



UNIVERSIDADE DE BRASÍLIA  
INSTITUTO DE CIÊNCIAS BIOLÓGICAS  
PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA

**A LUZ ARTIFICIAL A NOITE EM PARQUES URBANOS AFETA  
OS MORCEGOS OU AS PRESAS DE MORCEGOS?**

LUCAS DAMÁSIO EVANGELISTA REIS

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## Resumo

Com a crescente urbanização no mundo é essencial saber como as espécies animais respondem aos impactos desse processo. A iluminação artificial noturna altera ambientes que eram naturalmente escuros a noite para ambientes iluminados, impactando a biodiversidade noturna. Como morcegos são ativos durante a noite a iluminação pode interferir no padrão de atividade e estruturação da comunidade de morcegos em ambientes urbanos. Assim, vou avaliar se a iluminação artificial afeta a atividade dos morcegos, e se esta está relacionada a abundância de presas de morcegos nos parques urbanos. Para isso, testei as hipóteses da atividade de morcegos ser maior próximo aos locais com iluminação artificial, assumindo que a abundância de insetos será maior por serem atraídos pela luz; espécies de voo rápido serão mais ativas próximo à iluminação artificial, e as de voo lento vão apresentar resposta negativa à iluminação artificial; e a última, que as espécies de voo rápido estarão positivamente relacionadas com os artrópodes, pois são mais ativas próximo aos locais com iluminação artificial onde há maior abundância de artrópodes. Verifiquei a atividade dos morcegos por meio de gravações acústicas, e a abundância de artrópodes com armadilhas de cartolina amarela adesiva instaladas em cada um dos locais de amostragem. Foi verificado o efeito direto e indireto da luz artificial sobre a atividade dos morcegos, via abundância de artrópodes, utilizando modelos de equações estruturais. A abundância de artrópodes foi afetada positivamente pela iluminação artificial, e a atividade de morcegos não. No entanto, a atividade dos morcegos foi afetada positivamente pela abundância de artrópodes. A atividade das espécies de voo rápido aumentou em resposta à luz artificial e à abundância de artrópodes. O grupo *Eumops sp/Nyctinomops macrotis* mostrou uma resposta negativa a abundância de artrópodes. A atividade de *Eptesicus sp.*, espécie de voo lento, teve resposta negativa à luz artificial. A iluminação artificial influencia diretamente a abundância de artrópodes, mas afeta indiretamente a atividade dos morcegos. Este efeito parece ser espécie específico: espécies de voo rápido se beneficiam com a presença da iluminação artificial.

**Palavras-chave:** artrópodes, ecolocalização de morcegos, ecossistemas urbanos, insetivoria, remanescentes verdes.

## **Introdução Geral**

Urbanização é o resultado de um processo de extrema alteração no uso da terra (Shochat et al. 2006; Grimm et al. 2008). A taxa de mudança na qual as áreas urbanas estão evoluindo é devido ao crescimento natural da população humana, incluindo a migração significativa das áreas rurais para urbanas e a expansão espacial das cidades (Grimm et al. 2008; Montgomery 2008; UN 2012; McDonnell e Hahs 2013). Além disso, projeta-se que mais da metade do crescimento da população urbana ocorra em cidades menores (UN 2012). A urbanização não é um evento concentrado localmente, mas sim um processo disperso e um acontecimento mundial (McDonald 2008). Países desenvolvidos já passaram por amplos processos de urbanização e esse crescimento acontece de forma lenta, diferente do que ocorre em países em desenvolvimento como na Ásia, África e América Latina onde há um grande aumento da urbanização, inclusive em muitos *hotspots* de biodiversidade (Myers et al. 2000).

A urbanização é responsável por um grande declínio da biodiversidade global (Sala et al. 2000). Os efeitos do processo da urbanização operam a partir de escalas locais (por meio da expansão das cidades) e também em escalas globais (por meio da emissão de gases de efeito estufa) (McDonald et al. 2008), agindo diretamente na expansão das áreas urbanas, quanto indiretamente, no crescimento da infraestrutura e mudanças no consumo e poluição (McIntyre et al. 2000; Pickett et al. 2001). Além da remoção da área natural, a expansão das cidades também promove impactos nos ambientes rurais e naturais devido ao aumento da fragmentação e dos efeitos de borda, provocando aumento dos níveis de temperatura e ruído, introduzindo novos estressores antropogênicos em ecossistemas marginais (Grimm et al. 2008) e áreas protegidas próximas (McDonald et al. 2008; McDonald e Marcotullio 2013). No entanto, apesar da transformação radical da terra e da perda de habitat causada pela urbanização, muitas espécies ainda podem



persistir em ambientes urbanos e algumas até mesmo experimentar aumentos populacionais (McKinney 2006; Russo e Ancillotto 2015).

A resposta das espécies à urbanização varia de acordo com suas características funcionais. Algumas espécies são tolerantes ao novo ambiente, enquanto outras podem ser beneficiadas e outras espécies são prejudicadas (Russo e Ancillotto 2015). Assim, as pressões geradas pela urbanização atuam como um filtro ambiental, selecionando as espécies a partir das suas características funcionais, permitindo que estas sobrevivam nesses novos habitats (Aronson et al. 2016). As espécies que requerem um habitat mais conservado são geralmente as mais prejudicadas pela urbanização. Geralmente as espécies mais raras e menos tolerantes desaparecem ou mudam as relações com o meio para se manterem nesses ambientes (Kurta e Teramino 1992; Hourigan et al. 2006; Luck et al. 2013).

Além da perda de habitat, fragmentação e efeito de borda gerados no processo de urbanização (Grimm et al. 2008), outros estressores antropogênicos são introduzidos nesse processo. A poluição sonora, gerada por fontes de sons antropogênicos, por exemplo, diminui a riqueza de espécies de aves em ambientes com alta perturbação sonora. Entretanto, facilita o sucesso reprodutivo de indivíduos que nidificam em áreas urbanas como resultado da perturbação da interação predador-presa (Francis et al. 2009). Outro exemplo do impacto dos sons antropogênicos é evidenciado com a espécie de morcego *Myotis myotis* que apresentou um baixo sucesso de captura de presas quando exposto ao som do tráfego de veículos (Schaub et al 2008).

Além do som, outro impacto gerado pela urbanização é a poluição por agentes químicos. A biota aquática é uma das mais impactadas pelos poluentes químicos, sendo observados efeitos negativos na comunidade de peixes e invertebrados aquáticos, que tem a riqueza e abundância diminuídas nos ambientes aquáticos (Kalogianni et al. 2017; Karaouzas et al. 2017). Diversos

trabalhos mostram que anfíbios são fortemente afetados por poluentes químicos na água, gerando uma baixa sobrevivência de girinos quando expostos a diferentes compostos químicos e quando estes sobrevivem, apresentam baixa mobilidade e problemas de má formação de algumas estruturas corporais (Carey e Bryant 1995; Rowe et al 1998; Rubbo e Kiesecker 2005; Relyea e Diecks 2008).

Entre esses impactos a poluição luminosa vem ganhando destaque (Longcore e Rich 2004; Gaston et al. 2013). Poluição luminosa é definida pela alteração dos níveis de luminosidade na paisagem noturna provocada por fontes de iluminação artificial (Falchi et al. 2011; Kyba e Hölker 2013). A iluminação artificial tem aumentando em resposta à crescente urbanização, do crescimento das populações humanas, do desenvolvimento econômico e dos avanços nas tecnologias de iluminação que proporcionam benefícios econômicos, comerciais, recreacionais e de segurança (Hölker et al. 2010a). A iluminação artificial cresce em média 6% ao ano e a poluição luminosa é tratada por ecólogos como uma ameaça a biodiversidade (Hölker et al. 2010a, b), pois 30% dos vertebrados e mais de 60% dos invertebrados são noturnos (Hölker et al. 2010b). O crescente aumento da iluminação artificial no ambiente noturno afeta fortemente os padrões naturais desses animais. Um dos efeitos da poluição luminosa é a capacidade de atrair para a fonte de iluminação artificial alguns animais ou repelir outros. Consequentemente, afetando esses animais na procura de alimentos, reprodução e comunicação, e alterando a estrutura das comunidades noturnas (Longcore e Rich 2004). Assim, a compreensão de como a iluminação artificial afeta as comunidades em ambientes urbanos será de grande valor para a conservação de espécies em ambientes urbanos, e para que as novas cidades sejam planejadas já pensando na minimização de efeitos negativos na biodiversidade.

Diversos grupos animais são afetados pela iluminação artificial, principalmente os noturnos. Estudos recentes vêm demonstrando que aves migratórias noturnas diminuíram suas rotas de voo e selecionaram habitats menos conservados em resposta a iluminação artificial (Cabrera- Cruz et al. 2018; McLaren et al. 2018). O comportamento reprodutivo das aves também foi afetado. Machos que ocupavam territórios periféricos a iluminação artificial vocalizaram significativamente mais cedo e apresentaram um maior sucesso na obtenção de parceiros extrapares (Kempnaers et al.2010). Conseqüentemente também é observado um aumento no período de atividade das aves quando expostas a iluminação artificial (de Jong et al. 2017; Raap et al. 2017).

Em tartarugas marinhas também foi observada uma relação negativa entre a densidade de locais de desova a medida que havia aumento na intensidade de iluminação artificial (Weishampel et al. 2016; Hu et al. 2018). Anfíbios apresentaram baixa sobrevivência de girinos, baixa velocidade de movimentação e ficaram mais susceptíveis ao parasitismo quando expostos a iluminação artificial (May et al. 2019). Também foi observada mudança no comportamento de algumas espécies de sapos, quando expostos a iluminação artificial. Estes vocalizaram durante toda a noite com maior gasto de energia quando comparado a locais sem a iluminação artificial (Dias et al. 2019).

Em mamíferos o efeito da iluminação artificial tem sido observado em diferentes grupos. Marsupiais apresentaram uma redução nos níveis de melatonina e conseqüente um aumento no período da gestação (Robert et al. 2015). Em pequenos mamíferos não voadores foi observado um aumento nas distâncias percorridas durante a noite, e a redução na área de sobreposição dos indivíduos quando expostos a iluminação artificial (Hoffmann et al. 2018; 2019). Assim, esses trabalhos mostram que mesmo níveis muito baixos de poluição luminosa podem interferir no

comportamento e na fisiologia dos mamíferos noturnos. Essa constatação é preocupante pois das espécies de mamíferos noturnas, mais de 1300 espécies são de morcegos, representantes da ordem Chiroptera,(Fenton e Simmons 2014) e com a expansão das áreas iluminadas, a cobertura de habitats adequados e naturalmente escuros diminuem rapidamente (Hölker et al. 2010a; Stone et al. 2015; Falchi et al. 2016)

Em geral, a urbanização afeta negativamente a atividade e abundância dos morcegos (Jung e Threlfall 2016). Curiosamente os morcegos são uma das ordens de mamíferos com maior riqueza de espécies registradas em ambientes urbanos (Santini et al. 2019). Essa riqueza pode estar relacionada às estruturas urbanas, como edifícios e pontes, que se assemelham com os abrigos naturais dos morcegos (Ávila-Flores e Fenton 2005; Jung e Kalko 2010). Entretanto, por possuírem hábito noturno, morcegos se tornam passíveis de impacto com a expansão da iluminação artificial (Stone et al. 2015). O efeito da iluminação artificial nos morcegos interfere diretamente na eficiência de forrageio das espécies (Rowse et al. 2016). A iluminação artificial atrai uma grande abundância de artrópodes como Lepidopteras, Coleoptera e Dipteras geralmente predados morcegos (Eisenbeis 2006; van Langevelde et al. 2011) e os morcegos se aproveitam da luz para aumentar sua eficiência de forrageio (Rydell 1992, 2006). Essa relação já foi observada em locais próximos as fontes de iluminação artificial onde há maior atividade de morcegos e consequentemente são os locais onde há maior abundância de artrópodes (Ávila-Flores e Fenton 2005; Jung e Kalko 2010).

Em geral, artrópodes são fortemente afetados pela iluminação artificial com aumento do número de espécies atraídas para as fontes de iluminação artificial durante a noite (Eisenbeis 2006). A composição espectral da luz pode influenciar no grau de atratividade dos insetos (van Grunsven et al. 2014; Longcore et al. 2015), e variáveis como comprimento de onda, saturação de

cor e brilho da luz são características importante que influenciam a atração de insetos (Antignus 2000). van Langevelde et al. (2011) observaram que luzes com menores comprimentos de onda atraíram uma maior riqueza e abundâncias de espécies de mariposas. Estas luzes podem atrair em média mariposas com maior massa corporal, maior dimensão de asa e olhos maiores. Além do comprimento de onda, outras características influenciam a atratividade de insetos pela iluminação artificial, como a saturação da cor e brilho da luz (Antignus 2000). Os Dípteros, por exemplo, são mais atraídos por luz ultravioleta (UV), luz azul e verde (Burkett et al. 1998; Burkett e Butler 2005), enquanto Lepidópteros são atraídos pela luz UV e azul (Solano Lamphar e Kocifaj 2013; van Langevelde et al. 2011). Já Hemípteros são mais atraídos por luz azul e luzes brancas de baixa intensidade (Pacheco-Tucuch et al. 2012).

Consequentemente, a atividade das espécies de morcegos responde as características da luz artificial, como o tipo de lâmpada (Rowse et al. 2016). Frank et al (2019) observaram na Costa Rica, que a atividade dos morcegos foi maior próximo as lâmpadas fluorescentes de cor azul e branca. Entretanto, essa resposta variou entre as espécies de morcegos. *Pipistrellus pipistrellus* teve menor atividade próximo a lâmpadas de LED (Light Emitting Diode, em inglês) em relação às lâmpadas de vapor de mercúrio (MV, em inglês). O contrário foi encontrado para espécies do gênero *Myotis*, que apresentou maior atividade próximo as lâmpadas de LED (Lewanzik e Voigt 2017). Lâmpadas de LED emitem baixas quantidades de raio UV, atraindo menos insetos (Longcore et al. 2015) refletindo no resultado observado para *P. pipistrellus*, e diferente do resultado observado para o gênero *Myotis*, que normalmente são intolerantes a iluminação artificial. As lâmpadas de LED são menos prejudiciais do que as MV (Gorresen et al. 2015).

No geral, é observado que a atividade de morcegos insetívoros é maior próximo aos locais com iluminação artificial (Jung e Kalko 2010; Rowse et al. 2016; Frank et al. 2019). No entanto,

esse efeito da iluminação artificial afeta de forma diferente as espécies de morcegos insetívoros. Espécies de voo rápido, que possuem hábito de forrageio em áreas abertas, são mais tolerantes e são frequentemente registradas próximas à iluminação artificial (Rowse et al. 2016). Forrageadores de área aberta possuem asas longas e estreitas e necessitam de um voo rápido para se manterem no ar, assim, utilizam chamadas de ecolocalização de banda estreita de alta intensidade e baixa frequência para detectarem os insetos a longas distâncias (Norberg e Rayner 1987; Rydell 2006; Kalko et al. 2008). Morcegos de áreas abertas quando forrageam próximos aos locais iluminados, voam acima dos postes de iluminação artificial e no momento da captura dos insetos passam pela luz (Jung e Kalko 2010).

Entretanto espécies que possuem voo lento e que forrageiam em ambiente com mais obstáculos, como bordas de matas, em geral, apresentam intolerância a iluminação artificial e raramente são registrados próximos aos locais iluminados (Rowse et al. 2016). Os forrageadores de borda possuem asas largas e curtas que facilitam a manobrabilidade em ambientes com muitos obstáculos (Norberg and Rayner 1987) e por conta dessa morfologia, a exposição aos predadores é uma das prováveis explicações para essa intolerância (Jones e Rydell 1994; Rydel et al 1996). Além disso, a estratégia de captura de presas também pode estar relacionado a essa intolerância, pois essas espécies são adaptadas para caçar presas em ambientes com muitos obstáculos (Norberg e Rayner 1987) e como os postes de iluminação pública estão localizados em áreas semi-abertas/abertas, esses locais podem não ser propícios para o forrageio dos morcegos de voo lento. Com isso, a proliferação da iluminação artificial tem o potencial de alterar a estrutura da comunidade de morcegos insetívoros aéreos, selecionando as espécies que se beneficiam e restringindo as que evitam locais com iluminação artificial (Stone et al. 2015; Rowse et. al. 2016; Frank et al. 2019).

Grande parte do conhecimento acerca do impacto da iluminação artificial sobre os morcegos provém de trabalhos que foram realizados no hemisfério norte (Gorresen et al. 2015; Rowse et al. 2016; Lewanzik e Voigt 2017). Poucos trabalhos sobre o efeito da iluminação artificial nos morcegos foram realizados na região neotropical (Jung e Kalko 2010; 2011; Frank et al. 2019). Há, portanto, muito a se descobrir sobre como os morcegos neotropicais respondem a iluminação artificial. Em especial no Brasil onde há um aumento significativo da urbanização (Klink e Machado 2005; Strassburg et al. 2017). Em recente revisão foram registradas 87 espécies de morcegos em ambientes urbanos (Nunes et al. 2017) no Brasil, representando aproximadamente metade da ordem presente no país. Entretanto, grande parte dos trabalhos realizados são relacionados a saúde pública em relação as possíveis doenças que os morcegos são potenciais vetores (Nunes et al. 2017). Poucos são os trabalhos que abordam questões ecológicas dos morcegos em ambientes urbanos no Brasil, como feito por Silva de Araújo e Bernard (2016). Esses autores observaram que a atividade de morcegos insetívoros em remanescentes verdes de áreas urbanas é maior do que nos locais onde esses remanescentes não estão presentes.

Nesse contexto que o meu trabalho se insere. O estudo foi conduzido na cidade de Brasília, Distrito Federal do Brasil. Brasília é uma cidade nova, com 59 anos, com uma população de aproximadamente 3 milhões de habitantes (IBGE 2019), totalmente inserido no Bioma Cerrado. Está cercada por extensas áreas de agricultura. Brasília possui um clima com duas estações bem definidas: de outubro a março é período chuvoso, e de abril à setembro o período de seca. Brasília está localizada a 1.000 metros do nível do mar (GDF 2019). Brasília é uma cidade bosque, com uma grande cobertura arbórea. Uma das características de Brasília são os Parques Urbanos, frequentemente utilizados pela população para atividade de recreação. Esses parques possuem em

sua composição a vegetação nativa do bioma Cerrado (IBRAM 2013) e assim podem ser utilizados pela fauna local.

