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Detection and molecular characterization of piroplasms species from naturally infected dogs in southeast Brazil

Deteccão e caracterização molecular de piroplasmas em cães naturalmente infectados no Sudeste do Brasil

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Abstract

Rangelia vitalii is a protozoon described from dogs in the south and southeast regions of Brazil. It is phylogenetically related to *Babesia* spp. that infects dogs, but data on this enigmatic parasite is still limited. The aim of this work was to detect piroplasm species in dogs in the state of Rio de Janeiro, Brazil, by 18S rRNA gene-based PCR assay, restriction fragment length polymorphism (RFLP) and sequence analyses. Of 103 dogs examined, seven (6.8%) were positive for *Babesia* spp. by PCR. The amplified products were digested by restriction enzymes to differentiate the *Babesia* species, and one sample was identified as *Babesia vogeli*. The pattern observed for the other six amplification products did not match with pattern described for large *Babesia* infecting dogs. Sequencing analysis confirmed these six samples as *R. vitalii*, with high homologies (99-100%) with a sequence from south Brazil. This study confirms the presence of *Babesia vogeli* and *Rangelia vitalii* circulate in domestic dogs in Teresópolis, Rio de Janeiro, Brazil.

Keywords: *Babesia vogeli*, *Rangelia vitalii*, piroplasmas, dogs, PCR.

Resumo

Rangelia vitalii é um protozoário que infecta cães e foi descrito nas regiões Sul e Sudeste do Brasil. *R. vitalii* é filogeneticamente próxima à *Babesia* spp., mas dados deste misterioso parasito ainda são escassos. O objetivo deste trabalho foi detectar a presença de piroplasmas em cães naturalmente infectados no estado do Rio de Janeiro, através da amplificação do gene 18S rRNA pela PCR, clivagem com enzimas de restrição (RFLP) e caracterização genética através do sequenciamento. De 103 cães, sete (6,8%) foram positivos para *Babesia* spp. pela PCR. Os produtos amplificados foram digeridos por enzimas de restrição para a diferenciação das espécies de *Babesia* e uma amostra foi identificada como *Babesia vogeli*. O padrão de amplificação observado nas outras seis amostras não correspondeu ao padrão descrito para babesias que infectam cães. O sequenciamento das seis amostras confirmou ser uma espécie geneticamente idêntica a *R. vitalii* apresentando grande homologia (99-100%) com a sequência do sul do Brasil. Este estudo confirma a presença de *Babesia vogeli* e *Rangelia vitalii* infectando cães em Teresópolis, Rio de Janeiro, Brasil.

Palavras-chave: *Babesia vogeli*, *Rangelia vitalii*, piroplasmas, cães, PCR.

Introduction

Canine babesiosis is a parasitosis caused by the intraerythrocytic protozoa of the genus *Babesia* (LEVINE, 1973). Traditionally, *Babesia* classification in dogs is performed microscopically

based on the parasite morphology in the erythrocytes. Large *Babesia* are usually referred to as “*B. canis*”, whereas small *Babesia* as “*B. gibsoni*” (SOLANO-GALEGO et al., 2008). Recent advances in molecular techniques have enabled the detection and identification of piroplasmid species that infect dogs. *Babesia canis* is divided into three genetically distinct species: *B. canis*, *B. vogeli* and *B. rossi* (UILENBERG, 2006). A new species of large (as yet unnamed) *Babesia* has been described infecting

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dogs in North Carolina, USA (BIRKENHEUER et al., 2004; LEHTINEN et al., 2008). Nowadays, other small piroplasms are known to infect dogs including *Theileria annae* (ZÄHLER et al., 2000), *B. conradae* (CONRAD et al., 1991; KJEMTRUP; CONRAD, 2006), *T. annulata* (CRIADO et al., 2006), *T. equi* (CRIADO-FORNELIO et al., 2003) and *Theileria* sp. (MATJILA et al., 2008).

