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Comparative morphology of the spermathecae of some species of *Chrysomya* Robineau-Desvoidy and *Cochliomyia* Townsend (Diptera, Calliphoridae)

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ABSTRACT. Comparative morphology of the spermathecae of some species of *Chrysomya* Robineau-Desvoidy and *Cochliomyia* Townsend (Diptera, Calliphoridae). Little is known about the morphology of the chitinized structures of the spermathecae of the Calliphoridae. In this work, the spermathecae of *Chrysomya albiceps* Wiedemann, 1819, *C. megacephala* Fabricius, 1794, *Cochliomyia macellaria* Fabricius, 1775 and *C. hominivorax* Coquerel, 1858 are described and illustrated. The occurrence in one species of four spermathecae, an atypical form for blow flies, was recorded for the first time. The analysis of these structures will allow a better understanding of this group as well as provide taxonomic characters for future phylogenetic studies.

KEYWORDS. Blowflies; females; morphology; taxonomy.

RESUMO. Morfologia comparada das espermatecas de espécies de *Chrysomya* Robineau-Desvoidy e *Cochliomyia* Townsend (Diptera, Calliphoridae). Pouco se conhece sobre a morfologia das estruturas quitinizadas das espermatecas de Calliphoridae. Nesse trabalho as espermatecas de *Chrysomya albiceps* Wiedemann, 1819, *C. megacephala* Fabricius, 1794, *Cochliomyia macellaria* Fabricius, 1775 e *C. hominivorax* Coquerel, 1858 são descritas e ilustradas. Foi registrada pela primeira vez a ocorrência em uma espécie com quatro espermatecas, uma forma atípica em califorídeos. A análise dessas estruturas possibilitará uma melhor compreensão do grupo e fornecerá caracteres taxonômicos para futuros estudos filogenéticos.

PALAVRAS-CHAVE. Fêmeas; morfologia; mosca varejeira; taxonomia.

Calliphorid blowflies belong to the superfamily Oestroidea. Chrysomyinae is divided into two tribes: Chrysomyini and Phormiini; only the former occurs in the Neotropics (Dear 1985). Two genera are important in medicine and/or forensic entomology: *Chrysomya* Robineau-Desvoidy, 1830, introduced in Brazil in the 70's and currently distributed in almost all Brazilian territory (Guimarães *et al.* 1978) and *Cochliomyia* Townsend, 1915, endemic to the Neotropics, with four species (Dear 1985; Guimarães *et al.* 1983; Guimarães & Papavero 1999).

A peculiarity of this group of insects is the relative uniformity of the external characters normally used for identification (McAlpine 1989). Although there are studies of the external morphology and dichotomous keys that help to recognize the involved taxa (de Carvalho & Ribeiro 2000; Mello 2003; Amat *et al.* 2008; de Carvalho & Mello-Patiu 2008), little is known about the morphology of the internal female genitalia. An alternative to aid in the identification of these species is to establish and identify new characters, especially based on the study of the spermathecae.

The spermatheca store and maintain the viability of the spermatozoa and are indirectly related to the processes of mating, fertilization and oviposition (McAlpine 1981; Gschwentner & Tadler 2000). This structure is covered by a glandular epithelium (Dallai *et al.* 1993) originating from ectodermal tissue and is located at the end of the spermathecal duct. This duct is inserted inside the bursa (McAlpine 1981).

Depending on the group of insects, the capsule of the spermathecae may be found in different forms and numbers. The ovoid and subcylindrical-shaped are the most common ones and represent the plesiomorphic condition (Toma & de Carvalho 1995). In Diptera, the predominant number of spermathecae is three, a plesiomorphic condition in the infraorder Muscomorpha. This number is often reduced to one or two spermathecae; a few groups present four (McAlpine 1981).