Meu objetivo é avaliar como a iluminação artificial afeta os morcegos insetívoros nos parques urbanos de Brasília, a capital de um país megadiverso sem estudos que avaliem como os morcegos respondem à presença de luz artificial em ambientes urbanos. Vou testar as seguintes hipóteses: a primeira é que a atividade dos morcegos insetívoros será maior próximo aos locais com iluminação artificial, pois a abundância de insetos também será maior próximos aos locais com iluminação artificial; a segunda, é que a atividade das espécies de voo rápido será maior próximos aos locais com iluminação artificial, enquanto as espécies de voo lento apresentarão uma resposta negativa à iluminação artificial, provavelmente para evitar a exposição a predadores em áreas abertas; e a terceira, as espécies de voo rápido estarão positivamente relacionadas com ordens de artrópodes, pois são mais ativas próximo aos locais com iluminação artificial onde há maior abundância de artrópodes.

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## Nocturnal artificial light in Urban Parks: Does it affect bats or bats' prey?

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Running title: Artificial light: Does it affect bats or bats' prey.

### Highlights

- Artificial lights lure insects, thus attracting bats, in a bottom-up effect system.
- The response is species specific, with fast-flying bat species being more tolerant to light.
- Fast flying bats are positively correlated with arthropod orders.

**Abstract**

Increasing global urbanization turns essential the understanding of the role of artificial lighting at night (ALAN) on bat's activity and community structuring in urban environments. We aimed to evaluate if artificial lighting influences the bat's activity and prey abundance in urban parks. We hypothesized that bat activity would be higher near sites with ALAN, supposing that the abundance of insects, attracted by the light, would be higher. Second, we expected that the activity of fast-flying species would be higher near ALAN while slow-flying species would present a negative response to ALAN, probably to avoid exposition to predators in more illuminated areas. Third, fast-flying species are positively related to arthropod orders, because both are positively related to ALAN source. We established bat activity through acoustic recordings. We measured arthropods abundance using yellow sticky traps installed in each sampling site. We checked the direct and indirect effect of artificial light on bat activity, via arthropod abundance using structural equation models. The abundance of arthropods was positively influenced by ALAN, while bat activity was not directly affected. However, bat activity changed positively with arthropod abundance. The activity of most fast-flying species increased in response to artificial light and arthropod abundance. *Eumops* sp/*Nyctinomops macrotis* group presented a negative response to arthropod abundance. The activity of slow-flying species *Eptesicus* sp. presented a negative response to artificial light. ALAN poses a direct influence on arthropod abundance, indirectly affecting bat activity, though this effect is species-specific, with fast-fly species benefiting with ALAN.

**Keywords:** arthropods, bat echolocation, green remnants, insectivory, urban ecosystems

## 1 **1. Introduction**

2           One of the leading causes of global biodiversity decreasing is urbanization (Sala et al.,  
3 2000) because it converts natural habitats into artificial environments composed of human  
4 constructions. This process initially produces habitat fragmentation (Strassburg et al., 2017) and  
5 later noise pollution (Bunkley, McClure, Kleist, Francis, & Barber, 2015), and artificial lighting  
6 (E. G. Rowse, Lewanzik, Stone, Harris, & Jones, 2016). Those processes act as environmental  
7 filter structuring communities in urban environments (Aronson et al., 2016) selecting species that  
8 present functional traits that enable them to survive in this new habitat. Some species, like free-  
9 tailed bats (Molossidae) and evening bats (Vespertilionidae), possibly are more tolerant or even  
10 favored by urbanization, according to their ecological requirements, occurring in both urban and  
11 suburban environments (Santini et al., 2019). Species that require conserved natural habitat are  
12 usually hampered by urbanization, with rare or less tolerant species disappearing, or changing  
13 their ecology (Avila-Flores & Fenton, 2005; Hourigan, Johnson, & Robson, 2006; Kurta &  
14 Teramino, 1992; Loeb, Post, & Hall, 2009; Luck, Smallbone, Threlfall, & Law, 2013).

15           Artificial lighting at night (ALAN) is one of the most detrimental stresses caused by  
16 urbanization. The rapid increase of ALAN is affecting a considerable portion of the global  
17 biodiversity because approximately 30% of vertebrates and over 60% of invertebrates are  
18 nocturnal (Hölker, Wolter, Perkin, & Tockner, 2010). Recently studies have shown that birds  
19 (Kempnaers, Borgstrom, Loes, Schlicht, & Valcu, 2010), turtles (Bourgeois, Gilot-Fromont,  
20 Viallefont, Boussamba, & Deem, 2009), amphibians (Wise, 2007) and insects (van Langevelde,  
21 Ettema, Donners, WallisDeVries, & Groenendijk, 2011), have their activity patterns affected by  
22 ALAN. Animals can be attracted or repelled by light pollution, and are affected in foraging,  
23 reproduction, and communication. Therefore, altering nocturnal communities' structures

24 (Longcore & Rich, 2004). Thus, understanding how ALAN affects communities in urban  
25 environments will be of worth value for urban planning actions.

26         Although bats are the mammal order with the highest diversity in urban areas and a  
27 primary component of urban nocturnal communities, (Nunes, Rocha, & Cordeiro-Estrela, 2017;  
28 Santini et al., 2019) there is still a lack of knowledge on how they use urban environments and  
29 the role they may have in maintaining ecological processes in urban matrices, especially in the  
30 Neotropical region. There is an urgency and global priority on urban bat studies (Bernard et al.,  
31 2012; Pacheco & Sodré, 2010; Russo & Ancillotto, 2015) because they are highly prone to be  
32 affected by ALAN (Stone, Harris, & Jones, 2015). Some studies have observed that bat  
33 communities in urban areas are restricted to few species specially composed of generalist species  
34 of frugivore, nectarivore and insectivore species (Frank, Gabbert, Chaves-Campos, & LaVal,  
35 2019; Jung & Kalko, 2010). Because urban areas are already associated with low bat diversity  
36 (Russo and Ancillotto 2015), it is vital to understand how ALAN affects different bat species and  
37 their activity in green remnants of urban environments, to avoid the loss of bat species at local  
38 scales.

39         It is possible to classify aerial insectivore bats into functional guilds based on whether  
40 they forage in open areas or edge spaces near environmental clutter (Denzinger, Tschapka, &  
41 Schnitzler, 2018). Studies have shown that artificial lights benefit aerial insectivore bats that  
42 catch prey on the wing, while other bats are not benefited (Jung & Kalko, 2010; Lewanzik &  
43 Voigt, 2017). There are also studies registering that fast-flying bats, open-air foragers –like  
44 Molossidae- are attracted to artificial lights, whereas slow-flying bats, cluttered environment  
45 foragers –like vespertilionids - are not. The latter evade artificial lights to avoid exposition to  
46 predators (Jones & Rydell, 1994; Jung & Kalko, 2011). It seems that the type of functional guild

47 and the flying speed of the bats are essential traits that will determine the ability of aerial  
48 insectivores to deal with artificial lights of different types (Frank et al., 2019; Lewanzik & Voigt,  
49 2017).

50         However, studies evaluating the relationship between bat fauna and resource availability  
51 in urban environments are still scarce, especially in Neotropical areas, where the urbanization  
52 process happens without planning, and the maintenance of green areas in urban centers is not a  
53 priority (Nunes et al., 2017; Pickett et al., 2001; Silva de Araújo & Bernard, 2016). Few studies  
54 are dealing with the effects of artificial light on bats in the Neotropical region, and they refer to  
55 cities with neighboring tropical forest environments (Avila-Flores & Fenton, 2005; Frank et al.,  
56 2019; Jung & Kalko, 2010). We aim to assess how ALAN affects insectivorous bats in urban  
57 parks (UP) of a highly populated city surrounded by cerrado (savanna) vegetation and extensive  
58 farming. Therefore, we are testing the following hypothesis: the first is that bat activity is higher  
59 near sites with ALAN, because the abundance of insects is also higher; the second, is that the  
60 activity of fast-flying species is higher near ALAN while slow-flying species present a negative  
61 response to ALAN, probably to avoid exposition to predators in open areas; and the third, that  
62 fast-flying species are positively related to arthropod orders, as they are more active near the  
63 ALAN source where there is higher abundance of arthropods.

## 64 **2. Material and Methods**

### 65 *2.1. Study Area*

66         Brasília is a densely green city located within the Cerrado. The climate is characterized  
67 by a very marked dry period, from April to September, and a rainy season, from October to  
68 March. Brasília is located about 1,000 meters above sea level (GDF, 2019). We selected six  
69 urban parks located within Brasília. Two are multi-use parks: named Olhos d'Água – OAEMP -

70 with 21.54ha, and Asa Sul Multi-use Park – ASMP - with 21.73ha. The other Parks are named  
71 ecological parks: Dom Bosco Ecological Park – DBEP - with 131.14ha; Águas Claras Ecological  
72 Park – ACEP – with 86.10ha, Ezechias Heringer Ecological Park – EHEP – with 306.44ha, and  
73 Burle Marx Ecological Park –BMEP - with 280ha (Figure 1). All urban parks have typical  
74 Cerrado vegetation, with small portions of gallery forest, and some species of exotic plants. They  
75 are open to the public during the day and broadly used by the local population for recreational  
76 purposes (IBRAM, 2013).

## 77 *2.2. Data Collection*

78 To estimate the effects of artificial lighting in bat activity and arthropod availability, we  
79 selected eight sites in each park (four sites in lit areas and four sites in unlit areas), with a  
80 minimum distance of 130m between sites. High-pressure sodium (HPS) lamps were present in all  
81 of the Urban Parks sampled. Because of the small area with artificial light available in DBEP,  
82 only two sites in lit treatment we selected in this area, totaling 46 sampling sites. We collected  
83 data twice in each season, from September/2018 to February/2019. We registered artificial  
84 lighting intensity (in lux, with Instrutemp ITLD 270I) in each sampling site.

85 To assess insectivorous bat activity, we placed AudioMoth ultrasound detectors (vs.  
86 1.0.0, Open Acoustic Devices, UK) installed at a 3-m-height pole or trees from the ground  
87 (Figure 2), recording at frequencies from 0 to 384Hz. Recordings were performed every 5  
88 minutes (1-min recording, 4-min interval) during 12 hours, starting at sunset. Recorded files  
89 were analyzed in Raven Pro (vs. 1.5. Cornell Lab of Ornithology, USA) and bat passes were  
90 considered as a sequence of pulses with intervals smaller than 1s between them (Fenton,  
91 Jacobson, & Stone, 1973). Calls were identified to species-level whenever possible, using an  
92 identification guide written by Arias-Aguilar et al. (2018) that sampled bats in Cerrado. We



93 measured the main acoustics characteristics of the pulse such as minimum and maximum  
94 frequencies (kHz), peak frequency (kHz), duration of the pulse (ms), and the interval between  
95 pulses (ms) to proceed with bat sound identification. We assigned species to functional guilds  
96 (open vs. edge forager) following Denzinger et al. (2018), and as fast or slow flier following  
97 Jung and Kalko (2010). Because the public light pole structure could be found within the park or  
98 in the street outside the parks, the recorders were installed under the lit areas and were placed as  
99 close as possible to the artificial lighting pole (Figure 2).

100 To quantify resource availability, we measured arthropod abundance by conducting  
101 captures with 25×50cm yellow sticky traps (BioControle, Brazil), installed at a 5-m-height pole  
102 from the ground approximately 2m apart from the ultrasound detectors (Figure 2). During all  
103 night, sticky traps were checked every hour, we use pens with different colors and geometrical  
104 shapes that represented each hour. Those sticky traps were then taken to the laboratory to  
105 proceed with the arthropods identification and were identified to the family level under the  
106 criteria of (Rafael, Melo, Carvalho, Casari, & Constantino, 2012).

### 107 *2.3. Data analysis*

108 We developed a Structural Equation Model (SEM) to assess how direct and indirect  
109 predictors associated with ALAN affect insectivorous bat activity and buzz ratio. We included  
110 artificial lighting intensity, represented by lux, and hour after sunset (1 to 12) as exogenous  
111 (indirect) factors, which would indirectly affect bat activity. As an endogenous (direct) factor,  
112 which would directly influence bat activity, we included only arthropod abundance (considered  
113 number of arthropods/hour). We also developed different SEMs to evaluate how direct and  
114 indirect predictors affect the activity of most abundant bat species, with over 100 individual  
115 passes.

116 We conducted a Redundancy Analysis (RDA) to evaluate how different bat species  
117 respond to different arthropod families throughout the night comparing matrices containing data  
118 for bats (dependent variable) and arthropods (independent variable) (Legendre & Legendre,  
119 1998). Bat species matrix was transformed (Hellinger distances) (Legendre & Gallagher, 2001)  
120 in order to determine the relationship between the occurrence of each bat species and the  
121 arthropods orders via Multiple Regression Models (Legendre, 2005; Legendre & Gallagher,  
122 2001). Statistical significance for the RDA was based on permutation tests of pseudo-F values  
123 computed as the ratio between constrained and unconstrained ordination (Legendre & Legendre,  
124 1998). Species groups were then visually identified (RDA graphics) based on the strength of  
125 their relationship with arthropod orders. Only the most abundant bat species and arthropod orders  
126 with more than 10 individuals were used for the analysis.

127 We conducted all analysis on R software (vs. 3.5.2, R Core Team, 2018) by using  
128 packages “Vegan” (Oksanen et al., 2008) for RDA, and “sem” (Fox et al., 2017) and “Lavaan”  
129 (Rosseel, 2012) for SEM.

### 130 **3. Results**

#### 131 *3.1. General Results*

132 We performed twenty-four nights of sampling, where 13,824 sound files were recorded in  
133 the WAV format. We used 12,821 files for the analyses, totaling 769,260 minutes sampled. The  
134 1003 files were not used due to recording problems. We recorded a total of 11,773 individual  
135 passes and 1,683 feeding buzzes. Six thousand two hundred thirty-three passes were in lit sites,  
136 and 5,540 in dark sites and 954 feeding buzzes in the lit site and 729 in the dark site. We  
137 managed to identify 11,015 bat passes from four families, 14 genera, and 20 species. The  
138 Molossid family was the most registered, with 5,690 passes in the lit sites and 3,607 passes in the

139 unlit site, followed by the Vespertilionid family with 107 passes in the lit sites and 1,226 passes  
 140 in the unlit sites. The Emballonurid Family was the third most registered, with 72 passes in the lit  
 141 sites and 293 in the unlit sites. (Table S1). *Molossus molossus* (n = 4,945; 42.01%), followed by  
 142 *Eumops sp./Nyctinomops macrotis* (n = 1,212; 10.29%) and *Molossus currentium* (n = 1,013;  
 143 8.61%) were the most abundant species.

144 Ninety-six sticky plates were installed for arthropod sampling during the 24 sampling  
 145 nights and 91 sticky plates were used for analysis. Five sticky plates were not used because of  
 146 problems in the traps. We captured 3,447 arthropods, from which 3,343 were identified as ten  
 147 orders and 32 families, among insects and arachnids. The most abundant in general were Diptera  
 148 (n = 2,436; 70.7%), followed by Hemiptera (n = 726; 21.1%) and Coleoptera (n = 92; 2.7%). The  
 149 most common orders in the lit and unlit site is Diptera, Hemiptera, and Coleoptera (Table S3).

### 150 3.2. Effect of ALAN on Bat Activity and Arthropod Abundance

151 Artificial lighting increased the general abundance of arthropods (estimate = 0.285,  $p <$   
 152 0.01) but did not affect bat activity (estimate = 0.001,  $p = 0.75$ ) nor buzz ratio (estimate = 0.001,  
 153  $p = 0.51$ ) directly. The latter was positively affected only by arthropod abundance ( $p < 0.01$  for  
 154 both) (Figure 3). Among the most abundant species, activity of fast-flying species *M. molossus*  
 155 (estimate = 0.177  $p < 0.01$ ), *Molossus sp1* (estimate = 0.012,  $p < 0.01$ ), and *Cynomops sp1*  
 156 (estimate = 0.024,  $p < 0.01$ ) were positively affected by arthropod abundance but had no direct  
 157 response to ALAN. Activity of *Eumops sp./N. macrotis* was not affected by ALAN but was  
 158 negatively affected by arthropod abundance (estimate = -0.013,  $p = 0.037$ ). Activity of *Eumops*  
 159 *perotis* and *M. currentium* increased in areas near ALAN (estimate = 0.014,  $p < 0.01$  and  
 160 estimate = 0.021,  $p = 0.001$ , respectively) and with higher arthropod abundance (estimate =  
 161 0.005,  $p = 0.033$  and estimate = 0.060,  $p < 0.01$ , respectively). Activity of slow-flying species

162 *Eptesicus furinalis* (estimate = -0.024,  $p = 0.001$ ) and *Eptesicus brasiliensis* (estimate = -0.031,  $p$   
163 = 0.012) were negatively affected by ALAN, being higher in the dark areas of the parks. Activity  
164 of *Peropteryx macrotis*, *Peropteryx trinitatis*, and *Eumops* sp1 was not affected by ALAN or  
165 arthropods abundance.

### 166 3.3. Bats Species $\times$ Arthropod Group

167 Redundancy Analysis (RDA) significantly classified bat species groups according to  
168 arthropod orders presence throughout the night ( $F = 4.08$ ,  $p = 0.001$ ). First axis explained 21.3%  
169 of species variation in relation to hour after sunset and second axis explained 16.1% of total  
170 variation. Two main groups were observed. The first group included most of Molossidae species  
171 (*M. molossus*, *Molossus* sp1., *M. currentium*, and *Cynomops* sp1.) were explained by arthropod  
172 orders (Diptera, Hemiptera, Coleoptera, Hymenoptera and Aranae) responding to the same  
173 temporal pattern. Second group was formed by *Eumops* sp./*N. macrotis*, *Eumops* sp, *E. perotis*,  
174 *E. furinalis*, *E. brasiliensis*, *P. macrotis* and *P. trinitatis*, with no relationship to the arthropods  
175 present in the records (Figure 4).

## 176 4. Discussion

### 177 4.1 Effects of ALAN on Bat Activity and Arthropod Abundance

178 In general, the increased bat activity near artificial light is a result of the higher resource  
179 availability provided in those illuminate areas as shown by several studies conducted in Europe,  
180 North and Central America (Frank et al., 2019; Rowse, Harris, & Jones, 2016). The relationship  
181 between higher arthropod abundance and bat activity is well-documented in literature with  
182 studies done in forest environments (Avila-Flores & Fenton, 2005; Jung & Kalko, 2010).  
183 However, little is known about how this relationship takes place in open areas, such as savanna  
184 habitats. This is the first study evaluating how insectivorous bats in an urban environment within

185 a Neotropical Savanna respond to light presence, showing indirect effects of ALAN on bat  
186 activity by the increasing arthropod abundance around ALAN poles. Light presence lures insects,  
187 thus attracting bats, in a bottom-up effect system, corroborating our study first hypothesis that  
188 bat activity is higher near sites with ALAN, because the abundance of insects is also higher. We  
189 observed more activity of fast-flying species, such as *M. molossus*, *M. currentium*, *Molossus* sp1,  
190 *E. perotis* and *Cynomops* sp1. near ALAN sites, when compared to slow-flying species such as  
191 *E. furinalis* and *E. brasiliensis*. Thus, confirming our second hypothesis that the activity of fast-  
192 flying species is higher near ALAN while slow-flying species present a negative response to  
193 ALAN, probably to avoid exposition to predators in open areas. We also observed that *M.*  
194 *molossus*, *M. currentium*, *Molossus* sp1 and *Cynomops* sp1. are related to arthropod order,  
195 partially confirming our third hypothesis that fast-flying species is positively related to arthropod  
196 orders, as they are more active near the ALAN source where there is higher abundance of  
197 arthropods. However, the large molossids (*Eumops* sp/*Nyctinomops macrotis*, *Eumops* spp and *E.*  
198 *perotis*) were unrelated.