Rangelia vitalii is an intracellular parasite belonging to the phylum Apicomplexa and order Piroplasmida (LORETTI; BARROS, 2004). This protozoan infects dogs and is supposedly transmitted by ticks and causes a disease that is popularly known as “nambiuú” (bloody ears) or “plague of blood”. This illness has been described only in the south and southeast regions of Brazil (BRAGA, 1935; LORETTI; BARROS, 2004, 2005; DA SILVA et al., 2011). In a recent study conducted in southern Brazil *R. vitalii* was molecularly characterized (SOARES et al., 2011).

Rangelia vitalii is described as infecting mainly young dogs, but adult dogs can become infected. Dogs that live in both rural and urban areas may be affected. It is believed that the vectors of this agent are ticks of the species *Rhipicephalus sanguineus* and *Amblyomma aureolatum* (LORETTI; BARROS, 2004, 2005).

The life cycle of *R. vitalii* consists of an erythrocytic stage of development and exoerythrocytic form occurring in the cytoplasm of neutrophils, monocytes and endothelial cells (LORETTI; BARROS, 2005; SOARES et al., 2011). Morphologically, intraerythrocytic forms are very similar to *B. vogeli*, and pyriform to round shapes can be arranged in pairs on the same erythrocyte. Piroplasms are found in the blood circulation, mainly in the acute phase of infection. Parasites are numerous in histological sections of organs such as liver, spleen, kidneys, lungs and bone marrow. In these organs, the parasites are found in capillary endothelial cells and reticuloendothelial cells (CARINI; MACIEL, 1914; CARINI, 1948; LORETTI; BARROS, 2005).

Recent molecular studies characterized *B. vogeli* (PASSOS et al., 2005; SÁ et al., 2006), *B. gibsoni* (TRAPP et al., 2006) and *R. vitalii* (DA SILVA et al., 2011; SOARES et al., 2011) as the causative agents of canine piroplasmosis in Brazil. Studies on this disease are limited, especially those involving the molecular characterization of these piroplasmids in different regions of Brazil. Therefore, it is important to better characterize the piroplasm species infecting dogs in Brazil in order to improve the diagnosis and treatment of the diseases they cause.

The aim of this study was to detect and characterize piroplasm species in dogs from southeastern Brazil using both parasitological (buffy coat and capillary blood smear examination) and molecular methods (polymerase chain reaction (PCR), restriction fragment length polymorphism (RFLP) and sequence analyses).

Material and Methods

1. Samples and optic microscopy

Blood samples from 103 dogs examined in a private veterinary clinic in Teresópolis, Rio de Janeiro, were used in this experiment. The area (22° 24' 44" N and 42° 57' 56" W)

(<http://www.apolo11.com>) is located in the mountainous region of Rio de Janeiro, has a tropical latitude climate, with an annual average temperature of 20 °C (<http://www.teresopolis.rj.gov.br/>). The animals were selected with no preference for breed, age or sex during the period between November 2006 and May 2008.

Blood samples for DNA extraction were collected in tubes containing 10% ethylenediaminetetraacetic acid anticoagulant. Capillary blood smears were made at the moment of sample collection and were stained by quick-stain procedures (Instant-Prove®) for hemoparasite screening. The feathered edge and the lateral borders of the smears were examined.

2. Extraction, amplification, RFLP and DNA sequencing

DNA extraction was performed with 200 µL of total blood, using the Illustra Blood Genomicprep Kit (GE Health Care®) according to the manufacturer's instructions. From the DNA extracted from each sample, a fragment of the 18S rRNA was amplified with primers PIRO A (5'-AAT ACC CAA TCC TGA CAC AGG G-3') and PIRO B (5'-TTA AAT ACG AAT GCC CCC AAC-3'). The program consisted in an initial denaturation step at 94 °C for 5 minutes, followed by 30 denaturation cycles at 94 °C for 1 minute, annealing at 55 °C for 1 minute and extension at 72 °C for 1 minute. A final extension step at 72 °C for 5 minutes was used (CARRET et al., 1999).

To define the species of *Babesia*, the amplification products were digested with the enzymes *Hinf*I (Biolabs®, New England) and *Taq* I (Biolabs®, New England) for 3 hours (CARRET et al., 1999) and then analyzed by 2% agarose gel electrophoresis.