Studies of the structures of the spermathecae of the Calliphoridae are scarce. Those that exist, in most cases, are found in works describing species and illustrate only the capsule, without mentioning the ducts (*i.e.*, Mello 1968, 1969, 1972a, b, 1974; Mariluis 1978, 1982) (Table I). The identification and description of comparable structural characteristics of the spermathecae may aid in the identification of blowfly species and in future works on the phylogeny of the group. For this reason, we aim to provide a detailed description of the morphological structures of the spermathecae of four species of Calliphoridae: *Chrysomya albiceps* (Wiedemann, 1819), *Chrysomya megacephala* (Fabricius, 1794), *Cochliomyia macellaria* (Fabricius, 1775) and *Cochliomyia hominivorax* (Coquerel, 1858).

MATERIAL AND METHODS

We examined approximately 50 specimens of each species, except for *C. hominivorax*, represented by only 15 speci-

mens. Specimens were collected in the Distrito Federal, Brazil and some reared in laboratory. For microscopic examination, the abdomen was sectioned transversely on the median region of the first tergite, then boiled in lactic acid for about 20 minutes to remove soft tissues and to clarify sclerotized portions. Subsequently the abdomen was transferred to a Petri dish with water and dissected with a cut along the abdominal pleura to remove the spermathecae. Afterwards, microscope slides were mounted with glycerin for analysis. Dissection and mounting of microscope slides was done using a Leica S8APO stereoscopic microscope. Morphological analysis was performed on the structures and images using a Leica DM 2000 microscope equipped with a Leica DFC 295 camera. The morphometric analysis of the images was performed using Leica's LAS V3.6 program. Reference specimens of each species were stored in microtubes containing glycerin and deposited in the Entomological collection da Universidade de Brasília (UnB). The terminology used to describe the morphological structures of the spermathecae follows that proposed by Artigas (1971) and McAlpine (1981), with some modification.

RESULTS

The female reproductive tract of the four species examined, in ventral view, consists of a pair of ovaries, lateral oviducts, accessory glands, median oviduct, bursa, vaginal chamber and three spermathecae (Fig. 1). Two of three spermathecae are encircled by the same glandular tissue, and are associated with an accessory gland positioned to the right. The third spermatheca is isolated and encircled by a glandular tissue, on the opposite side. It is also associated with an accessory gland (Fig. 1). The spermathecae are positioned between the sixth and seventh segments of the telescoping abdomen. Based on the constriction and dilation of the spermathecal duct, it was divided into the following regions: proximal, median and distal (Figs. 2b, c, d).

We found similarities in the morphological patterns of the structures of the spermathecae for the species belonging to the same genus; they are described below.

Chrysomya albiceps (Fig. 3a) has three spermathecae, 0.130 x 0.160 mm in length, semi-spherical in shape, with a smooth surface and brown color. The spermathecal duct has a transverse ridge, is thinner in the proximal region and extends gradually to reach maximum caliber in the median region of the duct. The proximal region is approximately two to three times longer than the distal duct.

In *Chrysomya megacephala* (Fig. 3b), the same pattern was observed: three spermathecae, 0.130 x 0.160 mm in length, semi-spherical in shape, with a smooth surface and brown color. The spermathecal duct has a transverse ridge, is thinner in the proximal region and extends gradually to reach maximum caliber in the median region of the duct. However, in two dissected specimens we observed the presence of four capsules (Fig. 4), two of which were united by a single duct and separated only by the branched proximal region of the spermathecal duct.

Cochliomyia macellaria (Fig. 5a) has three spermathecae, 0.120 x 0.210 mm in length, ovoid in shape, with a smooth surface and brown color. The spermathecal duct has a transverse ridge, is thinner in the proximal and distal regions and gradually reaches maximum caliber in the median region of the duct.

Cochliomyia hominivorax (Fig. 5b) has three spermathecae, 0.136 x 0.261 mm in length, reniform in shape, with a smooth surface and brown color. The spermathecal duct has a transverse ridge, is thinner in the proximal and distal regions and gradually reaches maximum caliber in the median region of the duct.

The common duct in all the studied species is inserted into the bursa. This duct bifurcates and one branch of this fork bifurcates again, linking the common duct to the duct of the spermathecae.