199 Another factor influencing ALAN effects on bat activity is the type of light used for  
200 public illumination (Stone, Wakefield, Harris, & Jones, 2015). Lewanzik & Voigt (2017)  
201 observed higher bat activity in areas with mercury vapor lamps when compared to LED lamps.  
202 However, according to Rowse et al. (2016), sodium vapor lamps, which are used to illuminate  
203 urban parks in our study area, had similar effects on bat activity when compared to LED lamps.  
204 In our study area, overall bat activity was not significantly different between lit and unlit  
205 treatments, but for sure, sodium vapor lamps attracted arthropods as was observed in previous  
206 studies (Eisenbeis & Hänel, 2009).

207 We observed that ALAN adequately attracted Diptera; however, the vespertilionid *E.*  
208 *furinalis*, a predator of this order (Aguiar & Antonini, 2008) was intolerant to light presence,  
209 which could reduce the service provided by this bat species as a population regulator of those  
210 insects. This situation may be especially detrimental because Diptera is considered one of the  
211 most critical insect order regarding public health issues in Brazil, because of its role as vector of  
212 different diseases (Lourenço & Rodrigues, 2017). Increased abundance of arthropods near urban  
213 ALAN areas can result in a shift in the food chain of bats preying in urban ALAN areas, with  
214 significant shift on diet consumed items compared to the diet of bats foraging far from artificial  
215 lights (Cravens, Brown, Divoll, & Boyles, 2018; Holzhauer et al., 2015; Manfrin et al., 2017).

216 Studies conducted in the Neotropical region observed that the traits of slow flight and  
217 edge spaces bats are not predictors of a light-avoidant species (Frank et al., 2019; Jung & Kalko,  
218 2010). In our study, the species with these traits, *P. macrotis* and *P. trinitatis* did not have their  
219 activity affected by ALAN and neither by the abundance of arthropods. Though, *E. furinalis* and  
220 *E. brasiliensis* had their activity negatively affected by ALAN. The difference between our  
221 results and those of the others carried out in the Neotropical region may be related to the regional  
222 factor, since the Cerrado has savannah vegetation and large open areas, the presence of artificial  
223 lighting may not be beneficial for slow-flight and edge spaces, increasing the exposure to  
224 predators.

225 Contrary to slow-flight species, most Molossidae species, considered fast-flying species,  
226 were positively influenced by the higher arthropod abundance brought by ALAN presence, either  
227 directly (*E. perotis* and *M. currentium*) or indirectly (*M. molossus*, *Molossus* sp1, *Cynamops*  
228 sp1). High activity levels of fast-flying bats foraging near artificial lights have been recorded in  
229 previous studies conducted in Neotropical and European forests (Frank et al., 2019; Jung &

230 Kalko, 2010, 2011; Stone, Wakefield, et al., 2015). Because molossids are fast in flight, with  
231 little maneuverability to cluttered habitats, they are well adapted to forage in open spaces such as  
232 cities (Denzinger et al., 2018) , and more likely to escape from predators as they can fly faster  
233 from predators during foraging time (Jones & Rydell, 1994).

234 Three species, *M. molossus*, *Eumops* sp./*Nyctinomops macrotis*, and *M. currentium*  
235 accounted for 60.9% of all recorded bat passes, suggesting a dominance of few species in urban  
236 environments, as observed in previous studies (Azam et al., 2018; Jung & Kalko, 2010;  
237 Lewanzik & Voigt, 2017). In Panama, over 95% of recorded passes in urban areas were from  
238 molossids, a rate significantly higher than the one found in natural areas (Jung & Kalko, 2010).  
239 Artificial lighting may be acting as an environmental filter selecting species adapted to the new  
240 scenario provided by urbanization (Aronson et al., 2016), especially fast-flying species, such as  
241 molossids, leading to a pattern observed in urban areas that are entirely different from those  
242 observed in natural habitats.

#### 243 4.2 Bats Species × Arthropod Group

244 We observed temporal segregation on insectivorous bat activity based on the presence of  
245 arthropod groups, as demonstrated by the Redundancy analysis (RDA) results. Small fast-flying  
246 species of Molossidae (*M. molossus*, *M. currentium*, *Molossus* sp1 and *Cynomops* sp1) are the  
247 first to show up at night (Figure 4), and this behavior is related to the activity of the most  
248 abundant arthropod orders registered in this study. These molossids species are more active in  
249 the early evening (Table S5 and Figure S1, S2, S3 and S8) as indicated by the SEM analyses.  
250 Large molossids (*Eumops* sp./*N. macrotis*, *Eumops* sp, *E. perotis*) and *E. furinalis*, *E.*  
251 *brasiliensis*, *P. macrotis* and *P. trinitatis* compose the group not related to any of the most  
252 abundant arthropod orders (Figure 4). These species showed no variation in activity in response

253 arthropod abundance, except *E. perotis* (Table S3). For the small molossids, our results  
254 corroborate the results of Jones & Rydell (1994) who observed species that feed on small  
255 arthropods emerging during the first hours of the night.

256 On the other hand, larger species such as *Eumops* sp./*N. macrotis*, *Eumops* sp, *E. perotis*  
257 feed on larger arthropods and do not depend on the abundance of the smaller arthropods present  
258 in the first hours of the evening, foraging through all the night (Jones & Rydell, 1994) (Figure  
259 S5, S6 and S7). In our study the three most recorded arthropod orders (Diptera, Hemiptera, and  
260 Coleoptera) are among the main items consumed by molossids (Bohmann et al., 2011; Fenton et  
261 al., 1998; Lee & McCracken, 2002), suggesting that arthropods abundance may be explaining  
262 how bat species separate themselves throughout the night. The absence of Lepidoptera in our  
263 records, other common item in molossids diet (Bohmann et al., 2011; Fenton et al., 1998; Lee &  
264 McCracken, 2002) may be a result of the employed method of capture. Stick traps are selective  
265 in arthropods capture (Tang et al., 2016). Although, our results recorded the same orders,  
266 Diptera, Hemiptera, Coleoptera, Orthoptera and Hymenoptera, that are known to compose the  
267 diet of the species *E. brasiliensis*, *E. furinalis*, *P. macrotis* and *P. trinitatis* (Aguiar & Antonini,  
268 2008; Garcia, Machado, Nunez, Delgado-Jaramillo, & Aular, 2011). Our RDA results for these  
269 bat species, from the families Vespertilionidae and Emballonuridae, corroborate what was  
270 observed in SEM, where their activity was not correlated with the abundance of arthropods.  
271 (Figure 4) (Table S5). This result may be an indicative of the effects of urbanization on those bat  
272 families, usually adapted to forage in cluttered or background cluttered areas (Kalko, Handley, &  
273 Handley, 1996).

274 The higher abundance of Molossidae species in our study areas suggests they are highly  
275 tolerant to urban environments and are benefited from the high abundance of arthropods near the



276 ALAN. Furthermore, the presence of molossids in urban environments may also be related to  
277 higher roost availability in those areas since human-made structures can be similar to natural  
278 cracks used by molossids species (Gaisler et al. 1998, Keeley and Keeley 2004). Additionally,  
279 the presence of species intolerant to ALAN, such as *E. brasiliensis* and *E. furinalis* may be due  
280 to the high level of afforestation in the city of Brasília. Straka, Wolf, Gras, Buchholz, & Voigt,  
281 (2019) observed that a dense tree cover might potentially minimize effects of ALAN in intolerant  
282 species, which highlights the importance of preserving natural remnants in the urban area in  
283 order to preserve insectivorous bat communities and their ecosystem services (Silva de Araújo &  
284 Bernard, 2016).

## 285 **5. Conclusion**

286 The activity of insectivorous bats is indirectly affected by ALAN, which positively  
287 affects the abundance of arthropods, in a bottom-up effect. We also concluded that fast-flying  
288 bats that forage in open areas benefit from ALAN, with greater activity near the illuminated  
289 sites. The presence of species that are intolerant to ALAN in our study, such as *E. furinalis* and  
290 *E. brasiliensis*, may be an indication of the importance of urban parks, and the importance of  
291 green remnants. These structures can be significant for the maintenance and conservation of the  
292 populations of bats.

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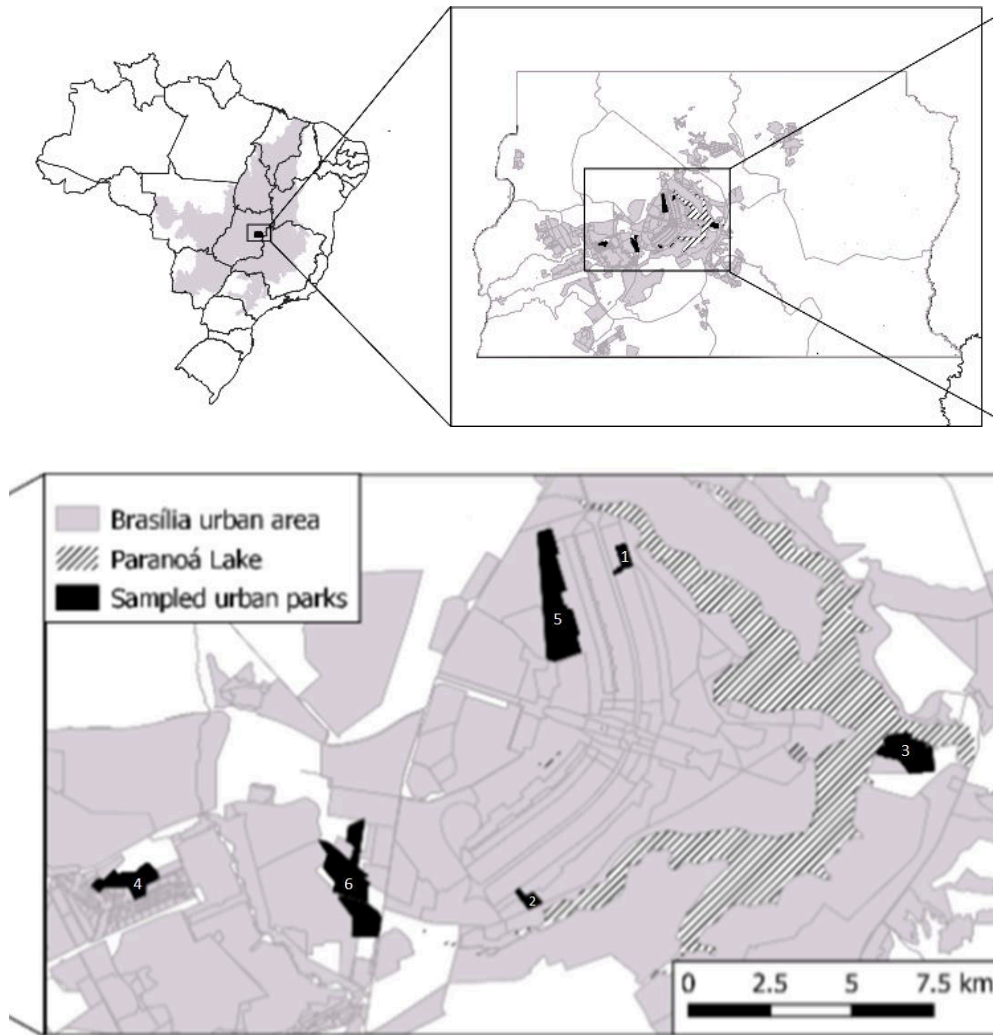
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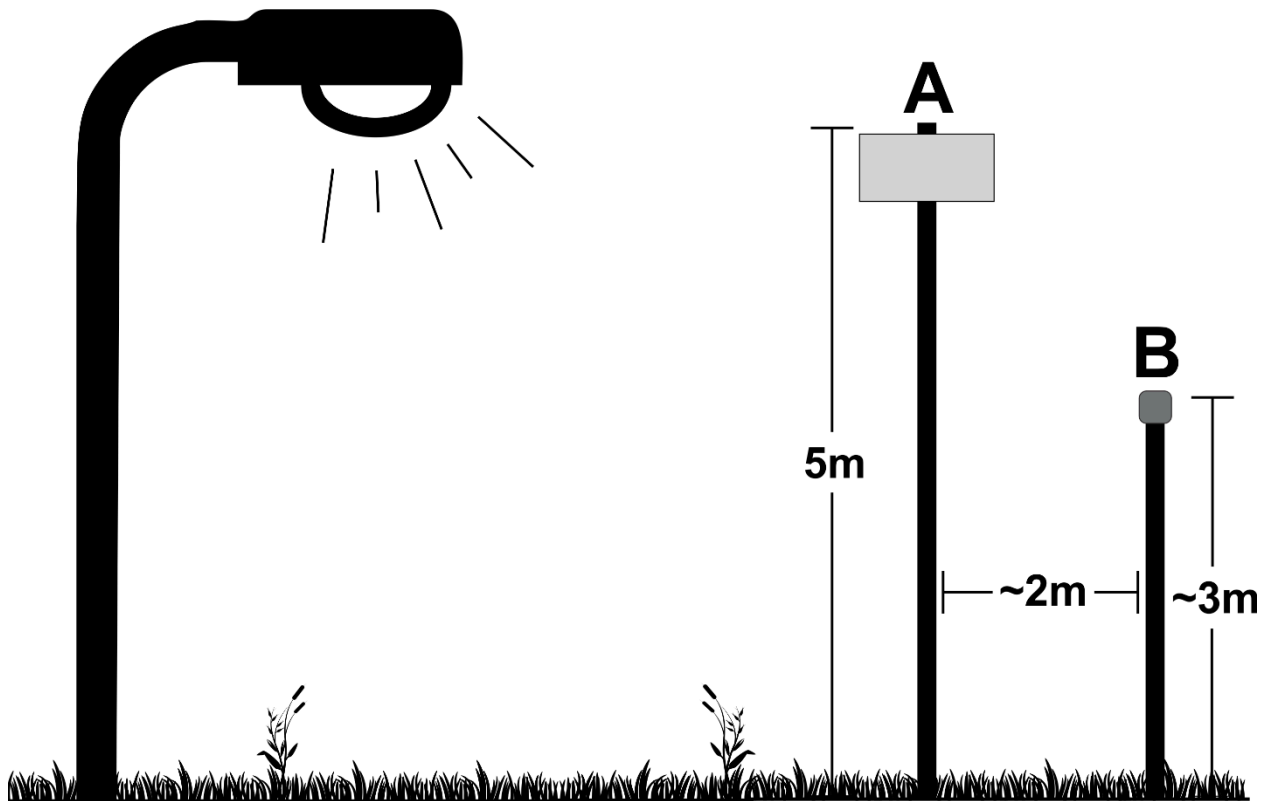
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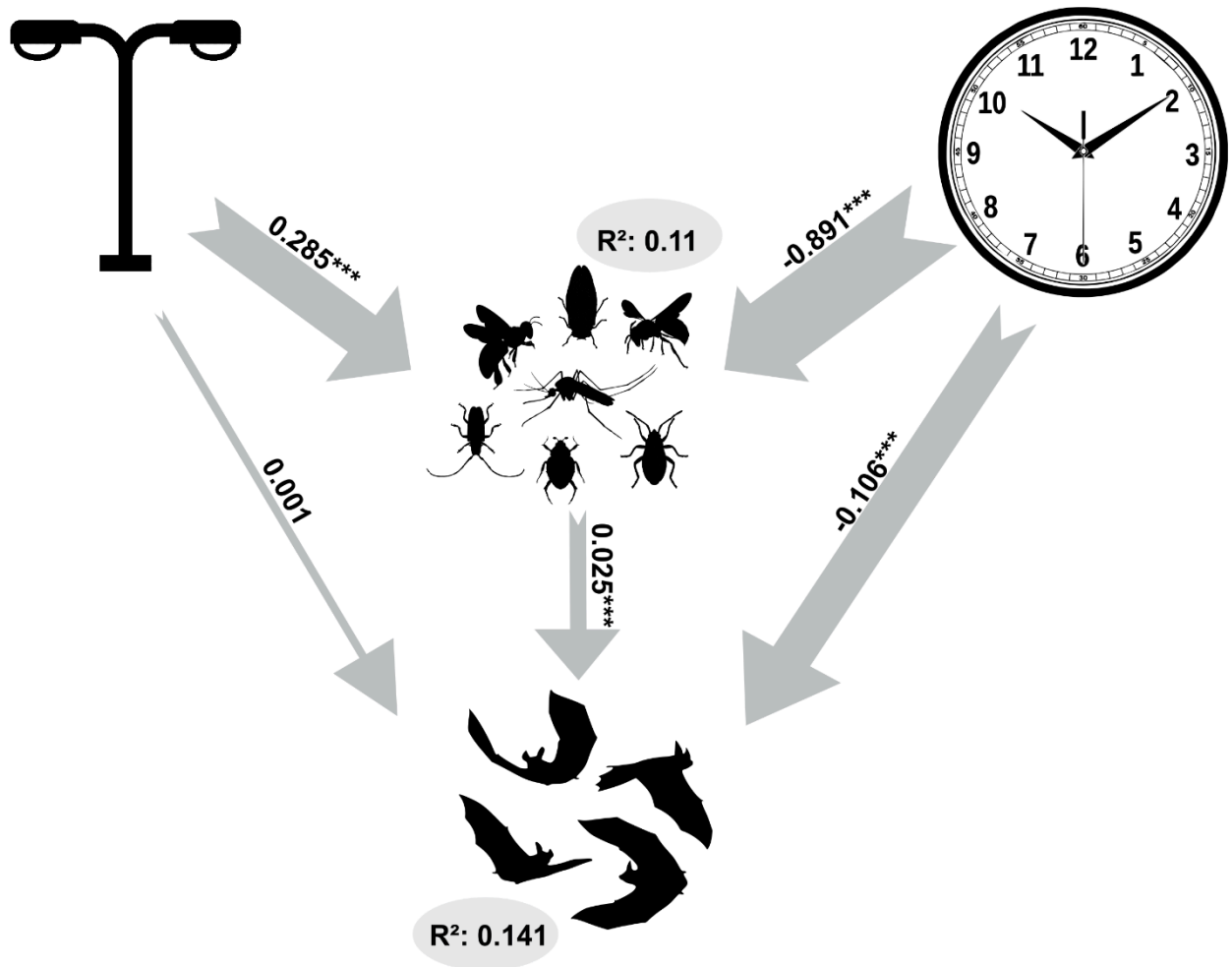
## 7. Figures