Only samples that showed different restriction patterns from piroplasm species described in the literature were sequenced from data of two previous amplifications of the 18S rRNA gene, as described by Soares et al. (2011) with modifications. The sequences were aligned using the program Mega version 5 (TAMURA et al., 2011) to generate the unique sequence of each sample. The first PCR was performed with primers Bab 33-57 (5'-GCC AGT AGT CAT ATG CTT GTC TTA A- 3') and Bab 432-409 (5'-TTC CTT AGA TGT GGT AGC CGT TTC- 3'), which amplify a fragment of ~370 bp. The program consisted of an initial denaturation step at 95 °C for 5 minutes, followed by 40 denaturation cycles at 95 °C for 40 seconds, annealing at 56 °C for 40 seconds and extension at 72 °C for 40 seconds. A final extension step at 72 °C for 5 minutes was performed. In the second amplification, the primers Bab 143-167 (5'-CCG TGC TAA TTG TAG GGC TAA TAC A- 3') and Bab 858-834 (5'-CCT CTG ACA GTT AAA TAC GAA TGC C- 3') were used, the latter being designed herein to amplify a fragment of ~715 bp. The reaction conditions were as follows: an initial denaturation step at 95 °C for 5 minutes, followed by 40 denaturation cycles at 95 °C for 1 minute, annealing at 64 °C for 1 minute and extension at 72 °C for 1 minute. A final extension step at 72 °C for 5 minutes was performed. For all amplifications, previously known positive *B. vogeli* samples and negative samples were used as controls.

The amplicons were purified with the commercial PureLink™ PCR Purification Kit (Invitrogen®, Carlsbad, CA, USA), according to the manufacturer's instruction. The purified DNA was sequenced by the automated sequencer ABI Prism 3130.

3. Phylogenetic analysis

The sequences obtained by amplifying the gene fragment encoding the 18S rRNA, when sequenced with primers BAB 33-57/BAB 432-409 and BAB 143-167/BAB 858-834, were compared with those available in the GenBank through the BLAST program. The genetic similarity tree was constructed using the algorithm Neighbor Joining of the program MEGA version 5 (TAMURA et al., 2011). For phylogenetic comparison, the following 18S rRNA gene sequences were included in the alignment: *Rangelia vitalii* (HQ150006.1), *B. motasi* (AY260180.1), *B. ovata* (AY603400.1), *B. gibsoni* (EU084677.1), *B. canis vogeli* (AB083374.1), *B. canis canis* (AY072926.1), *B. canis rossii* (DQ111760.1), *Theileria annae* (AF188001.1), *B. bigemina* (X59607.1), *B. caballi* (Z15104.1), *B. odocoilei* (AY237638.1), *B. divergens* (U07885.1), *B. microti* (U09833.1), *T. equi* (Z15105.1), *T. parva* (L02366.1), *T. annulata* (EU083801.1), *Cytauxzoon felis* (GU903911.1) and *Plasmodium falciparum* (M19172.1).

Results

1. Optic microscopy, PCR assay and RFLP

In the capillary and circulating blood smears, four (3.9%) of the 103 dogs evaluated showed intraerythrocytic parasites compatible with *Babesia* spp.

In all samples that were positive by light microscopy (samples 1, 3, 5 and 7) and three others (samples 2, 4 and 6), the 18S rRNA gene fragment of *Babesia* of approximately 400 bp was amplified. All samples that tested positive by PCR (6.8%) were submitted to RFLP analysis. Using the enzyme *Taq* I, only sample 1 was cleaved into three fragments of different sizes. With the exception of sample 1, the other six samples were cleaved with the enzyme *Hinf* I into two fragments of approximately 200 bp, which were not separated by electrophoresis due to their similar size (Figure 1). The pattern of cleavage of sample 1 is consistent with the subspecies *B. vogeli* (CARRET et al., 1999).

Amplifications using the primer pairs BAB 33-57/BAB 432-409 and BAB 143-167/BAB 858-834 produced products of ~370 and 715 bp, respectively.