DISCUSSION

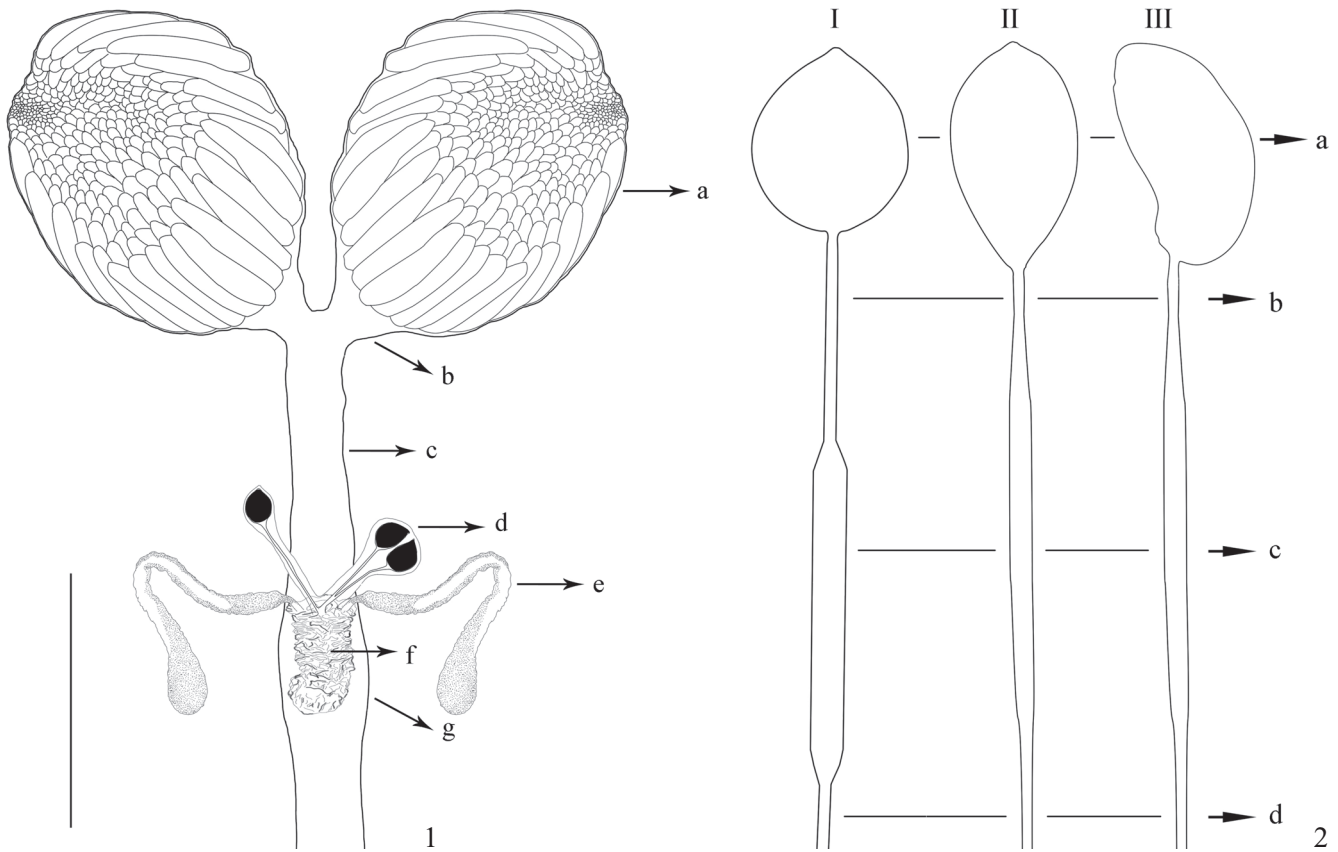
The morphology of the spermathecal structures of four species of blow flies is described and illustrated. Diagnostic characters useful for the identification of species were found only for the genus *Cochliomyia*, based upon differences in the shape of the capsule. However, in *Chrysomya*, such characters are not useful to identify the species. Several authors have described and/or illustrated the structures of the spermathecae of some calliphorids (Table I), but most of these works only described the shape of the capsules.