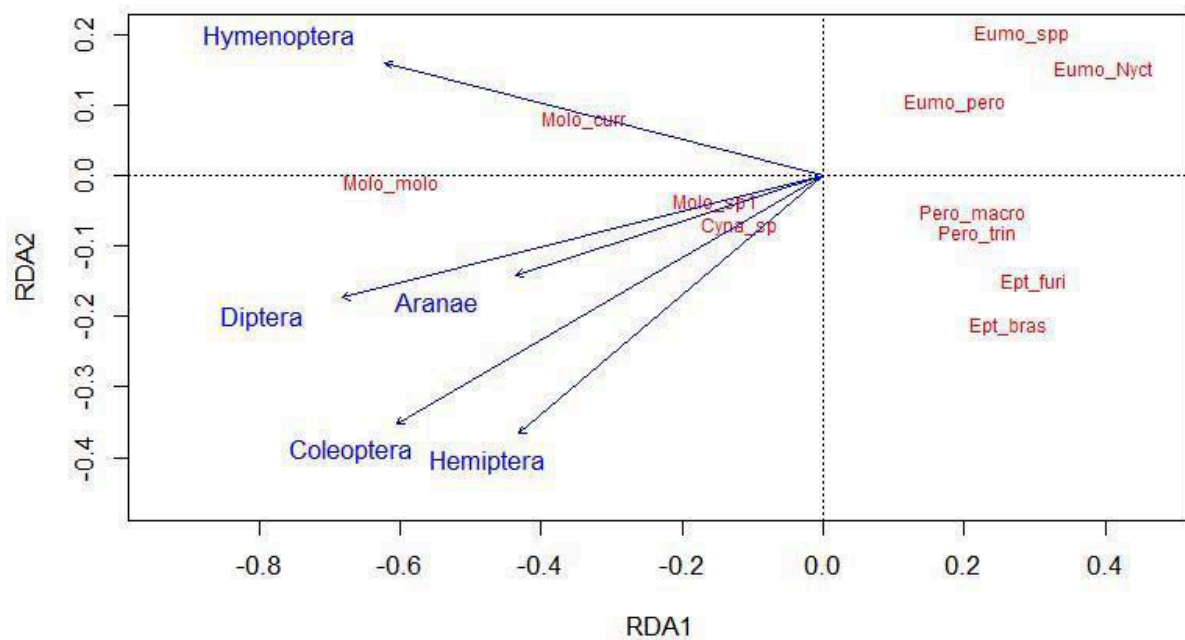
**Figure 1.** Study area showing Federal District within Cerrado and Brazil, and location of the six analyzed Urban Parks (UP) within Brasília urban matrix. 1 = OAEMP – Olhos d’Água Ecological Multi-use Park, 2 = ASMP – Asa Sul Multi-use Park, 3 = DBEP – Dom Bosco Ecological Park, 4 = ACEP – Águas Claras Ecological Park, 5 = BMEP – Burle Marx Ecological Park and 6 = EHEP – Ezechias Heringer Ecological Park.



**Figure 2:** Scheme of how recorder (B) and sticky trap (A) were positioned and the distance of both was approximately 2 meters. The recorders (B) installed at approximately 3-m-height pole or tree and the sticky trap (A) installed at a 5-m-height pole. The distance of both, recorder and the sticky trap, from the artificial light source varied in each park.



**Figure 3.** Structural Equation Model for bat species and arthropod orders located in the city of Brasília, Federal District of Brazil, from September/2018 to February/2019. Arrow width represents relative strength of each effect and significant effects are indicated with asterisks (\* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ ). Determination coefficient ( $R^2$ ) is represented in gray circles to both dependent variables (bat activity and insect abundance).



**Figure 4.** Redundancy Analysis (RDA) biplot showing the relationship between bat species and arthropod orders located in the city of Brasília, Federal District of Brazil, from September/2018 to February/2019. First axis represents 21.3% of variation according to night hours and axis 2 represents 16.1% of total variation. *Note:* Molo\_molo: *M. molossus*, Molo\_curr: *M. currentium*, Molo\_sp1: *Molossus sp1*, Cyno\_sp: *Cynomomops sp1*, Eumo\_spp: *Eumops spp*, Eumo\_pero: *E. perotis*, Eumo\_Nyct: *Eumops sp/ Nyctinomops macrotis*, Pero\_macro: *P. macrotis*, Pero\_trin: *P. trinitratis*, Ept\_furi: *E. furinalis*, Ept\_bras: *E. brasiliensis*.

## 8. Supplementary Material

**Table S1.** Bat species recorded in Urban Parks located in the city of Brasília, Federal District of Brazil, from September/2018 to February/2019 in lit and unlit treatments with its respective passes number and relative abundance (RA). Species were classified into functional guilds as either open-air foragers or edge foragers based on Denzinger et al.(2018), and as fast or slow fliers based on Jung & Kalko (2010).

Family	Foraging	Passes in unlit areas	Passes in lit areas	RA (%)
Species				
<b>Emballonuridae</b>				
<i>Peropteryx macrotis</i>	Slow edge	175	55	1.95
<i>Peropteryx trinitatis</i>	Slow edge	118	17	1.15
<b>Molossidae</b>				
<i>Cynomops sp1</i>	Fast open	290	300	5.01
<i>Cynomops planirostris</i>	Fast open	30	36	0.56
<i>Cynomops</i> spp.	Fast open	23	20	0.36
<i>Eumops</i> sp./ <i>Nyctinomops macrotis</i>	Fast open	503	709	10.29
<i>Eumops</i> spp.	Fast open	275	408	5.8
<i>Eumops perotis</i>	Fast open	96	138	1.99
<i>Molossus currentium</i>	Fast open	311	702	8.61
<i>Molossus molossus</i>	Fast open	1831	3114	42.01
<i>Molossus rufus</i>	Fast open	0	5	0.04
<i>Molossus sp1</i>	Fast open	153	155	2.62
<i>Molossops temminckii</i>	Fast open	38	52	0.76
<i>Molossops</i> spp.	Fast open	17	41	0.49
<i>Nyctinomops laticaudatus</i>	Fast open	9	6	0.13
<i>Promops nasutus</i>	Fast open	7	2	0.08
<i>Promops centralis</i>	Fast open	1	1	0.02
<i>Promops</i> spp.	Fast open	23	1	0.2
<b>Phyllostomidae</b>				
NI		14	6	0.17
<b>Vespertilionidae</b>				
<i>Eptesicus brasiliensis</i>	Slow edge	622	21	5.46
<i>Eptesicus furinalis</i>	Slow edge	499	55	4.71

*Continue Table S1*

<b>Family</b>	Foraging	Passes in unlit areas	Passes in lit areas	RA (%)
Species				
<i>Histiotus diaphanopterus</i>	Slow edge	1	0	0.01
<i>Histiotus velatus</i>	Slow edge	7	1	0.07
<i>Lasiurus blossevillii</i>	Slow edge	3	1	0.03
<i>Lasiurus cinereus</i>	Slow edge	13	6	0.16
<i>Lasiurus ega</i>	Slow edge	28	13	0.35
<i>Myotis albescens</i>	Slow edge	4	1	0.04
<i>Myotis lavalii</i>	Slow edge	18	0	0.15
<i>Myotis spp.</i>	Slow edge	30	9	0.33
<i>Rhogeessa hussoni</i>	Slow edge	1	0	0.01
<b>NI</b>		400	358	6.44
<b>TOTAL</b>		5540	6233	100

**Table S2:** Bat species recorded in each Urban Parks sampled in the city of Brasília, Federal District of Brazil, from September/2018 to February/2019 in lit and unlit treatments with its respective passes number. OAEMP – Olhos d’Água Ecological Multi-use Park, ASMP – Asa Sul Multi-use Park, DBEP – Dom Bosco Ecological Park, ACEP – Águas Claras Ecological Park, BMEP – Burle Marx Ecological Park and EHEP – Ezechias Heringer Ecological Park.

Family Species	OAEMP		ASMP		DBEP		ACEP		BMEP		EHEP	
	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit
<b>Emballonuridae</b>												
<i>Peropteryx macrotis</i>	9	26	51	15	22	11	81	0	1	0	11	3
<i>Peropteryx trinitatis</i>	2	3	10	2	14	8	89	0	0	1	3	3
<b>Molossidae</b>												
<i>Cynomops greenhalli</i>	70	60	64	47	30	63	20	19	48	74	58	37
<i>Cynomops planirostris</i>	2	3	10	11	0	1	2	4	7	7	9	10
<i>Cynomops sp.</i>	8	8	2	7	5	3	3	1	3	0	2	1
<i>Eumops sp./Nyctinomops macrotis</i>	78	104	106	216	115	128	43	48	48	20	113	193
<i>Eumops spp.</i>	23	17	23	44	60	93	129	10	6	2	34	242
<i>Eumops perotis</i>	0	4	9	6	60	116	22	2	5	5	0	5
<i>Molossus currentium</i>	52	112	50	110	57	185	47	60	26	35	79	200
<i>Molossus molossus</i>	237	372	516	680	183	634	301	208	112	225	482	995
<i>Molossus rufus</i>	0	5	0	0	0	0	0	0	0	0	0	0
<i>Molossus sp.</i>	22	13	34	26	8	40	20	4	9	17	60	55
<i>Molossops temminckii</i>	0	21	22	8	0	2	6	1	2	14	8	6
<i>Molossops spp.</i>	1	5	3	11	3	2	0	0	6	21	4	2
<i>Nyctinomops laticaudatus</i>	0	1	2	1	0	0	0	2	5	2	2	0



*Continue Table S2*

Family	OAEMP		ASMP		DBEP		ACEP		BMEP		EHEP	
	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit
<i>Promops nasutus</i>	3	0	2	1	0	0	1	0	0	1	1	0
<i>Promops centralis</i>	0	0	0	1	0	0	0	0	1	0	0	0
<i>Promops</i> spp.	5	0	1	0	7	0	3	0	0	1	7	0
<b>Phyllostomidae</b>												
NI	3	0	10	6	0	0	0	0	0	0	1	0
<b>Vespertilionidae</b>												
<i>Eptesicus brasiliensis</i>	26	5	154	7	16	3	346	0	79	6	1	0
<i>Eptesicus furinalis</i>	40	2	87	37	2	2	232	0	138	14	0	0
<i>Histiotus diaphanopterus</i>	0	0	0	0	0	0	0	0	1	0	0	0
<i>Histiotus velatus</i>	2	0	1	0	4	1	0	0	0	0	0	0
<i>Lasiurus blossevillii</i>	0	1	1	0	2	0	0	0	0	0	0	0
<i>Lasiurus cinereus</i>	1	3	0	1	10	2	1	0	1	0	0	0
<i>Lasiurus ega</i>	0	0	27	12	1	1	0	0	0	0	0	0
<i>Myotis albescens</i>	2	0	1	0	0	0	1	1	0	0	0	0
<i>Myotis lavalii</i>	0	0	4	0	1	0	2	0	9	0	2	0
<i>Myotis</i> spp.	9	2	10	7	1	0	5	0	3	0	2	0
<i>Rhogeessa hussoni</i>	0	0	0	0	0	0	0	0	1	0	0	0

**Table S3.** Arthropod orders and families recorded in Urban Parks located in the city of Brasília, Federal District of Brazil, from September/2018 to February/2019 in lit and unlit treatments with its respective number of individuals and relative abundance (RA).

<b>FILO</b>			
<b>Order</b>	Number in unlit areas	Number in lit areas	RA(%)
Family			
<b>ARACHNIDA</b>			
<b>Aranae</b>	7	8	0.435
<b>INSECTA</b>			
<b>Blattodea</b>			
Blattellidae	1	0	0.029
<b>Coleoptera</b>			
Anthicidae	0	1	0.029
Bruchidae	18	27	1.305
Cantharidae	1	0	0.029
Chrysomelidae	13	3	0.464
Coccinelidae	0	5	0.145
Curculionidae	6	3	0.261
Latridiidae	4	3	0.203
Mycetophagidae	1	0	0.029
Staphylinidae	3	0	0.087
Tenebrionidae	1	3	0.116
<b>Diptera</b>			
NI	623	1813	70.67
<b>Hemiptera</b>			
Aleyrodidae	7	31	8.703
Alydidae	1	0	1.102
Aphididae	9	16	0.029
Cicadellidae	217	83	0.725
Fulgoridae	2	0	0.058
Lygaeidae	2	1	0.087
Membracidae	0	2	0.058
Miridae	6	4	0.29
Psyllidae	185	131	9.167
Reduviidae	18	7	0.725
Rhopalidae	0	1	0.029
Ricaniidae	0	1	0.029
Tingidae	1	0	0.029

*Continue Table S3*

<b>FILO</b>			
<b>Order</b>	Number in unlit areas	Number in lit areas	RA(%)
Family			
Tropiduchidae	1	0	0.029
<b>Ordem Hymenoptera</b>			
Apidae	0	5	0.145
Braconidae	0	2	0.058
Formicidae	14	26	1.16
Ichneumonidae	0	1	0.029
Vespidae	1	7	0.232
<b>Mantodea</b>			
Mantidae	0	1	0.029
<b>Neuroptera</b>			
Chrysopidae	0	2	0.058
<b>Orthoptera</b>			
Tettigoniidae	7	1	0.232
<b>Thysanoptera</b>			
NI	4	2	0.174
<b>Insecta NI</b>	59	45	3.017
<b>TOTAL</b>	1212	2190	100

**Table S4:** Arthropod orders and families recorded in each Urban Parks sampled in the city of Brasília, Federal District of Brazil, from September/2018 to February/2019 in lit and unlit treatments with its respective number of individuals. OAEMP – Olhos d'Água Ecological Multi-use Park, ASMP – Asa Sul Multi-use Park, DBEP – Dom Bosco Ecological Park, ACEP – Águas Claras Ecological Park, BMEP – Burle Marx Ecological Park and EHEP – Ezechias Heringer Ecological Park.

<b>FILO Order</b>	<b>OAEMP</b>		<b>ASMP</b>		<b>DBEP</b>		<b>ACEP</b>		<b>BMEP</b>		<b>EHEP</b>	
	<b>Dark</b>	<b>Lit</b>	<b>Dark</b>	<b>Lit</b>	<b>Dark</b>	<b>Lit</b>	<b>Dark</b>	<b>Lit</b>	<b>Dark</b>	<b>Lit</b>	<b>Dark</b>	<b>Lit</b>
<b>ARACHNIDA</b>												
<b>Aranae</b>	2	1	1	3	0	0	2	1	1	1	1	2
<b>INSECTA</b>												
<b>Blattodea</b>												
Blattellidae	1	0	0	0	0	0	0	0	0	0	0	0
<b>Cloleoptera</b>												
Anthicidae	0	0	0	0	0	1	0	0	0	0	0	0
Bruchidae*	0	5	15	16	2	1	0	4	0	0	1	1
Cantharidae*	1	0	0	0	0	0	0	0	0	0	0	0
Chrysomelidae*	3	0	6	0	1	2	1	0	1	0	1	1
Coccinellidae*	0	1	0	3	0	1	0	0	0	0	0	0
Curculionidae*	2	1	1	0	1	0	0	1	2	1	0	0
Latridiidae	2	1	1	0	0	0	0	0	0	1	1	1
Mycetophagidae	0	0	0	0	0	0	0	0	1	0	0	0
Staphylinidae*	1	0	1	0	1	0	0	0	0	0	0	0
Tenebrionidae*	0	1	0	0	1	2	0	0	0	0	0	0
<b>Diptera</b>												
NI	40	49	213	69	257	1517	61	103	21	38	31	37

*Continue Table S4*

<b>FILO</b>	<b>OAEMP</b>		<b>ASMP</b>		<b>DBEP</b>		<b>ACEP</b>		<b>BMEP</b>		<b>EHEP</b>	
<b>Order</b>	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit
<b>Hemiptera</b>												
Aleyrodidae*	1	0	0	1	3	0	0	30	3	0	0	0
Alydidae*	0	0	0	0	1	0	0	0	0	0	0	0
Aphididae*	3	6	3	3	1	2	0	4	1	0	1	1
Cicadellidae*	28	11	65	6	34	14	13	6	51	27	26	19
Fulgoridae*	1	0	0	0	0	0	0	0	1	0	0	0
Lygaeidae*	0	0	0	0	0	0	0	0	2	1	0	0
Membracidae	0	2	0	0	0	0	0	0	0	0	0	0
Miridae*	1	0	1	0	0	1	4	1	0	1	0	1
Psyllidae*	3	6	35	14	5	10	83	63	35	7	24	31
Reduviidae*	15	4	1	0	1	0	1	0	0	2	0	1
Rhopalidae	0	0	0	0	0	1	0	0	0	0	0	0
Ricaniidae	0	0	0	0	0	1	0	0	0	0	0	0
Tingidae*	0	0	0	0	0	0	0	0	1	0	0	0
Tropiduchidae	0	0	0	0	1	0	0	0	0	0	0	0
<b>Ordem Hymenoptera</b>												
Apidae*	0	0	0	1	0	4	0	0	0	0	0	0
Braconidae*	0	1	0	0	0	1	0	0	0	0	0	0
Formicidae*	3	0	3	0	3	20	1	2	1	2	3	2
Ichneumonidae*	0	0	0	0	0	0	0	1	0	0	0	0
Vespidae*	1	0	0	0	0	3	0	2	0	0	0	2

*Continue Table S4*

<b>FILO</b> <b>Order</b> Family	OAEMP		ASMP		DBEP		ACEP		BMEP		EHEP	
	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit	Dark	Lit
<b>Mantodea</b>												
Mantidae	0	0	0	0	0	1	0	0	0	0	0	0
<b>Neuroptera</b>												
Chrysopidae*	0	0	0	1	0	1	0	0	0	0	0	0
<b>Orthoptera</b>												
Tettigoniidae*	0	0	0	0	2	0	1	0	3	1	1	0
<b>Thysanoptera</b>												
NI	0	0	1	0	1	0	0	0	2	2	0	0

**Table S5.** Parameters estimated for disinfected Structural Equation Models (SEM). Related to response of the Hour after sunset, Arthropod Abundance and Lux to Arthropod Abundance, General Bat Activity, Buzz Ratio, *Molossus molossus*, *M currentium*, *Molossus sp1*, *Cynomops sp1*, *Eumops sp/Nyctinomops laticaudatus*, *Eumops sp1*, *E. perotis*, *Peropteryx macrotis*, *P. trinitratis*, *Eptesicus furinalis* and *E. brasiliensis*. Model of Arthropod Abundance same of all models.