2. DNA sequencing

Sample 1 (GenBank access number GQ214514.1), when sequenced with primers PIRO A and PIRO B, shared 100% identity with *B. vogeli* from Venezuela (GenBank access number DQ297390.1), Japan (GenBank access number AB083374.1) and Brazil (GenBank access number AY371196.1).

Samples 2 through 7, when sequenced with primers BAB 33-57/BAB 432-409 and BAB 143-167/BAB 858-834, were aligned using the program Mega version 5 (TAMURA et al., 2011) to generate the unique sequence of each sample. The final sequences were deposited into GenBank (GenBank access numbers JN880428, JN880429, JN880430, JN880431, JN880432 and JN880433). Sequences 2 through 6 and sequence 7 shared 99%

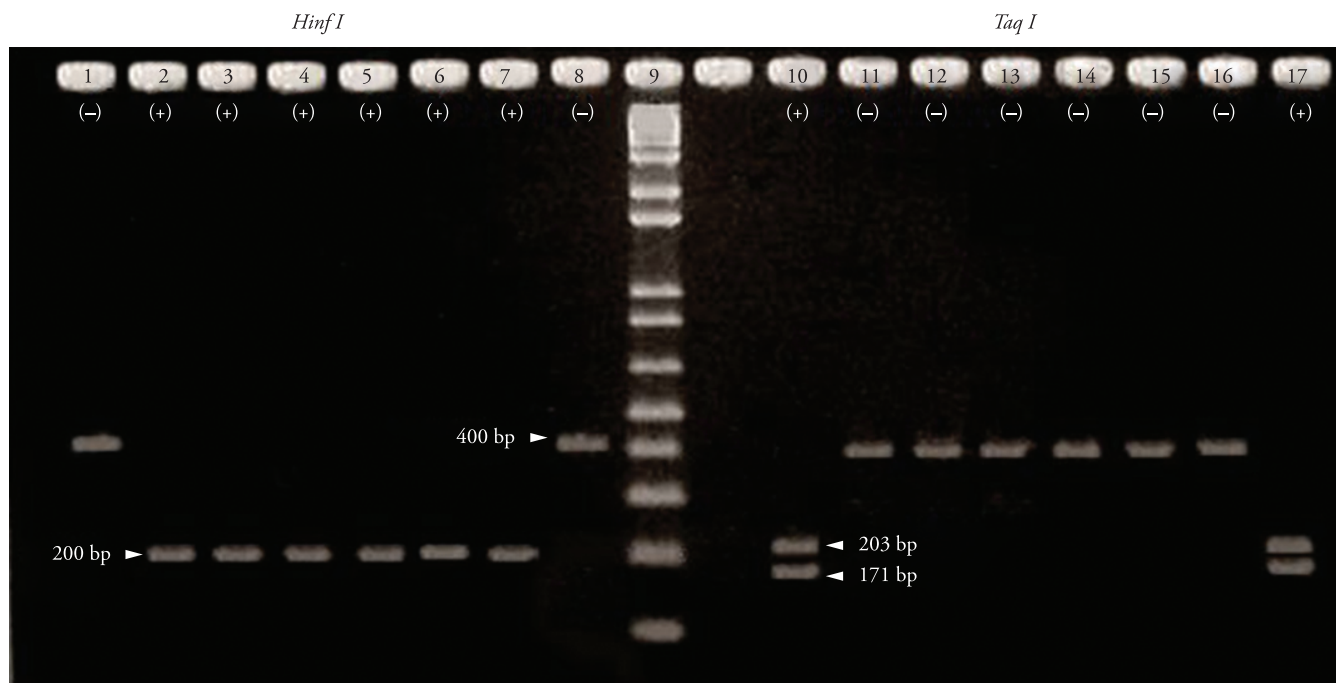


Figure 1. Analysis of PCR-RFLP products (*Hinf*I and *Taq*I enzymes) by 2% agarose gel electrophoresis and ethidium bromide staining. From left to right the lanes contain the following: cleavage with the *Hinf*I - *Babesia vogeli* positive sample (lane 1); *Babesia* spp. positive samples (lanes 2 to 7); *B. vogeli* positive control (lane 8); cleavage with the *Taq*I - *B. vogeli* positive sample (lane 10); *Babesia* spp. positive samples (lanes 11 to 16); *B. vogeli* positive control (lane 17); molecular weight marker (lane 9).