The structure of the female reproductive system for the four species described here included: a pair of ovaries, lateral oviducts, accessory glands, median oviduct, bursa, vaginal chamber and three spermathecae. These same structures were observed by Lawne (1870), who described and illustrated the female reproductive system of *Calliphora vomitoria* (Linnaeus, 1758).

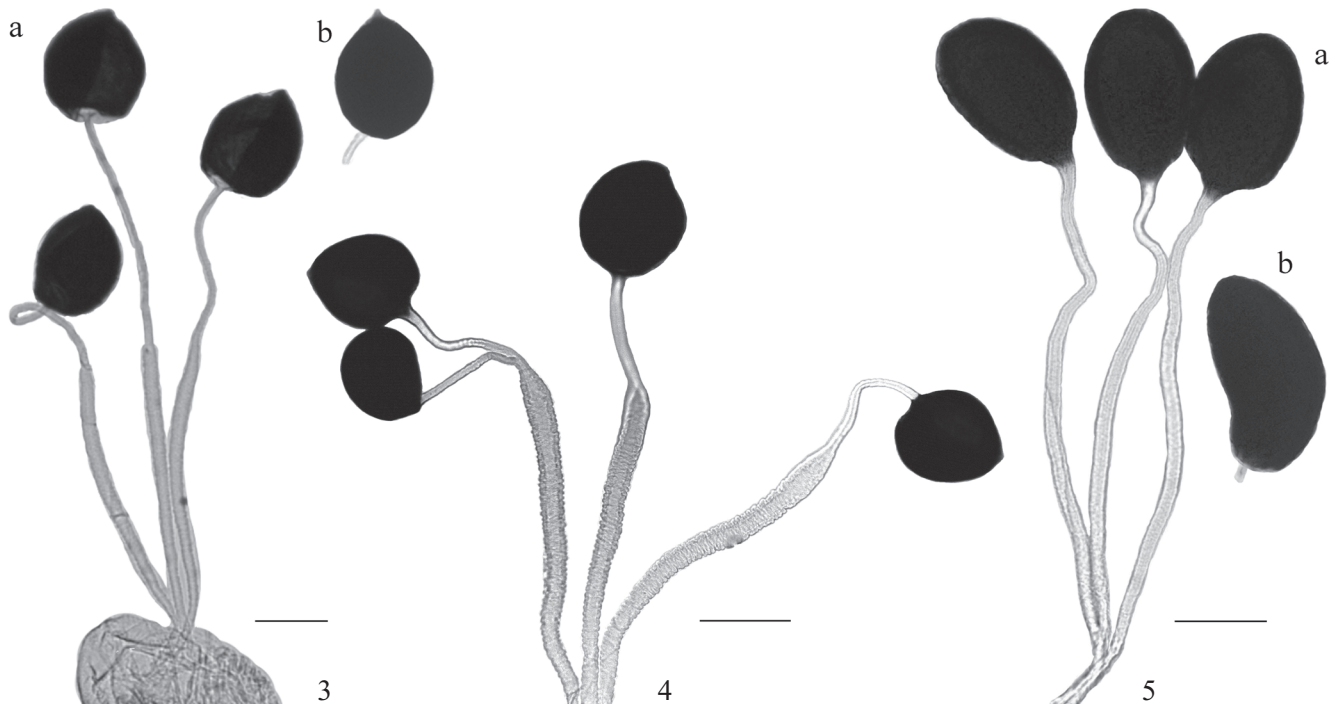
The spermathecae were found to be located between the sixth and seventh abdominal segments, as noted by Shewell (1987) and McAlpine (1981).

The capsule morphology and number of spermathecae observed in the present study is consistent with that reported in prior studies. A semispherical-shaped capsule and three spermathecae were found in *C. albiceps* and *C. megacephala*; Mariluis (1982) reported the same for the species *Lucilia sericata* (Meigen, 1826), *L. eximia* (Wiedemann, 1819), *L. purpurascens* (Walker, 1836). An ovoid-shaped capsule and three spermathecae were found in *C. macellaria* and *C. hominivorax*; Mariluis (1982) reported the same for the two species and Lawne (1870) for *Calliphora vomitoria*. A similar capsule pattern was also reported by Lopes & Albuquerque (1955) for *Callyntropyga humeralis* (Walker, 1837); however, the described texture of that capsule was different, featuring transverse grooves and spines at the base.