<b>Models</b>	<b>Predictor Variables</b>	<b>Estimate</b>	<b>Std.Err</b>	<b>z-value</b>	<b>P(&gt; z )</b>	<b>Std.lv</b>	<b>Std.all</b>	<b>R<sup>2</sup></b>
Arthropod abundance	Hour after sunset	-0.891	0.111	-8.002	>0.001	-0.891	-0.236	0.11
	Lux	0.285	0.035	8.018	>0.001	0.285	0.236	
General Bat Activity	Hour after sunset	-0.106	0.013	-8.421	>0.001	-0.106	-0.252	0.141
	Arthropod Abundance	0.025	0.003	7.266	>0.001	0.025	0.223	
	Lux	0.001	0.004	0.325	0.746	0.001	0.01	
Buzz Ratio	Hour after sunset	0.002	0.002	1.032	0.302	0.002	0.033	0.044
	Arthropod Abundance	0.003	0.001	6.442	>0.001	0.003	0.209	
	Lux	>0.001	0.001	0.656	0.512	>0.001	0.021	
<i>Molossus molossus</i>	Hour after sunset	-0.792	0.085	-9.28	>0.001	-0.792	-0.273	0.163
	Arthropod Abundance	0.177	0.023	7.599	>0.001	0.177	0.23	
	Lux	0.036	0.027	1.32	0.187	0.036	0.039	
<i>Molossus currentium</i>	Hour after sunset	-0.075	0.019	-3.94	>0.001	-0.075	-0.115	0.178
	Arthropod Abundance	0.06	0.005	11.581	>0.001	0.06	0.348	
	Lux	0.021	0.006	3.378	0.001	0.021	0.099	
<i>Molossus sp1</i>	Hour after sunset	-0.049	0.011	-4.284	>0.001	-0.049	-0.135	0.042
	Arthropod Abundance	0.012	0.003	3.793	>0.001	0.012	0.123	
	Lux	0.001	0.004	0.291	0.771	0.001	0.009	

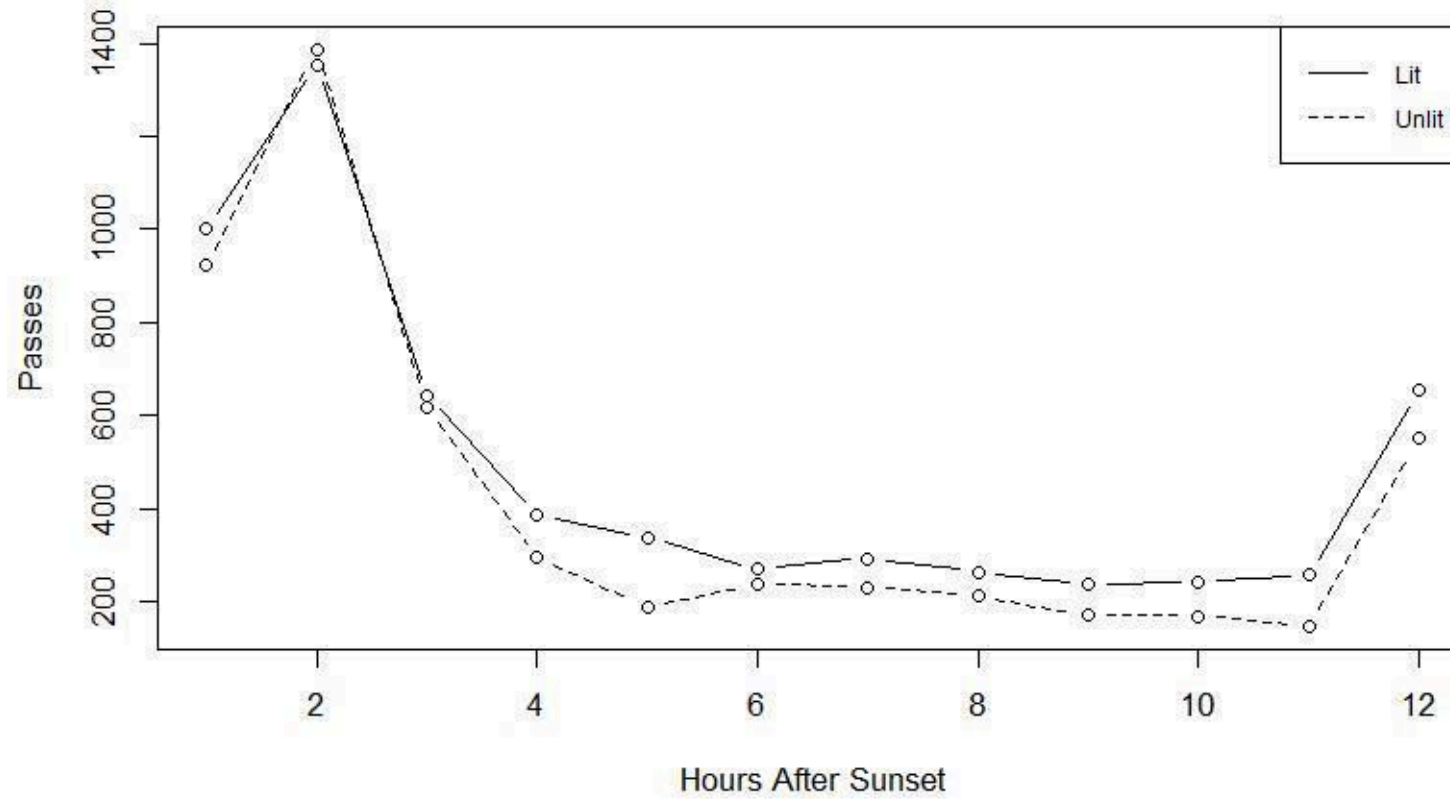
*Continue Table S5*

<b>Models</b>	<b>Predictor Variables</b>	<b>Estimate</b>	<b>Std.Err</b>	<b>z-value</b>	<b>P(&gt; z )</b>	<b>Std.lv</b>	<b>Std.all</b>	<b>R<sup>2</sup></b>
<i>Cynomops spl</i>	Hour after sunset	-0.085	0.016	-5.159	>0.001	-0.085	-0.161	0.066
	Arthropod Abundance	0.024	0.004	5.335	>0.001	0.024	0.171	
	Lux	-0.006	0.005	-1.19	0.234	-0.006	-0.037	
<i>Eumops sp/Nyctinomops laticaudatus</i>	Hour after sunset	0.004	0.023	0.165	0.869	0.004	0.005	0.006
	Arthropod Abundance	-0.013	0.006	-2.09	0.037	-0.013	-0.069	
	Lux	0.012	0.007	1.706	0.088	0.012	0.055	
<i>Eumops spl</i>	Hour after sunset	0.066	0.019	3.436	0.001	0.066	0.11	0.014
	Arthropod Abundance	0.003	0.005	0.548	0.584	0.003	0.018	
	Lux	0.008	0.006	1.296	0.195	0.008	0.041	
<i>Eumops perotis</i>	Hour after sunset	0.031	0.008	3.892	>0.001	0.031	0.122	0.053
	Arthropod Abundance	0.005	0.002	2.133	0.033	0.005	0.069	
	Lux	0.014	0.003	5.621	>0.001	0.014	0.176	
<i>Peropteryx macrotis</i>	Hour after sunset	-0.003	0.017	-0.2	0.841	-0.003	-0.006	0.002
	Arthropod Abundance	>-0.001	0.005	-0.062	0.951	>-0.001	-0.002	
	Lux	-0.007	0.005	-1.243	0.214	-0.007	-0.04	
<i>Peropteryx trinitatis</i>	Hour after sunset	-0.008	0.006	-1.24	0.215	-0.008	-0.04	0.002
	Arthropod Abundance	-0.001	0.002	-0.644	0.52	-0.001	-0.021	
	Lux	-0.001	0.002	-0.63	0.529	-0.001	-0.02	

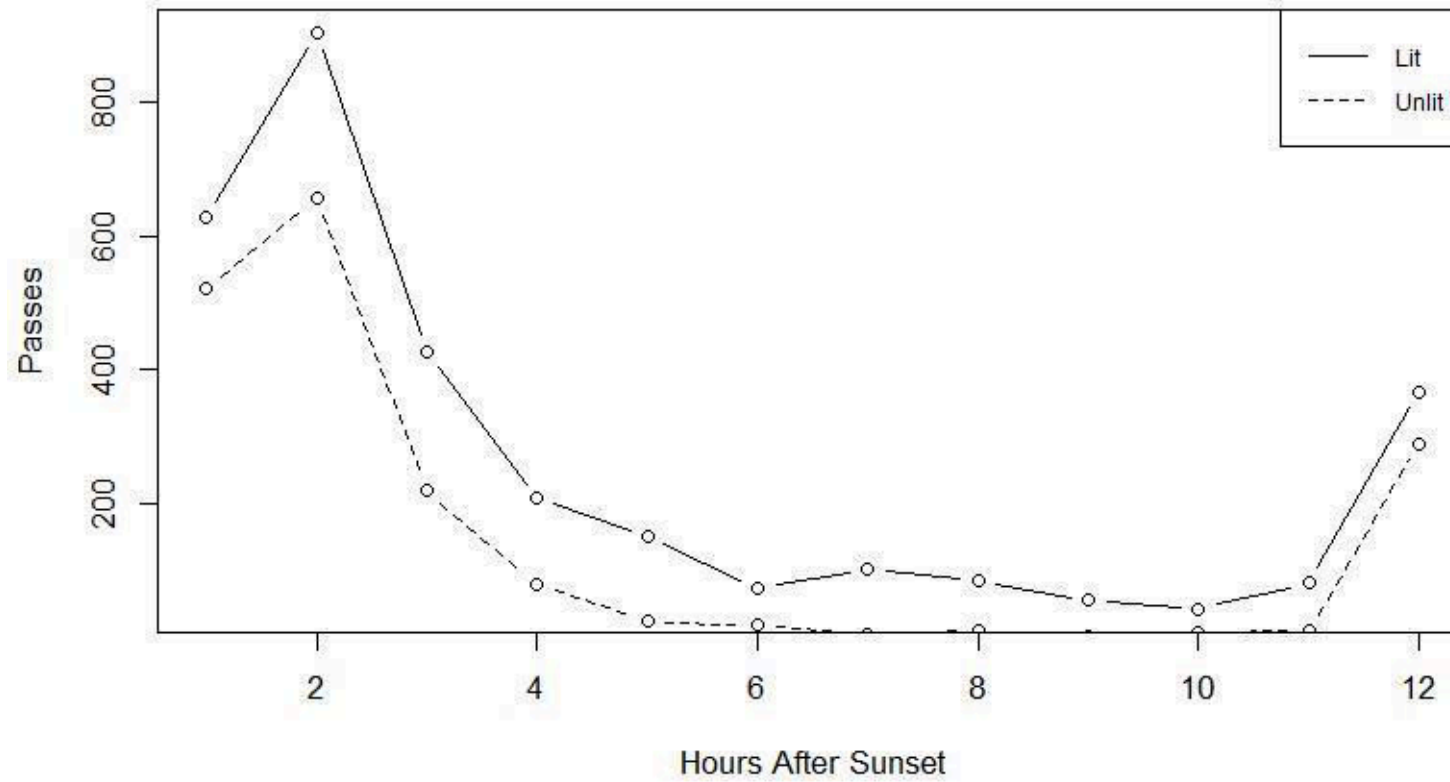


*Continue Table S5*

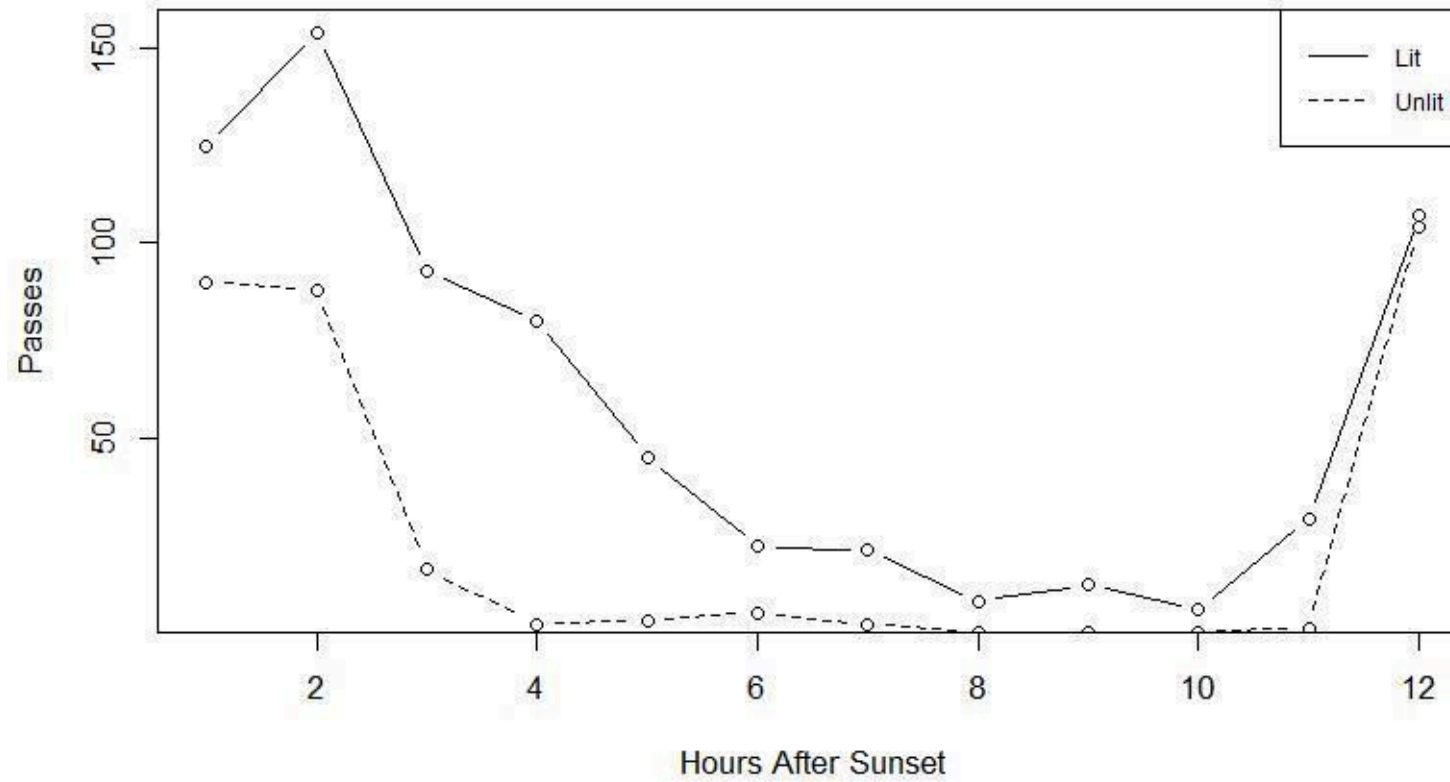
<b>Models</b>	<b>Predictor Variables</b>	<b>Estimate</b>	<b>Std.Err</b>	<b>z-value</b>	<b>P(&gt; z )</b>	<b>Std.lv</b>	<b>Std.all</b>	<b>R<sup>2</sup></b>
<i>Eptesicus furinalis</i>	Hour after sunset	-0.081	0.025	-3.254	0.001	-0.081	-0.104	0.021
	Arthropod Abundance	-0.004	0.007	-0.593	0.553	-0.004	-0.019	
	Lux	-0.024	0.008	-3.071	0.002	-0.024	-0.098	
<i>Eptesicus brasiliensis</i>	Hour after sunset	-0.125	0.038	-3.252	0.001	-0.125	-0.104	0.017
	Arthropod Abundance	-0.002	0.01	-0.236	0.813	-0.002	-0.008	
	Lux	-0.031	0.012	-2.516	0.012	-0.031	-0.08	



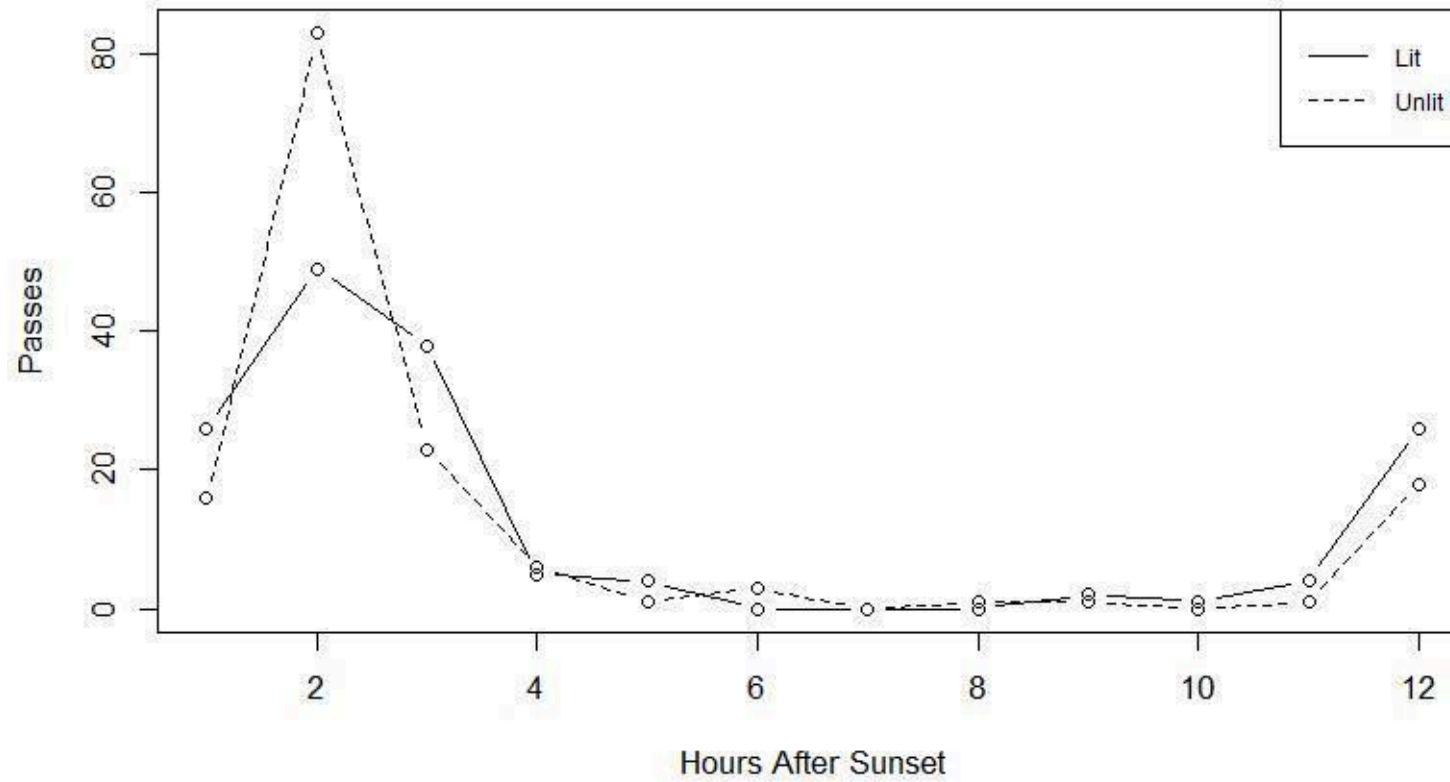
**Figure S1:** Line graph showing the all bat activity (passes) throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