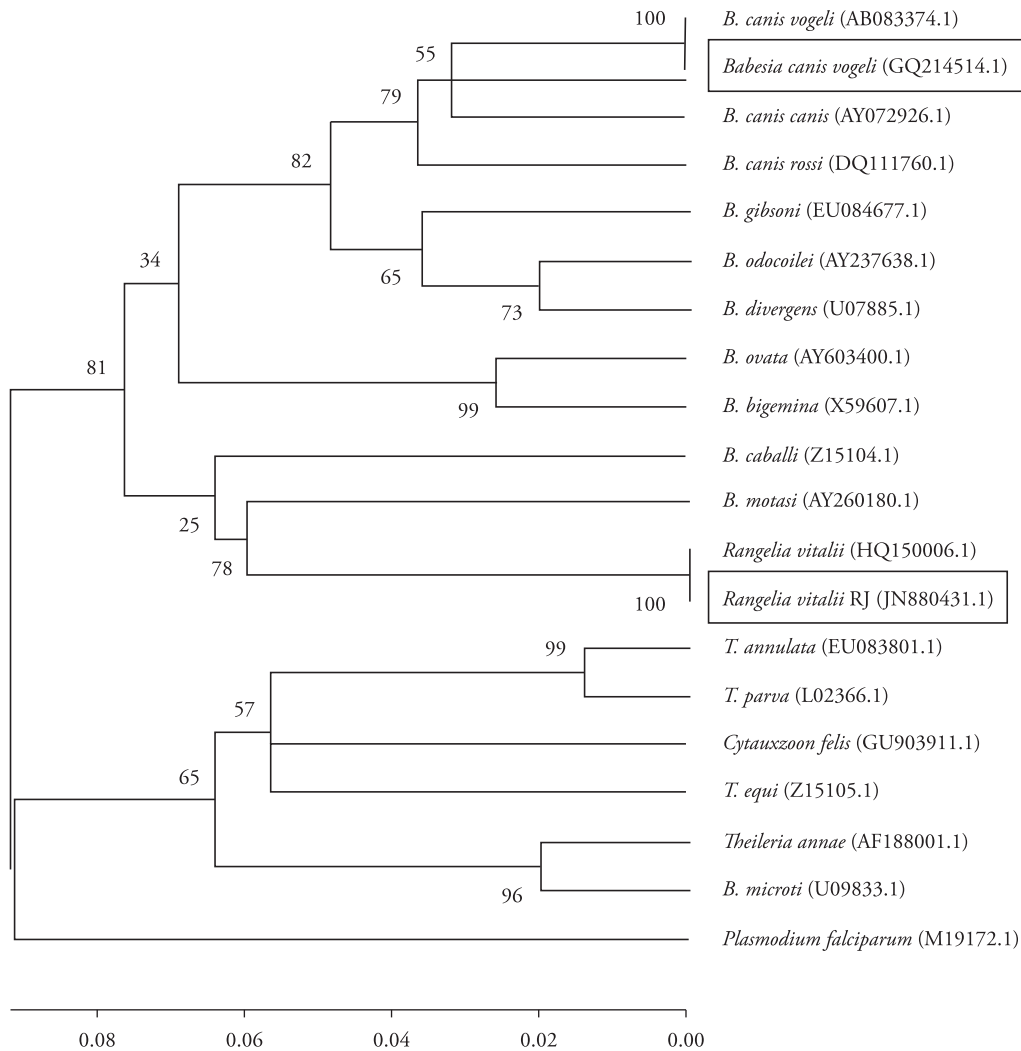


Figure 2. Neighbor-joining phylogenetic tree based on the partial 18S rRNA gene sequences of *Babesia vogeli* (311 bp) and *Rangelia vitalii* (796 bp) detected in Rio de Janeiro and similar sequences from GenBank. Bootstrap values were obtained after 1000 resampling. New sequences are marked by rectangles. The outgroup used in this work was *Plasmodium falciparum* (M19172.1).

and 100% identity, respectively, with *R. vitalii* from Santa Maria, Brazil (GenBank access number HQ150006.1).

