzaki LIB, Mello LS, Sampaio TL, Coimbra MVV, Da Mata ECG, Cavalcanti CC, et al. Detection of HERVs sequence in head and neck cancer patients in a public hospital (HBDF) in Brasilia, DF, Brazil. *World Conference on STDs, STIs & HIV/AIDS*. 2017 July 26-27; Vancouver, Canada.
- Kanzaki LIB. HTLV-1: A real pathogen or a runaway guest of a diseased cell? *J Biosci*. 2018;43(4):785-795.
- Katsura Y, Asai S. Evolutionary Medicine of Retroviruses in the Human Genome. *Am J Med Sci*. 2019;358(6):384-388. doi: 10.1016/j.amjms.2019.09.007.
- Khatun M, Ray R, Ray RB. Hepatitis C virus associated hepatocellular carcinoma. *Adv Cancer Res*. 2021;149:103-142. doi: 10.1016/bs.acr.2020.10.003.
- Kristensen MK, Christensen T. Regulation of the expression of human endogenous retroviruses: elements in fetal development and a possible role in the development of cancer and neurological diseases. *APMIS*. 2021;129(5):241-253. doi: 10.1111/apm.13130.
- Lawson JS, Glenn WK. Catching viral breast cancer. *Infect Agent Cancer*. 2021;16(1):37. doi: 10.1186/s13027-021-00366-3.
- LeBreton M, Switzer WM, Djoko CF, Gillis A, Jia H, Sturgeon MM, Shankar A, Zheng H, Nkeunen G, Tamoufe U, Nana A, Le DouxDiffo J, Tafon B, Kiyang J, Schneider BS, Burke DS, Wolfe ND. A gorilla reservoir for human T-lymphotropic virus type 4. *Emerg Microbes Infect*. 2014;3(1):e7. doi: 10.1038/emi.2014.7.
- Lehrer S, Rheinstein PH. The virology of breast cancer: viruses as the potential causative agents of breast tumorigenesis. *Discov Med*. 2019;27(148):163-166. Leitão EECV, Filho JE, da Mata ECG, Nakamura EK, Cavassani MOP, Silva ICR, et al. Screening of Papillomavirus Gene Sequences in Oral Cancer Patients in the Federal District, Brazil. *Oral Maxillofac Pathol J* 2015;6(1):537-543.
- Lessi F, Grandi N, Mazzanti CM, Civita P, Scatena C, Aretini P, et al. A human MMTV-like betaretrovirus linked to breast cancer has been present in humans at least since the copper age. *Aging (Albany NY)*. 2020;12(16):15978-15994. doi: 10.18632/aging.103780.
- Lewandowska AM, Rudzki M, Rudzki S, Lewandowski T, Laskowska B. Environmental risk factors for cancer - review paper. *Ann Agric Environ Med*. 2019;26(1):1-7. doi: 10.26444/aaem/94299.
- Li S, Hong X, Wei Z, Xie M, Li W, Liu G, et al. Ubiquitination of the HPV Oncoprotein E6 Is Critical for E6/E6AP-Mediated p53 Degradation. *Front Microbiol*. 2019;10:2483. doi: 10.3389/fmicb.2019.02483.
- Li Y, Hively WP, Varmus HE. Use of MMTV-Wnt-1 transgenic mice for studying the genetic basis of breast cancer. *Oncogene*. 2000;19(8):1002-9. doi: 10.1038/sj.onc.1203273.
- Ljubojevic S, Skerlev M. HPV-associated diseases. *Clin Dermatol*. 2014;32(2):227-34. doi: 10.1016/j.clindermatol.2013.08.007.
- Mansour M N J, Kanzaki LIB. Inquérito Soromolecular para Virus Humano T-linfotrópico entre pacientes com diferentes tipos de neoplasias [Human T-cell Lymphotropic virus Serological and Molecular Screening among patients with different types of neoplasia]. Poster presented at 24<sup>o</sup> Congresso de Iniciação Científica da UnB e 15<sup>o</sup> do DF [24<sup>th</sup> Congress of Scientific Initiation at the University of Brasilia and 15<sup>th</sup> of Federal District]; 2018 November 30; Brasilia, DF/Brazil.
- Mata EC, Bezerra RM, Proietti Júnior AA, Pamplona LK, Gomes LO, Corrêa VC, et al. HTLV-1/2 prevalence in two Amazonian communities. *J Virus Erad*. 2018;4(3):174-178.
- Matsuoka M, Mesnard JM. HTLV-1 bZIP factor: the key viral gene for pathogenesis. *Retrovirology*. 2020;17(1):2. doi: 10.1186/s12977-020-0511-0.
- McBride AA. Mechanisms and strategies of papillomavirus replication. *Biological Chemistry*. 2017; 398(8):919-927. doi:10.1515/hsz-2017-0113.
- Moles R, Sarkis S, Galli V, Omsland M, Purcell DFJ, Yurick D, Khoury G, Pise-Masison CA, Franchini G. p30 protein: a critical regulator of HTLV-1 viral latency and host immunity. *Retrovirology*. 2019;16(1):42. doi: 10.1186/s12977-019-0501-2.