Based on the constriction of the spermathecal ducts, a pattern was found among the genera studied (Figs. 2 I, II and III). Comparison with other species belonging to the same family was not possible due to a lack of information. How-



Figs. 1–2. 1, Female reproductive system of the Calliphoridae: (a) ovary, (b) lateral oviduct, (c) median oviduct, (d) spermatheca, (e) accessory gland; (f) bursa, (g) vaginal chamber. Scale bar: 3 mm. 2, Structures of the spermathecae of blow flies: (a) capsule, (b) proximal duct, (c) median duct, (d) distal duct. Pattern found for species: I) *Chrysomya albiceps* and *Chrysomya megacephala*; II) *Cochliomyia macellaria*; III) *Cochliomyia hominivorax*.



Figs. 3–5. 3, Spermathecae: a) *Chrysomya albiceps*; b) *Chrysomya megacephala*. 4, Four spermathecae in *Chrysomya megacephala*. 5, Spermathecae: a) *Cochliomyia macellaria*; b) *Cochliomyia hominivorax*. Scale bars: 0.100 mm.

Table I. List of authors who described the structures of the spermathecae of species belonging to the family Calliphoridae.

Author	Species	Structures of the spermathecae		
		Number	Capsule	Ducts
Lawne 1870	<i>Calliphora vomitoria</i> (Linnaeus, 1758)	3	Dark brown oval shape	Transverse ridge, highest caliber in the medium portion
Lopes & Albuquerque 1955	<i>Callintropyga humeralis</i> (Walker, 1837)	3	Rounded shape with transverse ridge and spines next to the duct	Not described
Mello 1968	<i>Paralucilia fulvicrura</i> (Robineau-Desvoidy, 1830)	Not described	Strongly pigmented ellipsoid shape	Not described
Mello 1969	<i>Paralucilia fulvinota</i> (Bigot, 1877)	Not described	Strongly pigmented globose shape	Not described
Mello 1972a	<i>Sarconesia chlorogaster</i> (Wiedemann, 1830)	Not described	Strongly pigmented long and sinuous shape	Not described
Mello 1972b	<i>Hemilucilia segmentaria</i> (Fabricius, 1805)	Not described	Strongly pigmented globose shape	Not described
	<i>Hemilucilia semidiaphana</i> (Rondani, 1850) = <i>H. flavifacies</i> = <i>H. hermanlenti</i>	Not described	Strongly pigmented elongate shape	Not described
	<i>Hemilucilia benoisti</i> Séguy, 1925	Not described	Strongly pigmented globose shape	Not described
	<i>Hemilucilia souzalopesi</i> Mello 1972	Not described	Strongly pigmented ellipsoid shape	Not described
Mello 1974	<i>Calliphora nigra</i> Mello, 1974	Not described	Strongly pigmented ellipsoid shape	Not described
Mariluis 1978	<i>Calliphora calcedoniae</i> Mariluis, 1978	3	Oval shape	Not described
Mariluis 1982	<i>Paralucilia fulvicrura</i> (Robineau-Desvoidy, 1830)	3	Ellipsoid shape	Not described
	<i>Paralucilia lyrcea</i> (Walker, 1849)	3	Ellipsoid shape	Not described
	<i>Cochliomyia hominivorax</i> (Coquerel, 1858)	3	Oval and enlarged shape	Not described
	<i>Cochliomyia macellaria</i> (Fabricius, 1775)	3	Oval and enlarged shape	Not described
	<i>Lucilia sericata</i> (Meigen, 1826)	3	Lemon shape	Not described
	<i>Lucilia eximia</i> (Wiedemann, 1819)	3	Lemon shape	Not described
	<i>Lucilia purpurascens</i> (Walker, 1836)	3	Lemon enlarged shape	Not described
	<i>Calliphora vicina</i> Robineau-Desvoidy, 1830	3	Oval shape	Not described
	<i>Calliphora antojuanae</i> Mariluis, 1982	3	Oval shape with a slightly curved beak	Not described
	<i>Neta chilensis</i> (Walker, 1836)	3	Spiral shape	Not described
	<i>Sarconesia chlorogaster</i> (Wiedemann, 1830)	3	Broad and sinuous shape distal portion	Not described
	<i>Chlorobrachycoma graciellae</i> Mariluis, 1982	3	Rounded and oval shape	Not described