The phylogenetic tree demonstrated that samples 2 through 7 and the *R. vitalii* sample from Santa Maria form a new branch of piroplasmids (Figure 2), a conclusion that is supported by the high bootstrap value of this phylogenetic analysis.

Discussion

Molecular studies have revealed a vast genetic diversity of piroplasmids infecting dogs (CRIADO-FORNELIO et al., 2003). PCR amplification of 18S rRNA partial sequences allows the identification of different *Babesia* species that were previously indistinguishable due to the similarity in the parasites' morphology (BIRKENHEUER et al., 2004; DUH et al., 2004; MATJILA et al., 2004; SÁ et al., 2006).

Passos et al. (2005) provided the first molecular characterization of *B. vogeli* in Brazil. These authors used blood samples from dogs

from the states of São Paulo and Minas Gerais. PCR amplification and sequencing of the 18S rRNA showed 100% similarity between the Brazilian isolates, which were identical to an isolate from Japan (AB083374.1) deposited into GenBank. Sá et al. (2006) performed the first detection of *B. vogeli* in Rio de Janeiro. They used the same protocols for PCR-RFLP as described by Carret et al. (1999).

In our study, PCR-RFLP was found to be a fast and practical method. DNA cleavage of other six samples by the enzyme *Hinf*I produced fragments that did not match any restriction pattern described in literature, suggesting that these samples were another piroplasmid species (CARRET et al., 1999; SOLANO-GALLEGO et al., 2008).

Recent advances in molecular techniques have enabled the detection and identification of piroplasmid species with high sensitivity and specificity when compared to other diagnostic methods (FÖLDVÁRI et al., 2005). Currently, new piroplasmid species have been reported as infecting dogs, including *Theileria annae* (ZÄHLER et al., 2000), *B. conradae* (CONRAD et al., 1991; KJEMTRUP et al., 2006), *T. annulata* (CRIADO et al., 2006),

T. equi (CRIADO-FORNELIO et al., 2003), *Theileria* sp. (MATJILA et al., 2008) and a yet undescribed species of large *Babesia* identified in North Carolina, USA (BIRKENHEUER et al., 2004; LEHTINEN et al., 2008).

Rangelia vitalii, which has been described in the south and southeast regions of Brazil, was shown to be phylogenetically closely related to a *Babesia* spp. *sensu stricto* group of piroplasmas that infects dogs (SOARES et al., 2011). This parasite was first described morphologically in the State of São Paulo (PESTANA, 1910a, b) and subsequently by immunohistochemistry in the State of Rio Grande do Sul (LORETTI; BARROS, 2005). Morphologically, intraerythrocytic forms of *R. vitalii* closely resemble *B. vogeli*. Piroplasms are often scarce in the bloodstream, but might be more frequent in the acute phase of infection (CARINI; MACIEL, 1914; CARINI, 1948; LORETTI; BARROS, 2005; DA SILVA et al., 2011). The present study demonstrated that 6.8% of the samples were positive by PCR, whereas 3.9% were positive by morphological examination. Thus, PCR can be considered to be a more sensitive assay.

The sequence analysis of the six samples in our study shows that they were genetically similar to *R. vitalii* from Santa Maria (SOARES et al., 2011). The phylogenetic tree demonstrated that the six *R. vitalii* samples from this study and the *R. vitalii* from Santa Maria form a new branch of piroplasms, a conclusion that is supported by the high bootstrap value.

In summary, PCR-RFLP and sequencing revealed the presence of *R. vitalli* in Rio de Janeiro, which suggests that the geographic distribution of this parasite in Brazil is probably larger than currently known.