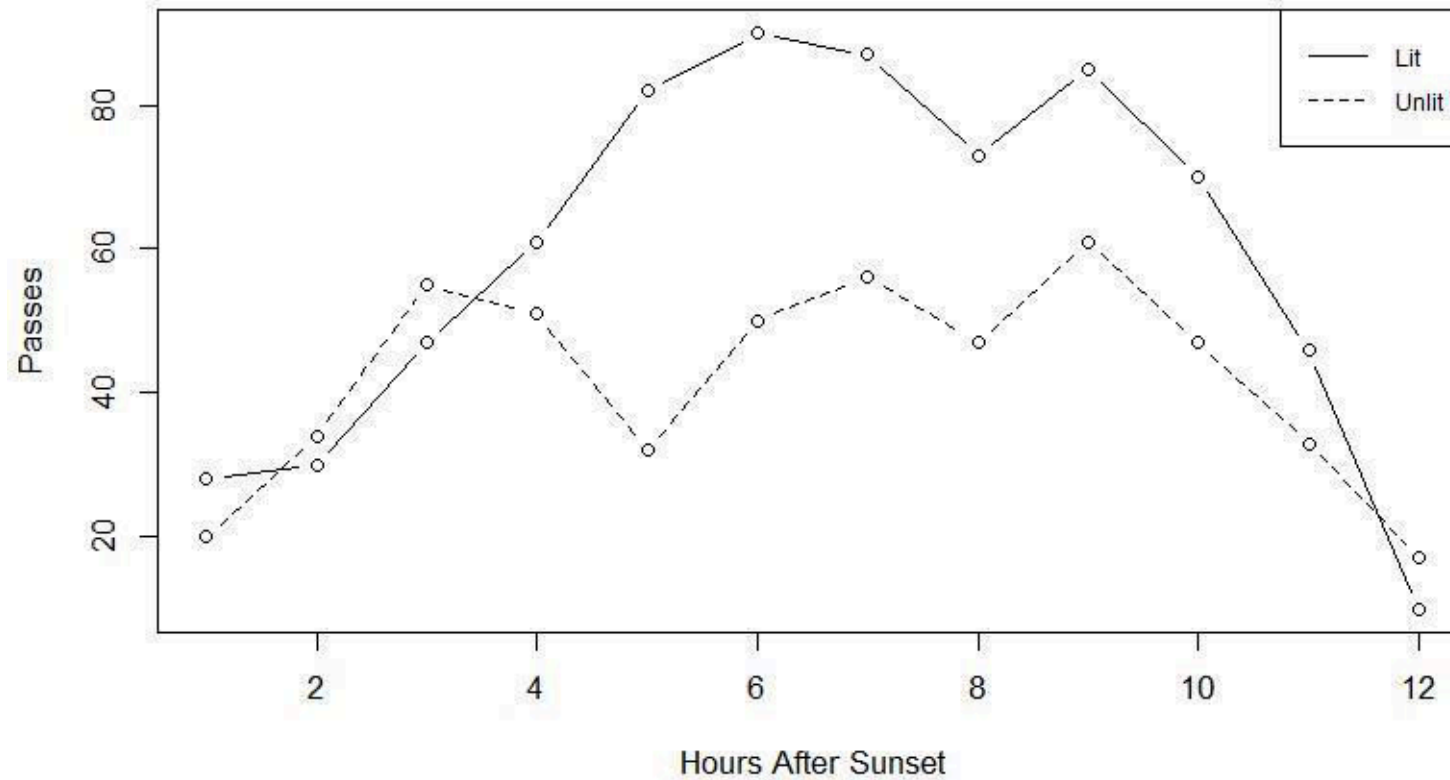
**Figure S2:** Line graph showing the activity (passes) of the species *M. molossus* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



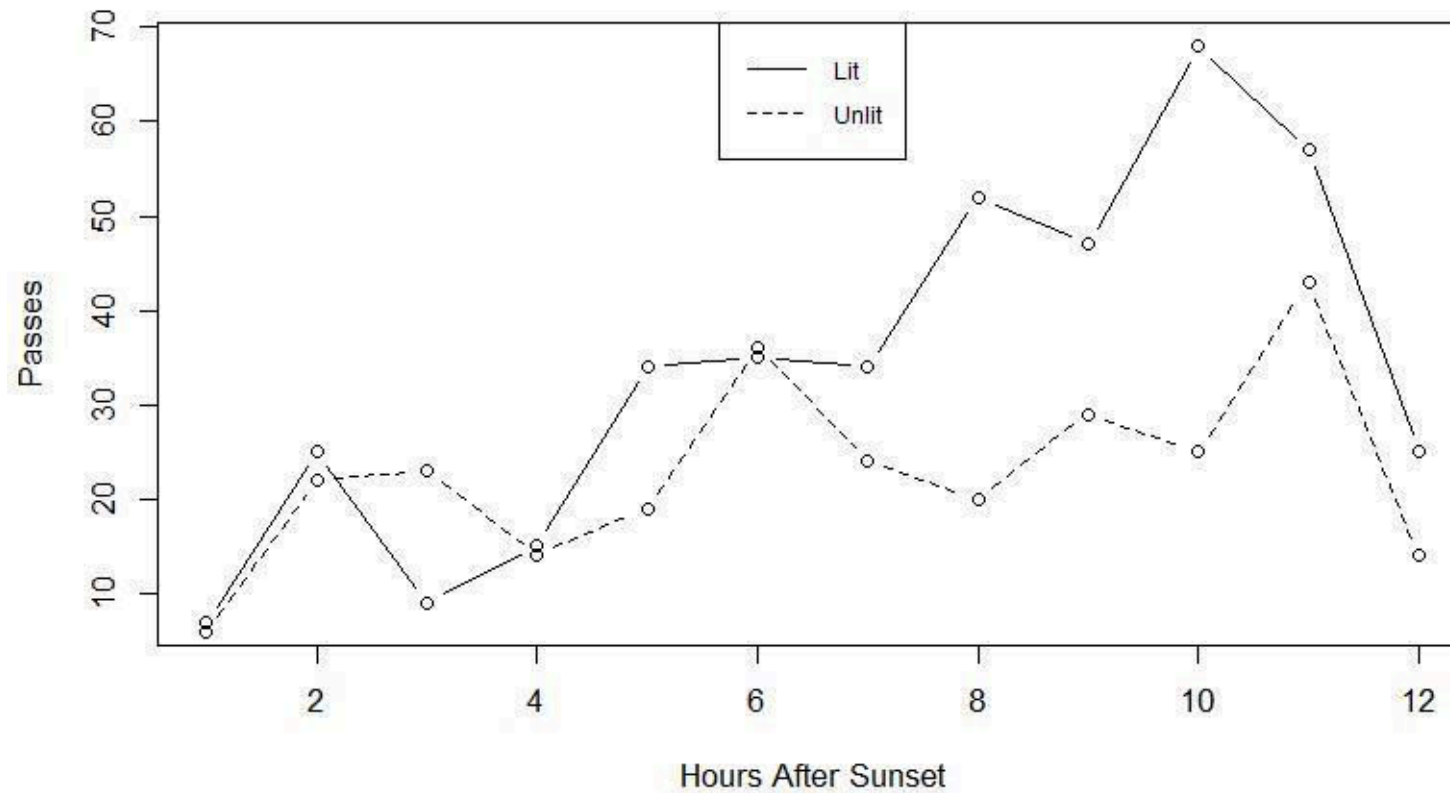
**Figure S3:** Line graph showing the activity (passes) of the species *M. currentium* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



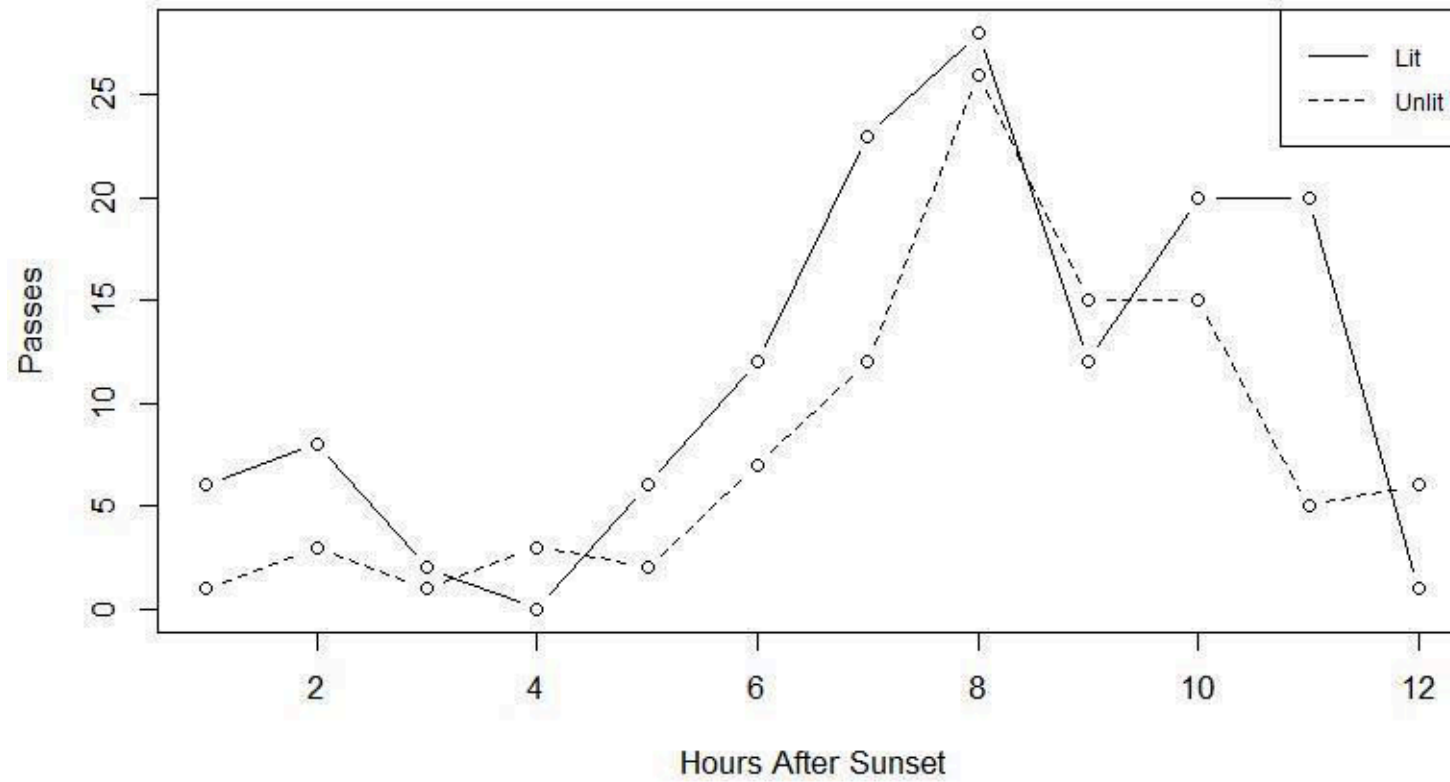
**Figure S4:** Line graph showing the activity (passes) of the species *Molossus. sp* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



**Figure S5:** Line graph showing the activity (passes) of the species *Eumops* sp/*Nyctinomops macrotis* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.

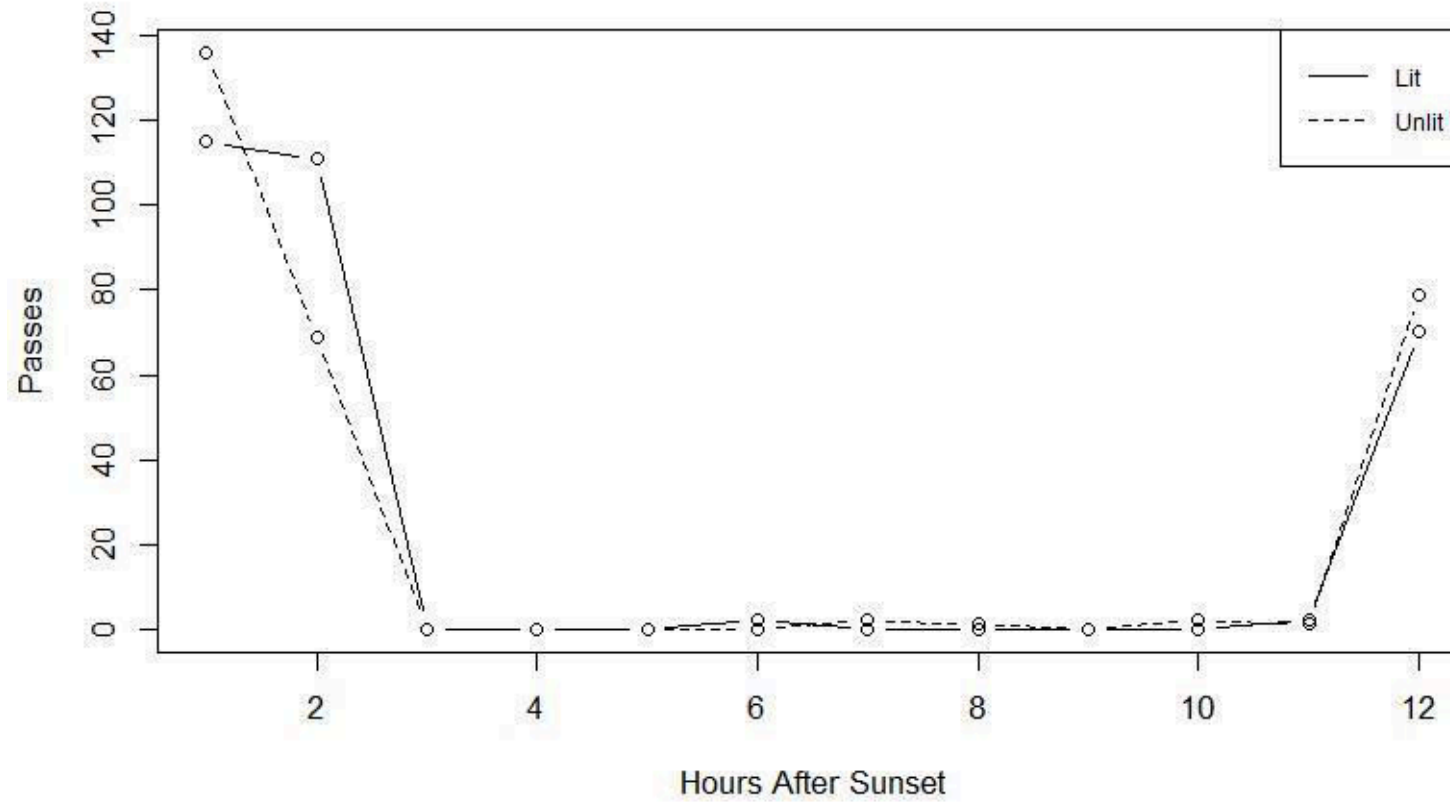


**Figure S6:** Line graph showing the activity (passes) of the species *Eumops*. spp throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.

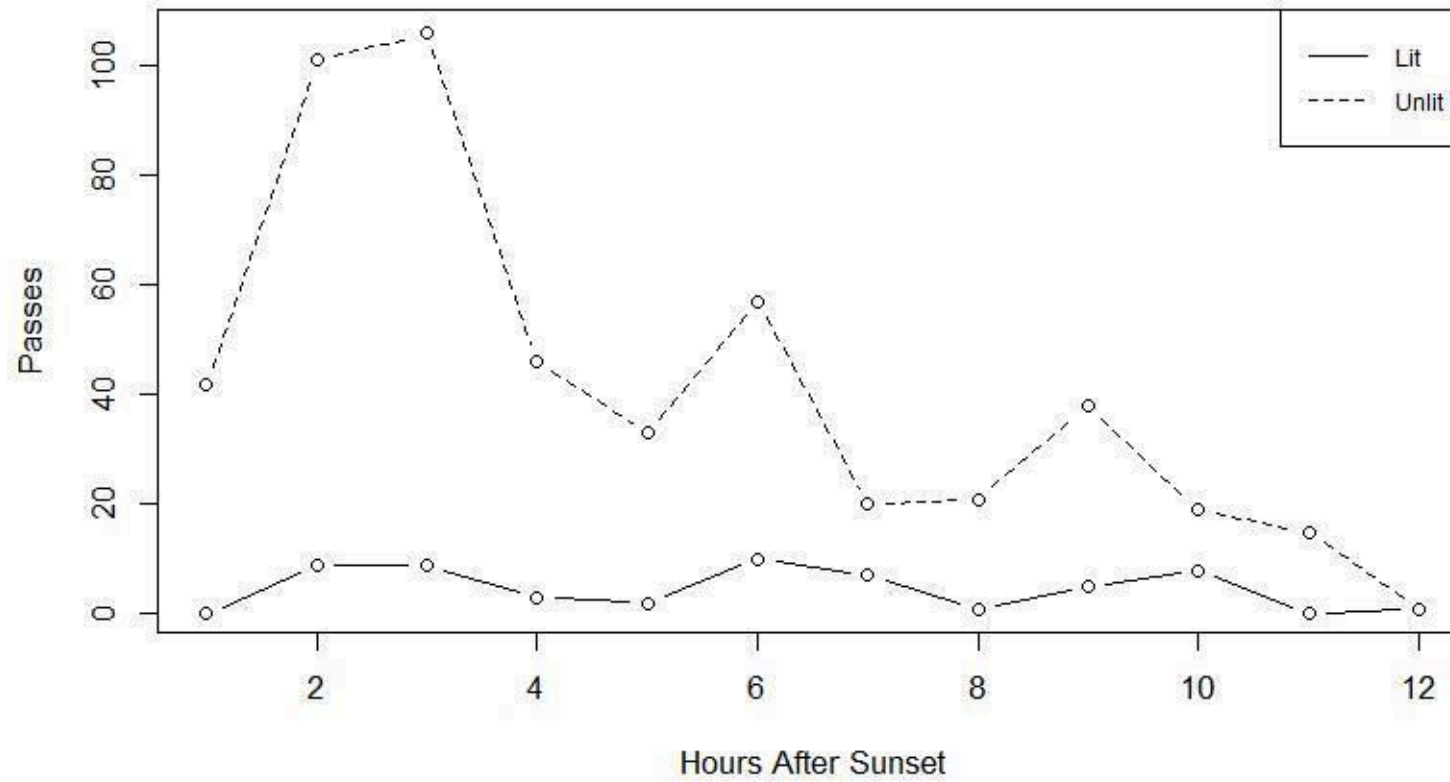


**Figure S7:** Line graph showing the activity (passes) of the species *E. perotis* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.