ever, Lawne (1870) observed that the ducts reached maximum size in the central portion, becoming thinner at the extreme on either side.

We observed four spermathecae in *C. megacephala*, which is an atypical form. The evolution from three to four spermathecae starts from the capsule and flows towards the common duct (Fig. 4). This same phenomenon was observed by Ward *et al.* (2008) in *Scathophaga stercoraria* (Linnaeus, 1758) (Scathophagidae), suggesting that such events may be linked to genetic variations between populations and/or associated with changes in environmental conditions (i.e., microhabitats). According to this author, the formation of a fourth spermatheca results in better genetic quality (i.e., less competition for the storage of sperm), but could generate costs for construction and maintenance of the additional structure. Such costs may result in fewer eggs (Ward *et al.* 2008) and lower total volume of stored sperm when compared to the typical form with three spermathecae (Parker *et al.* 1999).

The presence of three separate ducts emerging from a common duct is considered the plesiomorphic condition, while the branched common duct represents an apomorphic condition. The presence of three basally united ducts can be found in many nematoceros and ground-plan of the Brachycera.

However, the branched common duct is more often found in groups of apical Muscomorpha (McAlpine 1989).

Diagnostic characters based on the structures of the spermathecae were proposed to differentiate the genera *Chrysomya* and *Cochliomyia*. Given the lack of such information for other species of the family Calliphoridae, it is necessary to investigate a larger number of species, so these data may be used with them for the taxonomic and phylogenetic analysis of the group.