- Montano SM, Sanchez JL, Laguna-Torres A, Cuchi P, Avila MM, Weissenbacher M, et al; South American HIV Molecular Surveillance Working Group. Prevalences, genotypes, and risk factors for HIV transmission in South America. *J Acquir Immune Defic Syndr*. 2005;40(1):57-64. doi: 10.1097/01.qai.0000159667.72584.8b.
- Mühle M, Kroniger T, Hoffmann K, Denner J. The immunosuppressive domain of the transmembrane envelope protein gp41 of HIV-1 binds to human monocytes and B cells. *Immunol Res*. 2016;64(3):721-9. doi: 10.1007/s12026-015-8776-4.
- Nakauchi CM, Linhares AC, Maruyama K, Kanzaki LI, Macedo JE, Azevedo VN, Casseb JS. Prevalence of human T cell leukemia virus-I (HTLV-I) antibody among populations living in the Amazon region of Brazil (preliminary report). *Mem Inst Oswaldo Cruz*. 1990;85(1):29-33. doi: 10.1590/s0074-0276199000100004.
- Nakauchi CM, Maruyama K, Kanzaki LI, Linhares AC, Azevedo VN, Fukushima T, et al. Prevalence of HTLV-I antibody among two distinct ethnic groups inhabiting the Amazon region of Brazil. *Rev Inst Med Trop Sao Paulo*. 1992;34(4):323-8. doi: 10.1590/s0036-46651992000400009.
- Narat V, Kampo M, Heyer T, Rupp S, Ambata P, Njouom R, Giles-Vernick T. Using physical contact heterogeneity and frequency to characterize dynamics of human exposure to nonhuman primate bodily fluids in central Africa. *PLoS Negl Trop Dis*. 2018;12(12):e0006976. doi: 10.1371/journal.pntd.0006976.
- Nartey T, Mazzanti CM, Melana S, Glenn WK, Bevilacqua G, Holland JF, Whitaker NJ, Lawson JS, Pogo BGT. Erratum to: Mouse mammary tumor-like virus (MMTV) is present in human breast tissue before development of virally associated breast cancer. *Infect Agent Cancer*. 2017;12:16. doi:

- 10.1186/s13027-017-0126-9. Erratum for: *Infect Agent Cancer*. 2017;12:1.
- Nartey T, Moran H, Marin T, Arcaro KF, Anderton DL, Etkind P, Holland JF, Melana SM, Pogo BG. Human Mammary Tumor Virus (HMTV) sequences in human milk. *Infect Agent Cancer*. 2014 Jun 17;9:20. doi: 10.1186/1750-9378-9-20.
- Navas MC, Glaser S, Dhruv H, Celinski S, Alpini G, Meng F. Hepatitis C Virus Infection and Cholangiocarcinoma: An Insight into Epidemiologic Evidences and Hypothetical Mechanisms of Oncogenesis. *Am J Pathol*. 2019;189(6):1122-1132. doi: 10.1016/j.ajpath.2019.01.018.
- Noon MC, Wolford RG, Parks WP. Expression of mouse mammary tumor viral polypeptides in milks and tissues. *J Immunol*. 1975;115(3):653-8.
- Perzova R, Abbott L, Benz P, Landas S, Khan S, Glaser J, Cunningham CK, Poesz B. Is MMTV associated with human breast cancer? Maybe, but probably not. *Virology*. 2017;14(1):196. doi: 10.1186/s12985-017-0862-x.
- Piver E, Boyer A, Gaillard J, Bull A, Beaumont E, Roingard P, Meunier JC. Ultrastructural organisation of HCV from the bloodstream of infected patients revealed by electron microscopy after specific immunocapture. *Gut*. 2017;66(8):1487-1495. doi: 10.1136/gutjnl-2016-311726.
- Prati B, Marangoni B, Boccardo E. Human papillomavirus and genome instability: from productive infection to cancer. *Clinics (Sao Paulo)*. 2018;73(suppl 1):e539s. doi: 10.6061/clinics/2018/e539s.
- Ramírez-López IG, Ramírez de Arellano A, Jave-Suárez LF, Hernández-Silva CD, García-Chagollan M, Hernández-Bello J, et al. Interaction between 17 $\beta$ -estradiol, prolactin and human papillomavirus induce E6/E7 transcript and modulate the expression and localization of hormonal receptors. *Cancer Cell Int*. 2019;19:227. doi: 10.1186/s12935-019-0935-6.
- Rector A, Van Ranst M. Animal papillomaviruses. *Virology*. 2013;445(1-2):213-23. doi: 10.1016/j.virol.2013.05.007.
- Rocamonde B, Carcone A, Mahieux R, Dutartre H. HTLV-1 infection of myeloid cells: from transmission to immune alterations. *Retrovirology*. 2019;16(1):45. doi: 10.1186/s12977-019-0506-x.
- Sagan SM, Chahal J, Sarnow P. cis-Acting RNA elements in the hepatitis C virus RNA genome. *Virus Res*. 2015;206:90-8. doi: 10.1016/j.virusres.2014.12.029.