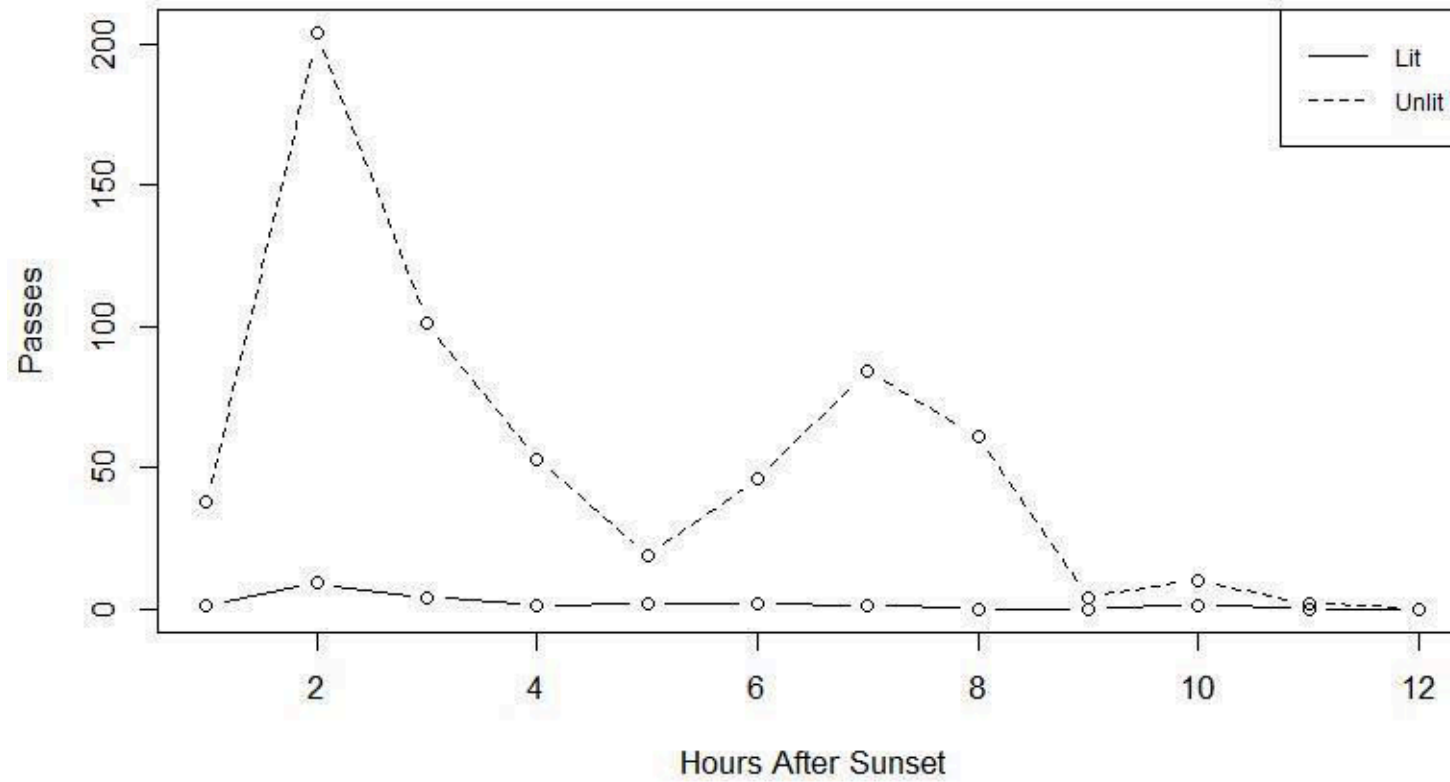




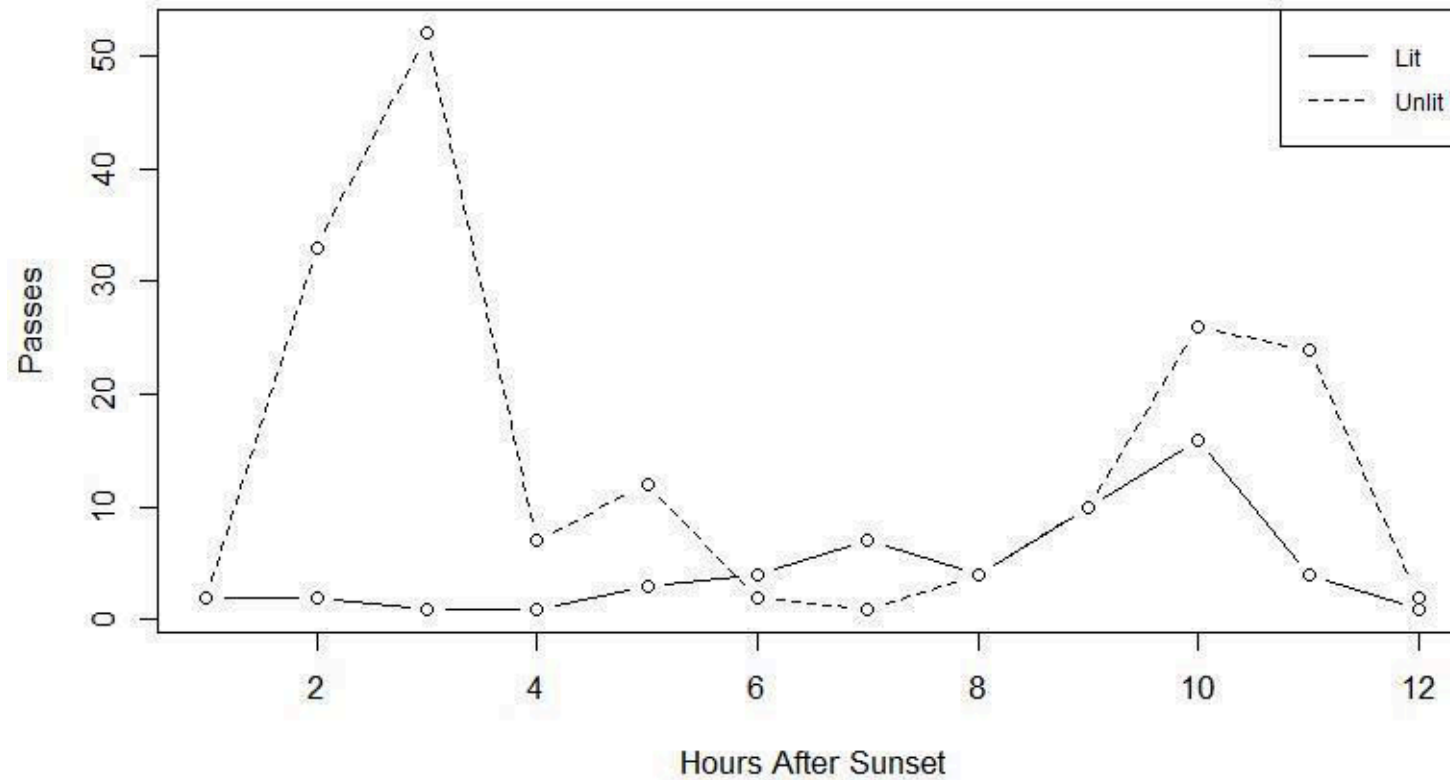
**Figure S8:** Line graph showing the activity (passes) of the species *Cynamops. sp* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



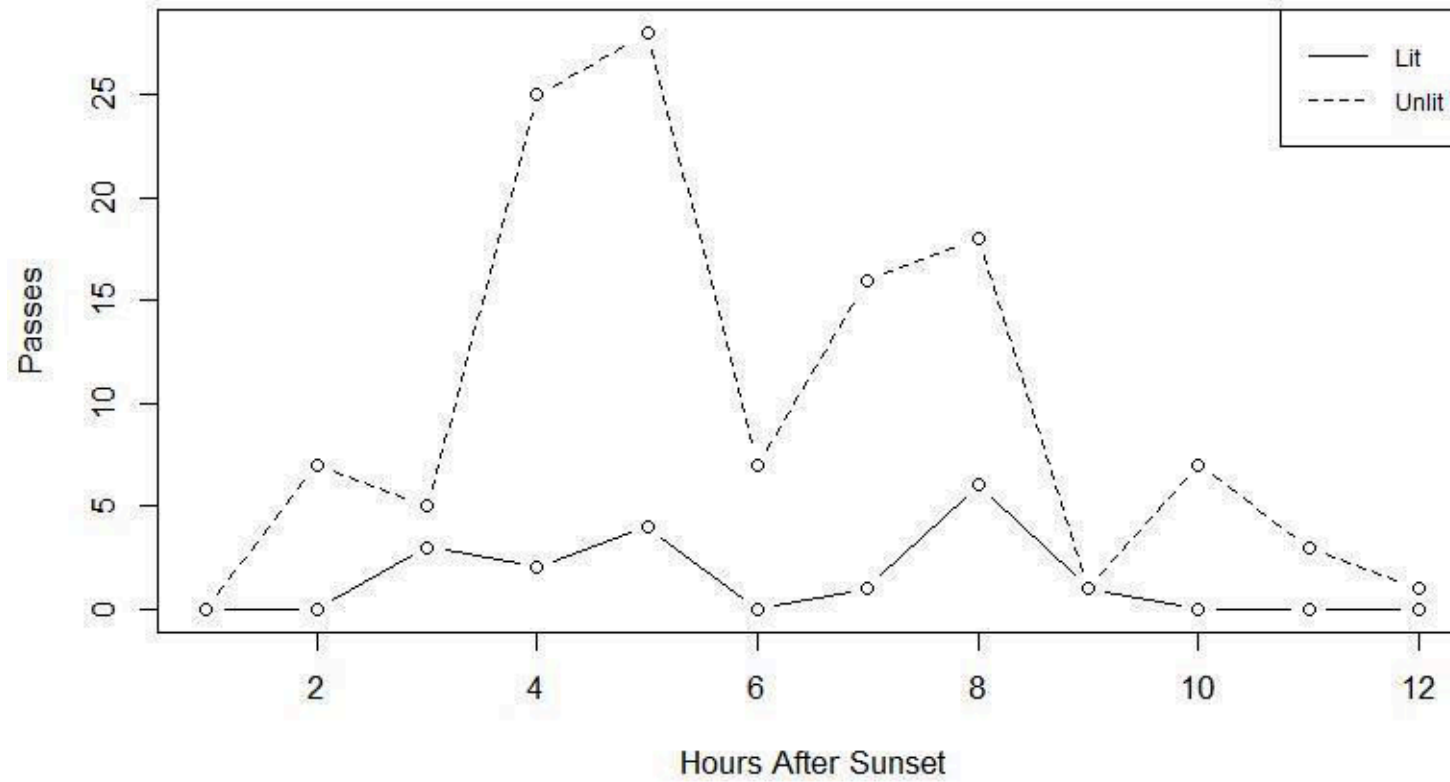
**Figure S9:** Line graph showing the activity (passes) of the species *E. furinalis* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



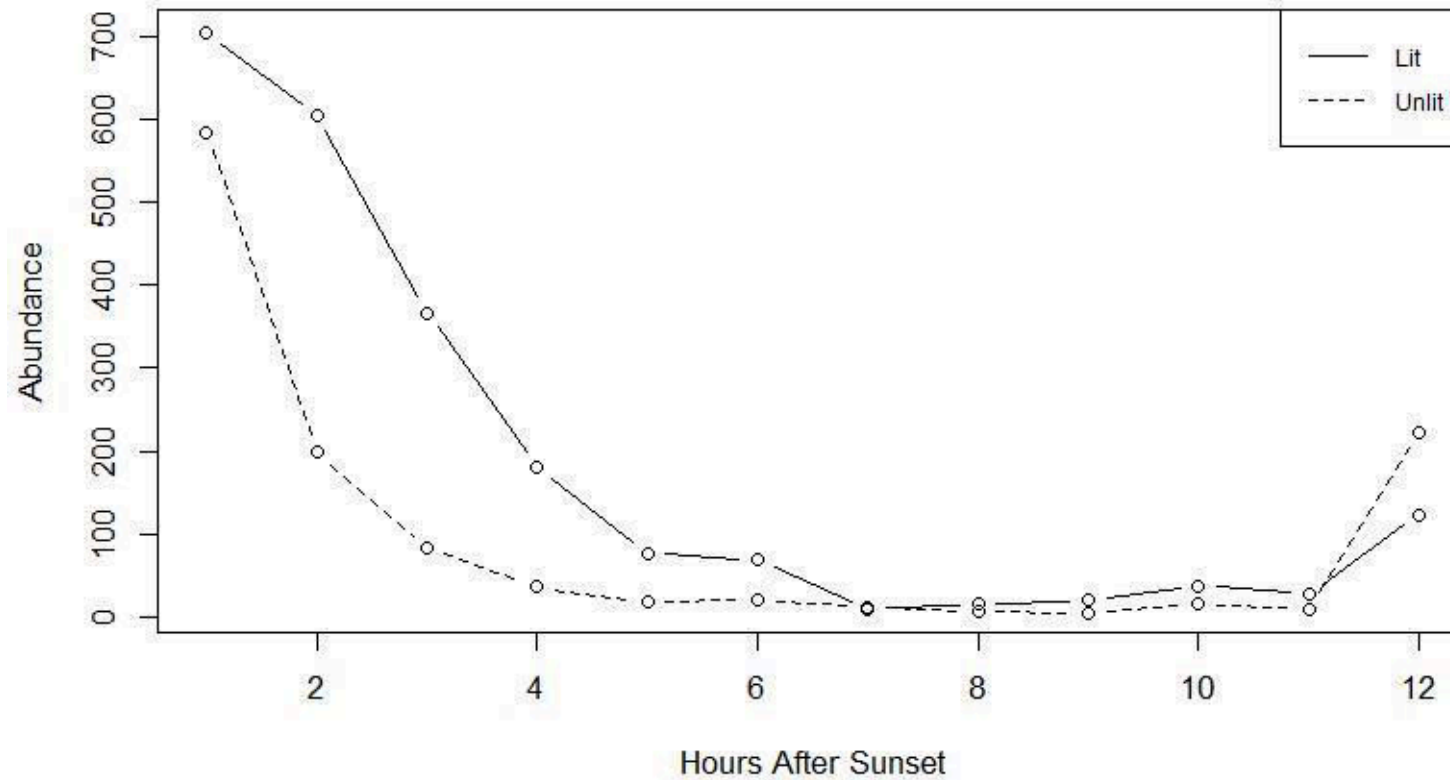
**Figure S10:** Line graph showing the activity (passes) of the species *E. brasiliensis* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



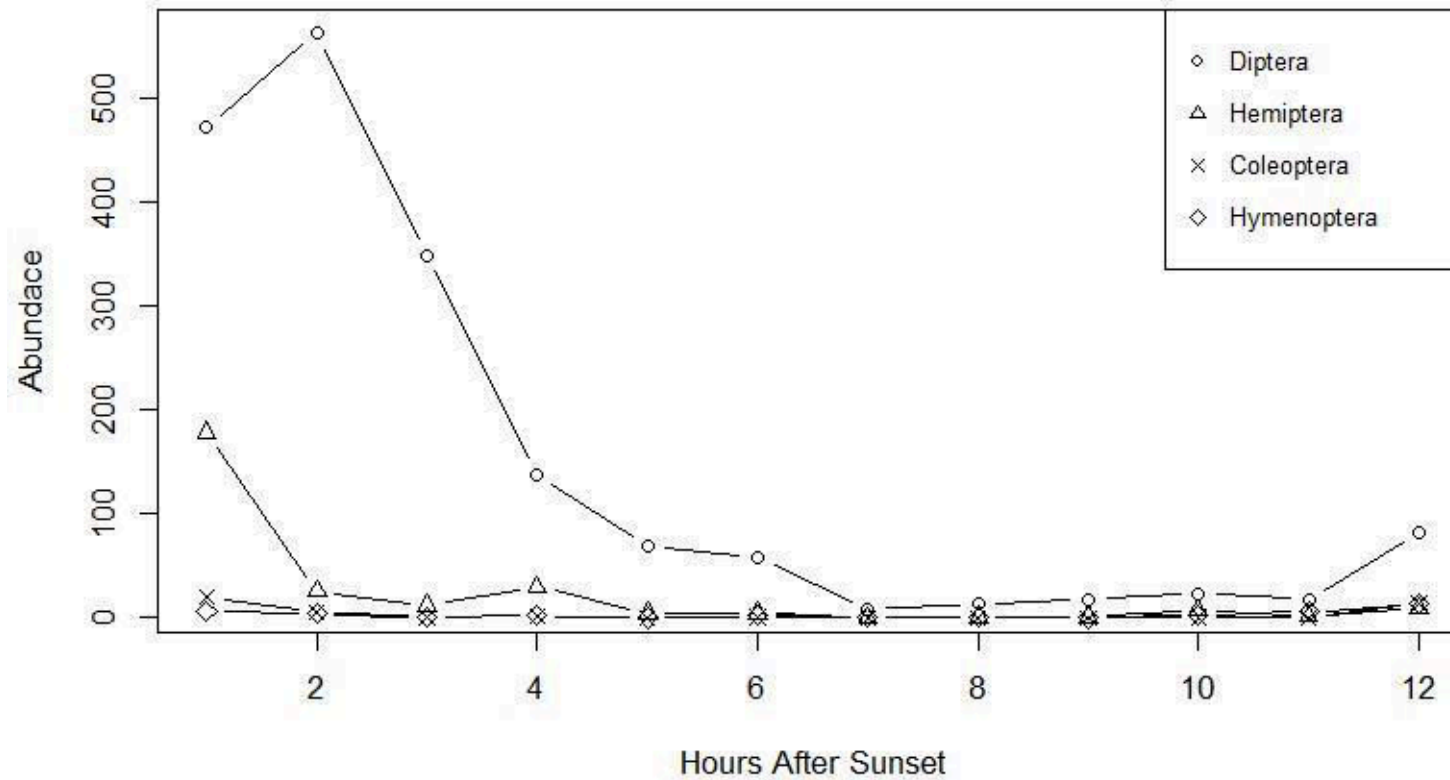
**Figure S11:** Line graph showing the activity (passes) of the species *P. macrotis* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