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REFERENCES

- Amat, E.; M. C Vélez & M. Wolff. 2008. Clave ilustrada para la identificación de los géneros y las especies de califóridos (Diptera: Calliphoridae) de Colombia. **Caldasia** **30**: 231–244.
- Artigas, J. N. 1971. Las estructuras quitinizadas de la spermatheca y funda del pene de los asilidos y su valor sistemático a través del estudio por taxonomía numérica. **Gayana Zoología** **18**: 1–106.
- de Carvalho, C. J. B. & P. B. Ribeiro. 2000. Chave de identificação das espécies de Calliphoridae (Diptera) do Sul do Brasil. **Revista Brasileira de Parasitologia Veterinária** **9**: 169–173.
- de Carvalho, C. J. B. & C. A. Mello-Patiu. 2008. Key to the adults of the most common forensic species of Diptera in South America. **Revista Brasileira de Entomologia** **52**: 390–406.
- Dallai, R.D.; Marchini, D. & Del Bene, G. 1993. The ultrastructure of the spermatheca in *Ceratitis capitata* Wied. and *Dacus oleae* Gmel. (Diptera: Tephritidae). **Redia** **76**: 147–167.
- Dear, J. 1985. A revision of the new world Chrysomyini (Diptera: Calliphoridae). **Revista Brasileira de Zoologia** **3**: 109–169.
- Gschwentner, R. & A. Tadler. 2000. Functional anatomy of the spermatheca and its duct in the seed bug *Lygaeus simulans* (Heteroptera: Lygaeidae). **European Journal of Entomology** **97**: 305–312.
- Guimarães, J. H.; A. P. Prado & A. X. Linhares. 1978. Three newly introduced blowfly species in southern Brazil (Diptera: Calliphoridae). **Revista Brasileira de Entomologia** **22**: 53–60.
- Guimarães, J. H.; N. Papavero & A. P. Prado. 1983. As míases na Região Neotropical (Identificação, Biologia, Bibliografia). **Revista Brasileira de Zoologia** **1**: 239–416.
- Guimarães, J. H. & N. Papavero. 1999. **Myiasis in man and animals in the Neotropical region**. São Paulo, Plêiade/FAPESP, 308 p.
- Lawne, B. T. 1870. **The anatomy & physiology of the blow-fly (*Musca vomitoria* Linn.)**. A Monograph. London, Thomson & Pinder, Printers, iv+121p.
- Lopes, H. S. & D. O. Albuquerque. 1955. Los insectos de las islas Juan Fernandez 22. Calliphoridae e Sarcophagidae (Diptera). **Revista Chilena de Entomología** **4**: 95–119.
- Mariluis, J. C. 1978. Contribucion al conocimiento del genero *Chlorobrachycoma* Townsend, 1918. (Diptera, Calliphoridae, Toxotarsinae). **Revista de la Sociedad Entomológica Argentina** **37**: 107–111.
- Mariluis, J. C. 1982. Contribucion al conocimiento de las Calliphoridae de la Argentina (Insecta, Diptera). **Opera Lilloana** **33**: 5–59.
- McAlpine, J. F. 1981. Morphology and Terminology – Adultos, p. 9–63. In: McAlpine, J.F.; Peterson, B.V.; Shewell, G.E.; Teskey, H.J.; Vockeroth, J.R. & Wood, D.M. (Eds). **Manual of Nearctic Diptera**. v.2, Monograph n°27, Research Branch, Agriculture Canada, vi+674 p.
- McAlpine, J. F. 1989. Phylogeny and classification of the Muscomorpha, p. 1397–1505. In: J. F. McAlpine & D. M. Wood (Eds.). **Manual of Nearctic Diptera**. v. 3, Monograph n° 32., Research Branch, Agriculture Canada., vi+1581 p.
- Mello, R. P. 1968. Contribuição ao estudo gênero “Paralucilia” Brauer & Bergenstamm, 1891 (Diptera, Calliphoridae). **Revista Brasileira de Biologia** **28**: 177–192.
- Mello, R. P. 1969. Contribuição ao estudo do gênero *Myiolucilia* Hall (Diptera, Calliphoridae). **Studia Entomologica** **12**: 1–4.
- Mello, R. P. 1972a. Contribuição ao estudo do gênero *Sarconesia* Bigot, 1857 (Diptera: Calliphoridae). **Revista Brasileira de Biologia** **32**: 533–537.
- Mello, R. P. 1972b. Revisão das espécies do gênero *Hemilucilia* Brauer, 1985 (Diptera, Calliphoridae). **Revista Brasileira de Biologia** **32**: 539–554.
- Mello, R. P. 1974. Nova espécie do gênero *Calliphora* Robineau-Devoidy, 1930 (Diptera, Calliphoridae). **Arquivos da Universidade Federal Rural do Rio de Janeiro** **4**: 59–63.
- Mello, R. P. 2003. Chave para identificação das formas adultas das espécies da família Calliphoridae (Diptera, Brachycera, Cyclorrhapha) encontradas no Brasil. **Entomologia y Vectores** **10**: 255–268.
- Parker, G. A.; L. W. Simmons; P. Stockley; D. M. McChristie & E. L. Charnov. 1999. Optimal copula duration in yellow dung flies: effects of female size and egg content. **Animal Behavior** **57**: 795–805.
- Shewell, G. E. 1987. Calliphoridae, p. 675–1333. In: J. F. McAlpine; B. V. Peterson; G. E. Shewell; H. J. Teskey; J. R. Vockeroth & D. M. Wood (Eds.). **Manual of Nearctic Diptera**, v.2, Monograph n°28, Ottawa, Agriculture Canada, Research Branch, vi+1332 p.
- Toma, R. & C. J. B. de Carvalho. 1995. Estudo filogenético de Mesembrinellinae com ênfase no gênero *Eumesbrinella* Townsend (Diptera, Calliphoridae). **Revista Brasileira de Zoologia** **12**: 127–144.
- Ward, P. I.; A. J. Wilson & C. Reim. 2008. A cost of cryptic female choice in the yellow dung fly. **Genetica** **134**: 63–67.