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References

- Birkenheuer AJ, Neel J, Ruslander D, Levy MG, Breitschwerdt EB. Detection and molecular characterization of a novel large *Babesia* species in a dog. *Vet Parasitol* 2004; 124(3-4): 151-160. PMID:15381295. <http://dx.doi.org/10.1016/j.vetpar.2004.07.008>
- Braga A. Contribuição ao estudo experimental das piroplasmoses dos cães. *Bol Vet Exército* 1935; 3:1-16.
- Carini A, Maciel J. Sobre a moléstia dos cães, chamada nambiuvú, e o seu parasita (*Rangelia vitalli*). *An Paul Med Cir* 1914; 3: 65-71.
- Carini A. Sobre o ciclo de desenvolvimento exo-eritrocitário de um piroplasma do cão. *Arq Biol* 1948; 285: 49-52.
- Carret C, Walas F, Carcy B, Grande N, Précigout E, Moubri K, et al. *Babesia canis canis*, *Babesia canis vogeli*, *Babesia canis rossii*: differentiation of the three subspecies by a Restriction Fragment Length Polymorphism analysis on amplified small subunit ribosomal RNA genes. *J Eukaryot Microbiol* 1999; 46(3): 298-303. PMID:10377990. <http://dx.doi.org/10.1111/j.1550-7408.1999.tb05128.x>
- Conrad P, Thomford J, Yamane I, Whiting J, Bosma L, Uno T, et al. Hemolytic anemia caused by *Babesia gibsoni* infection in dogs. *J Am Vet Med Assoc* 1991; 199(5): 601-605. PMID:1787120.
- Criado A, Martinez J, Buling A, Barba JC, Merino S, Jefferies R, et al. New data on epizootiology and genetics of piroplasms based on sequences of small ribosomal subunit and cytochrome b genes. *Vet Parasitol* 2006; 142(3-4): 238-247. PMID:16919391. <http://dx.doi.org/10.1016/j.vetpar.2006.07.004>
- Criado-Fornelio A, Martinez-Marcos A, Buling-Saraña A, Barba-Carretro, A. Molecular studies on *Babesia*, *Theileria* and *Hepatozoon* in Southern Europe: Part I. Epizootiological aspects. *Vet Parasitol* 2003; 113(3-4): 189-201. [http://dx.doi.org/10.1016/S0304-4017\(03\)00078-5](http://dx.doi.org/10.1016/S0304-4017(03)00078-5)
- Da Silva AS, França RT, Costa MM, Paim CB, Paim FC, Dornelles GL, et al. Experimental infection with *Rangelia vitalii* in dogs: acute phase, parasitemia, biological cycle, clinical-pathological aspects and treatment. *Exp Parasitol* 2011; 128(4): 347-352. PMID:21570966. <http://dx.doi.org/10.1016/j.exppara.2011.04.010>
- Duh D, Tozon N, Petrovec M, Strasek K, Avsic-Zupanc T. Canine babesiosis in Slovenia: molecular evidence of *Babesia canis canis* and *Babesia canis vogeli*. *Vet Res* 2004; 35: 363-368. PMID:15210084. <http://dx.doi.org/10.1051/vetres:2004018>
- Földvári G, Hell E, Farkas R. *Babesia canis canis* in dogs from Hungary: detection by PCR and sequencing. *Vet Parasitol* 2005; 127(3-4): 221-226. PMID:15710522. <http://dx.doi.org/10.1016/j.vetpar.2004.10.016>
- Kjemtrup AM, Conrad PA. A review of the small canine piroplasms from California: *Babesia conradae* in the literature. *Vet Parasitol* 2006; 138(1-2): 112-117. PMID:16522352. <http://dx.doi.org/10.1016/j.vetpar.2006.01.045>
- Kjemtrup AM, Wainwright K, Miller M, Penzhorn BL, Carreno RA. *Babesia conradae*, sp. Nov., a small canine *Babesia* identified in California. *Vet Parasitol* 2006; 138(1-2): 103-111. PMID:16524663. <http://dx.doi.org/10.1016/j.vetpar.2006.01.044>
- Lehtinen LE, Birkenheuer AJ, Droleskey RE, Holman PJ. In vitro cultivation of a newly recognized *Babesia* sp. in dogs in North Carolina. *Vet Parasitol* 2008; 151(2-4): 150-157. PMID:18083310. <http://dx.doi.org/10.1016/j.vetpar.2007.10.022>
- Levine ND. *Protozoan parasites of domestic animals and man*. 2nd ed. Burgess Publishing; 1973. 317 p.
- Loretti AP, Barros SS. Parasitismo por *Rangelia vitalii* em cães ("nambiuvú" "peste de sangue") - Uma revisão crítica sobre o assunto. *Arq Inst Biol São Paulo* 2004; 71(1):101-131.
- Loretti AP, Barros SS. Hemorrhagic disease in dogs infected with an unclassified intraendothelial piroplasm in southern Brazil. *Vet Parasitol* 2005; 134(3-4): 193-213. PMID:16153781. <http://dx.doi.org/10.1016/j.vetpar.2005.07.011>
- Matjila TP, Penzhorn BL, Bekker CPJ, Nijhof AM, Jongejan F. Confirmation of occurrence of *Babesia canis vogeli* in domestic dogs in South Africa. *Vet Parasitol* 2004; 122(2): 119-125. PMID:15177716. <http://dx.doi.org/10.1016/j.vetpar.2004.03.019>
- Matjila PT, Leisewitz AL, Ooshuizen MC, Jongejan F, Penzhorn B. Detection of a *Theileria* species in dogs in South Africa. *Vet Parasitol* 2008; 157(1-2): 34-40. PMID:18687528. <http://dx.doi.org/10.1016/j.vetpar.2008.06.025>