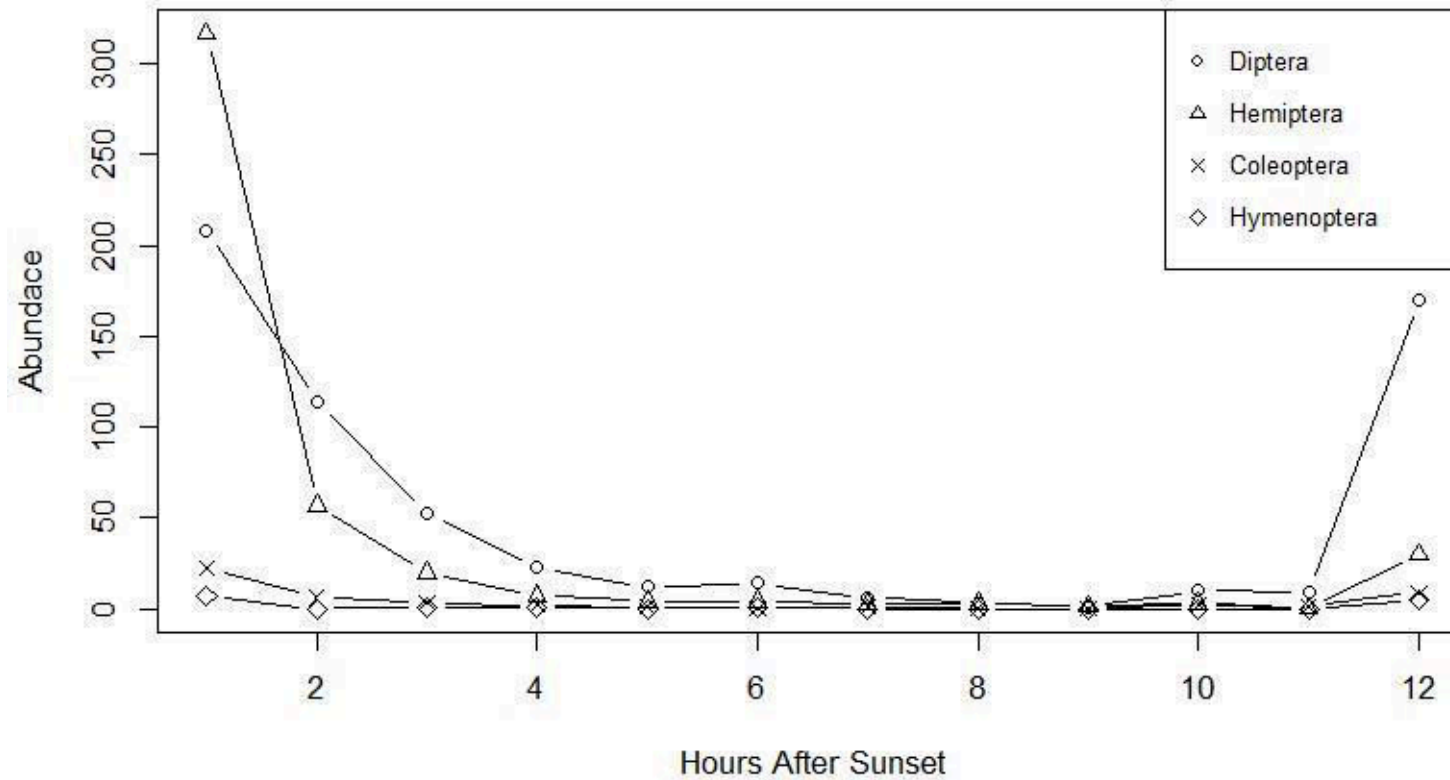
**Figure S12:** Line graph showing the activity (passes) of the species *P. trinitatis* throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



**Figure S13:** Line graph showing the abundance of the individuals of arthropods throughout the night (hours after sunset). Continuous line are the lit sites and dotted lines are the unlit sites.



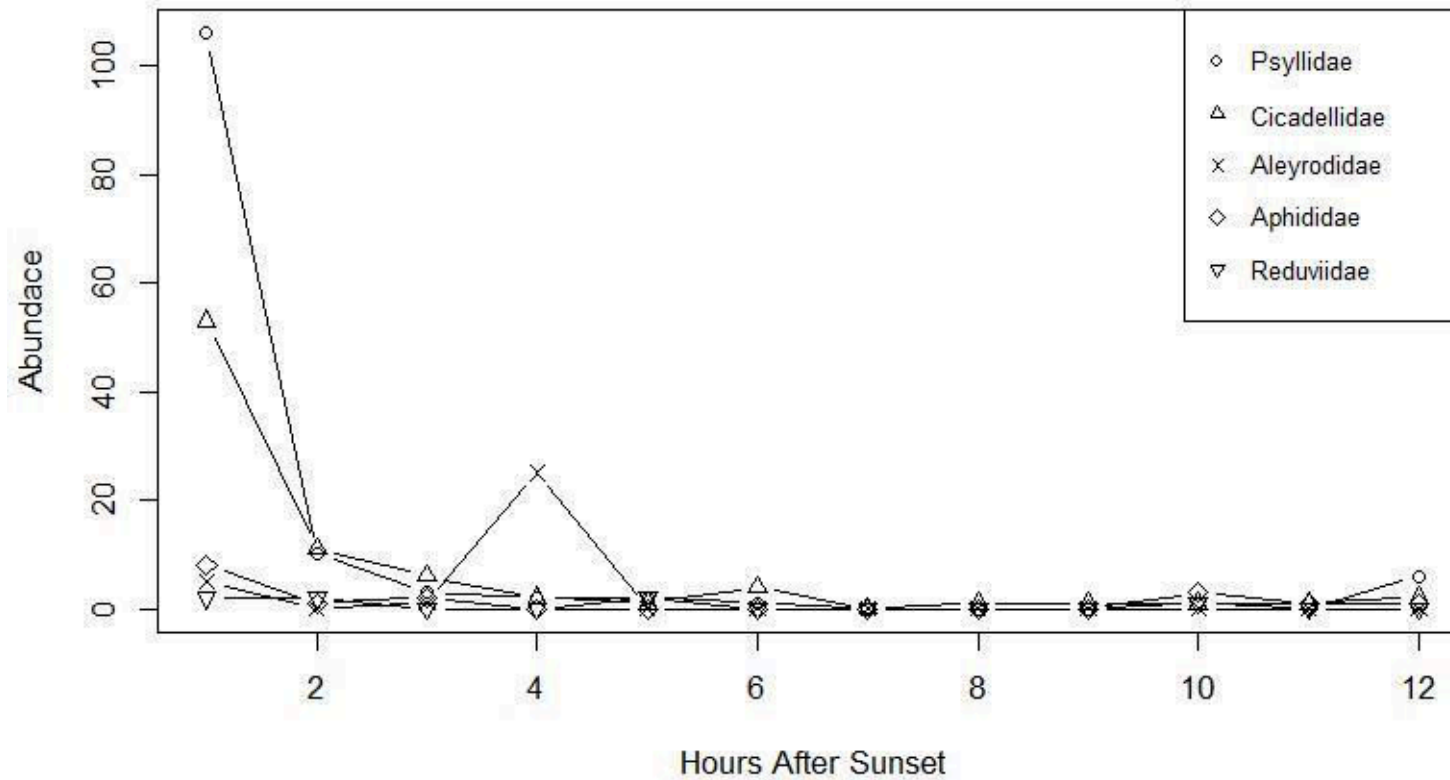
**Figure S14:** Line graph showing the abundance of the individuals of arthropods orders throughout the night (hours after sunset) in **lit sites**. Circles = Diptera order, Triangle = Hemiptera order, X = Coleoptera order and Rhombus = Hymenoptera.



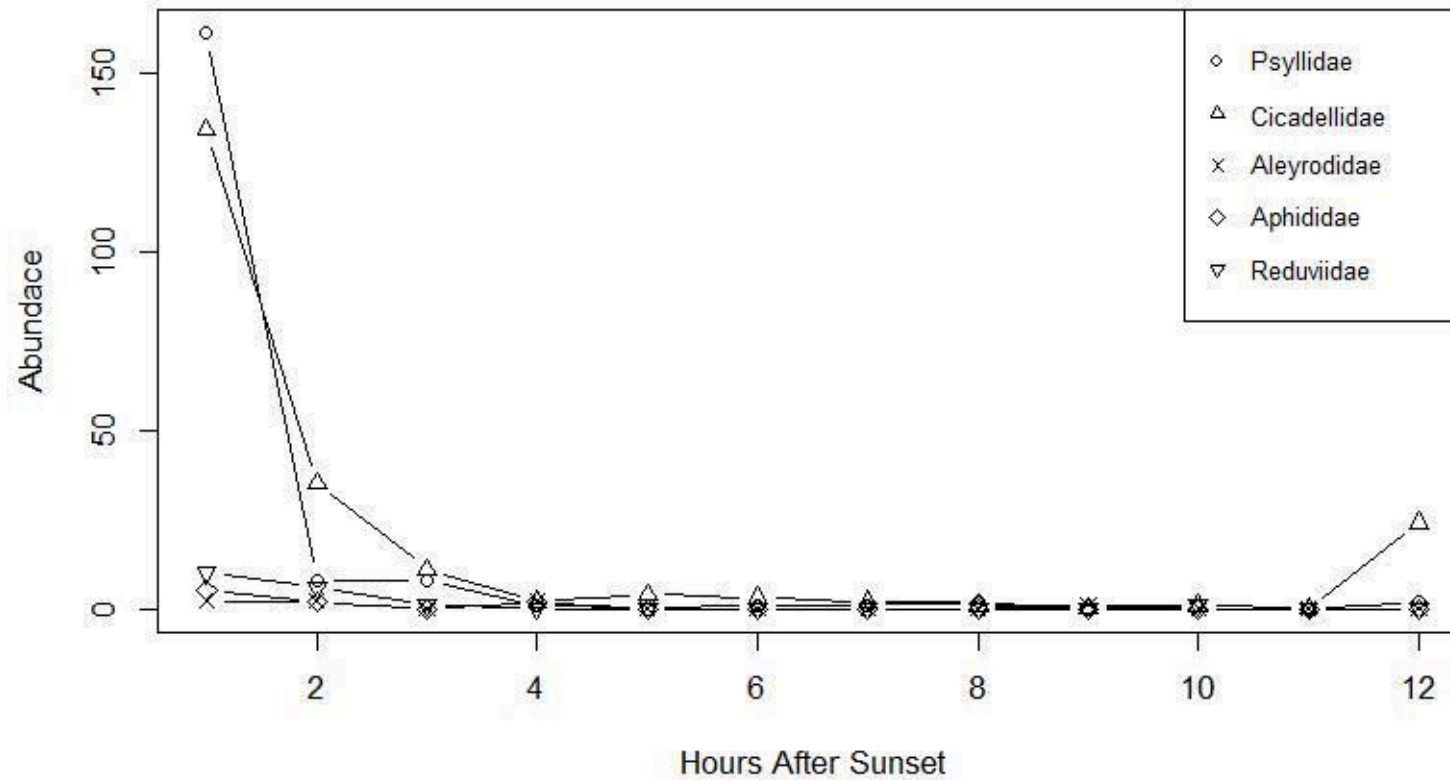
**Figure S15:** Line graph showing the abundance of the individuals of arthropods orders throughout the night (hours after sunset) in **unlit sites**.

Circles = Diptera order, Triangle = Hemiptera order, X = Coleoptera order and Rhombus = Hymenoptera.



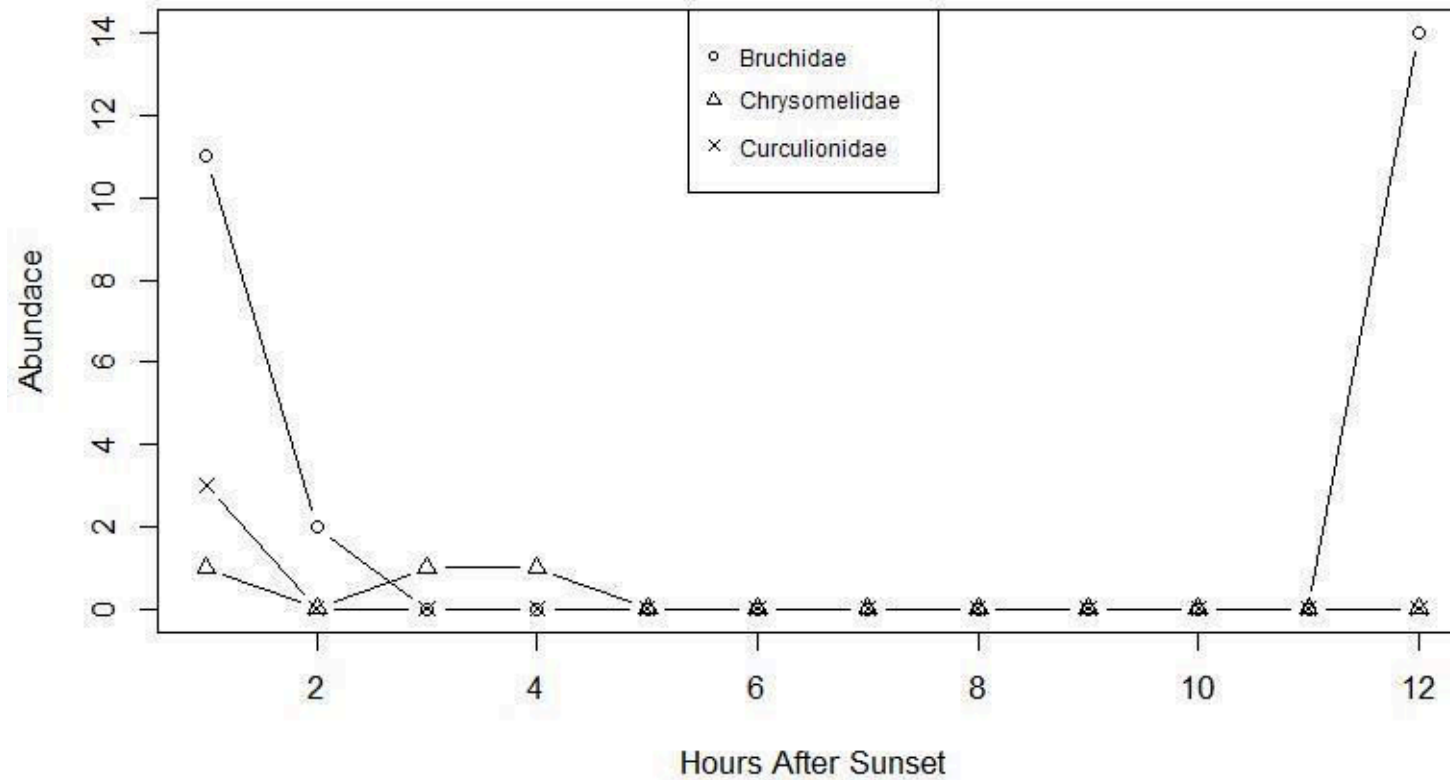


**Figure S16:** Line graph showing the abundance of the individuals of hemiptera families throughout the night (hours after sunset) in **lit sites**. Circles = Psyllidae, Triangle = Cicadellidae, X = Aleyrodidae, Rhombus = Aphididae and Reverse Triangle = Reduviidae.

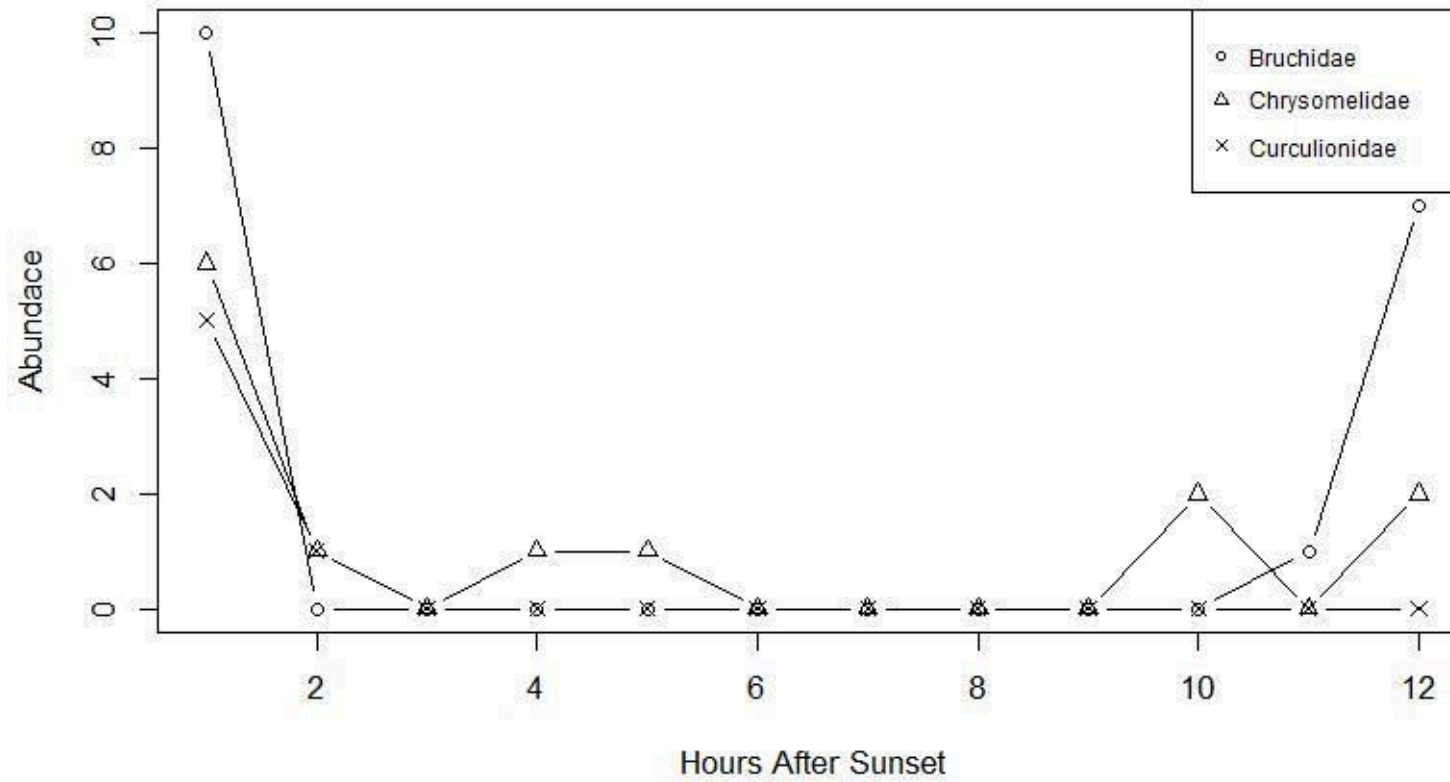


**Figure S17:** Line graph showing the abundance of the individuals of hemiptera families throughout the night (hours after sunset) in **unlit sites**.

Circles = Psyllidae, Triangle = Cicadellidae, X = Aleyrodidae, Rhombus = Aphididae and Reverse Triangle = Reduviidae.



**Figure S18:** Line graph showing the abundance of the individuals of coleoptera families throughout the night (hours after sunset) in **lit sites**. Circles = Bruchidae, Triangle = Chrysomelidae, X = Curculionidae.



**Figure S19:** Line graph showing the abundance of the individuals of coleoptera families throughout the night (hours after sunset) in **unlit sites**.

Circles = Bruchidae, Triangle = Chrysomelidae, X = Curculionidae.