- Sontakke BR, Ambulkar PS, Talhar S, Shivkumar PV, Bharambe MS, Pal A. Molecular Genetic Study to Detect Prevalence of High-risk Human Papilloma Virus Strains (type 16 and 18) in Cervical Lesions and Asymptomatic Healthy Subjects of Rural Central India. *J Cytol*. 2019;36(1):32-37. doi: 10.4103/JOC.JOC\_10\_18.
- Spearman CW, Dusheiko GM, Hellard M, Sonderup M. Hepatitis C. *Lancet*. 2019;394(10207):1451-1466. doi: 10.1016/S0140-6736(19)32320-7.
- Spurgeon ME, Uberoi A, McGregor SM, Wei T, Ward-Shaw E, Lambert PF. A Novel *In Vivo* Infection Model To Study Papillomavirus-Mediated Disease of the Female Reproductive Tract. *mBio*. 2019;10(2):e00180-19. doi: 10.1128/mBio.00180-19.
- Strickler HD, Rattray C, Escoffery C, Manns A, Schiffman MH, Brown C, et al. Human T-cell lymphotropic virus type I and severe neoplasia of the cervix in Jamaica. *Int J Cancer*. 1995 Mar 29;61(1):23-6. doi: 10.1002/ijc.2910610105.
- Taguchi H, Daibata M, Kitagawa T, Kubonishi I, Asai M, Sagara Y, et al. Generalized lymph node metastasis of early uterine cancer in an HTLV-I carrier. *Cancer*. 1988;62(12):2614-7. doi: 10.1002/1097-0142(19881215)62:12<2614::aid-cncr2820621227>3.0.co;2-x.
- Tahseen D, Rady PL, Tyring SK. Effects of  $\beta$ -HPV on DNA damage response pathways to drive carcinogenesis: a review. *Virus Genes*. 2021;57(1):23-30. doi: 10.1007/s11262-020-01813-w.
- Tanaka M, Kawazu Y, Yoshida T, Konishi T, Takenouchi N, Miwa M. Effects of radiation based on whole-body irradiation in HTLV-1-infected mice. *J Radiat Res*. 2019;60(5):705-708. doi: 10.1093/jrr/trz050. Erratum in: *J Radiat Res*. 2020;61(5):818.
- Toyoda K, Yasunaga JI, Matsuoka M. [Astute strategies of HTLV-1 with driven viral genes]. *Uirusu*. 2019;69(1):37-46. Japanese. doi: 10.2222/jsv.69.37.
- Us D. Viral süperantijenler [Viral superantigens]. *Mikrobiyol Bul*. 2016;50(3):491-504. Turkish. doi: 10.5578/mb.24250.
- Virzi A, Roca Suarez AA, Baumert TF, Lupberger J. Oncogenic Signaling Induced by HCV Infection. *Viruses*. 2018;10(10):538. doi: 10.3390/v10100538.
- Vonsky M, Shabaeva M, Runov A, Lebedeva N, Chowdhury S, Palefsky JM, Isagulians M. Carcinogenesis Associated with Human Papillomavirus Infection. Mechanisms and Potential for Immunotherapy. *Biochemistry (Mosc)*. 2019;84(7):782-799. doi: 10.1134/S0006297919070095.
- Williams LM, Gilmore TD. Looking Down on NF- $\kappa$ B. *Mol Cell Biol*. 2020 Jul 14;40(15):e00104-20. doi: 10.1128/MCB.00104-20.
- Wong RWJ, Tan TK, Amanda S, Ngoc PCT, Leong WZ, Tan SH, Asamitsu K, Hibi Y, Ueda R, Okamoto T, Ishida T, Iida S, Sanda T. Feed-forward regulatory loop driven by IRF4 and NF- $\kappa$ B in adult T-cell leukemia/lymphoma. *Blood*. 2020;135(12):934-947. doi: 10.1182/blood.2019002639.
- Wong RWJ, Tan TK, Amanda S, Ngoc PCT, Leong WZ, Tan SH, Asamitsu K, Hibi Y, Ueda R, Okamoto T, Ishida T, Iida S, Sanda T. Feed-forward regulatory loop driven by IRF4 and NF- $\kappa$ B in adult T-cell leukemia/lymphoma. *Blood*. 2020;135(12):934-947. doi: 10.1182/blood.2019002639.
- Wu S, Zhu W, Thompson P, Hannun YA. Evaluating intrinsic and non-intrinsic cancer risk factors. *Nat Commun*. 2018;9(1):3490. doi: 10.1038/s41467-018-05467-z.
- Zhi H, Guo X, Ho YK, Pasupala N, Engstrom HAA, Semmes OJ, Giam CZ. RNF8 Dysregulation and Down-regulation During HTLV-1 Infection Promote Genomic Instability in Adult T-Cell Leukemia. *PLoS Pathog*. 2020;16(5):e1008618. doi: 10.1371/journal.ppat.1008618.

\*\*\*\*\*