- Passos LMF, Geiger SM, Ribeiro MFB, Pfister K, Zahler-Rinder M. First molecular detection of *Babesia vogeli* in dogs from Brazil. *Vet Parasitol* 2005; 127(1): 81-85. PMID:15619377. <http://dx.doi.org/10.1016/j.vetpar.2004.07.028>
- Pestana BR. O Nambyuvú (nota preliminar). *Rev Soc Cie São Paulo* 1910a; 5: 14-17.
- Pestana BR. O Nambyuvú. *Rev Méd São Paulo* 1910b; 22: 423-426.
- Sá AG, Cerqueira AMF, O'Dwyer LH, Macieira DB, Abreu FS, Ferreira RF, et al. Detection and molecular characterization of *Babesia canis vogeli* from naturally infected Brazilian dogs. *Intern J Appl Res Vet Med* 2006; 4(2): 163-168.
- Soares JF, Giroto A, Brandão PE, Da Silva AS, França RT, Lopes STA, et al. Detection and molecular characterization of a canine piroplasm from Brazil. *Vet Parasitol* 2011; 180(3-4): 203-208. PMID:21489694. <http://dx.doi.org/10.1016/j.vetpar.2011.03.024>
- Solano-Gallego L, Trotta M, Carli E, Carcy B, Caldin M, Furlanello T. *Babesia canis canis* and *Babesia canis vogeli* clinicopathological findings and DNA detection by means of PCR-RFLP in blood from Italian dogs suspected of tick-borne disease. *Vet Parasitol* 2008; 157(3-4): 211-221. PMID:18789581. <http://dx.doi.org/10.1016/j.vetpar.2008.07.024>
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA 5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Mol Biol Evol* 2011; 28(10): 2731-2739. PMID:21546353. <http://dx.doi.org/10.1093/molbev/msr121>
- Trapp SM, Messick JB, Vidotto O, Jojima FS, Morais HSA. *Babesia gibsoni* genotype Asia in dogs from Brazil. *Vet Parasitol* 2006; 141(1-2): 177-180. PMID:16765518. <http://dx.doi.org/10.1016/j.vetpar.2006.04.036>
- Uilenberg G. *Babesia* - A historical overview. *Vet Parasitol* 2006; 138(1-2): 3-10. PMID:16513280. <http://dx.doi.org/10.1016/j.vetpar.2006.01.035>
- Zahler M, Rinder H, Schein E, Gothe, R. Detection of a new pathogenic *Babesia microti*-like species in dogs. *Vet Parasitol* 2000; 89(3): 241-248. [http://dx.doi.org/10.1016/S0304-4017\(00\)00202-8](http://dx.doi.org/10.1016/S0304-4017(00)00202-8)