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Laboratory characteristics of a cohort of patients with early rheumatoid arthritis

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ABSTRACT

Introduction/Objective: To characterize a population of patients with early rheumatoid arthritis (RA) according to laboratory aspects, comparing it with other similar cohorts. **Methods**: Data presented are part of a prospective incident cohort study that evaluated 65 patients with early RA, followed for 36 months from the diagnosis at Early Rheumatoid Arthritis Clinic of *Hospital Universitário de Brasília (HUB)*. We recorded demographics, clinical, and laboratory data relevant to the cohort initial assessment, including red blood cells, evidence of inflammatory activity, and presence of autoantibodies (rheumatoid factor (RF)), cyclic citrullinated peptide antibodies (anti-CCP), and antivimentin citrullinated (anti-Sa). **Results**: There was a preponderance of female (86%) with mean age of 45.6 years. Twelve patients (18.46%) had laboratory diagnosis of anemia (hemoglobin < 12 g / dL). Erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) were above the reference value for 51 (78.46%) and 46 (70.76%) patients, respectively. Thirty-two patients (49.23%) were positive for at least one of the RF isotypes, and 28 patients (43.07%) were positive for IgA RF, 19 (29.23%) for IgG, and 32 (49.23%) for IgM RF, respectively; 34 patients (52.30%) were positive for at least one of the techniques used in investigation of anti-CCP (CCP2, or CCP3, or CCP3.1), while 9 (13,85%) were positive for anti-Sa. **Conclusions**: The laboratory characteristics of patients enrolled in this Brazilian cohort are similar in many respects to those of North-American, European, and Latin-American cohorts previously published.

Keywords: early rheumatoid arthritis, RF, anti-CCP, anti-Sa, cohort, Brazilian population, early arthritis.

INTRODUCTION

Rheumatoid arthritis (RA) is a chronic condition with irreversible potential for bone and cartilage damage, which, despite recent advances regarding its management, remains costly for the affected individual and society.¹

Although it is well known that RA has varying characteristics according to the population affected, most available information, especially with regard to early RA, comes from Europe and the United States, with few studies in Latin American populations.²⁻⁴

There is no Brazilian cohort study involving patients with early RA. Thus, the laboratory features of early RA in the Brazilian population are unknown, as well as if there is difference from other populations previously studied.

The aim of this study was to characterize a population of patients with early RA according to laboratory tests, including hemoglobin levels; evidence of inflammatory activity by erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP); presence of autoantibody rheumatoid factor (RF), cyclic citrullinated peptide (CCP) antibody, and antivimentin citrullinated (anti-Sa).

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PATIENTS AND METHODS

Data presented are part of a prospective incident cohort study, which evaluated consecutive patients with early RA diagnosis, regularly followed-up for 36 months since diagnosis performed at Early Rheumatoid Arthritis Clinic of *Hospital Universitário de Brasília (HUB)*.

Early RA was defined as the occurrence of joint symptoms compatible with the disease (pain and swelling joints of inflammatory pattern, with or without morning stiffness or other manifestations suggestive of inflammatory joint disease, as assessed by a single observer) and lasting more than six weeks and less than 12 months, regardless of meeting the qualifying criteria of the American College of Rheumatology (ACR).⁵

We recorded demographic and clinical data, as well as laboratory parameters relevant to the cohort initial assessment. Complete blood count and evidence of inflammatory activity (ESR and CRP) were performed at the Laboratory of Clinical Pathology of *Hospital Universitário de Brasília*.

Measurement of RF (IgG, IgM and IgA) was performed at INOVA Diagnostics, Inc., San Diego, California, United States, using "Quanta LiteTM RF IgA ELISA", "Quanta LiteTM RF IgG ELISA", and "Quanta LiteTM RF IgM ELISA" assays (Inova Diagnostics, CA, USA), according to manufacturer protocol. We considered as cutoff points for positivity values higher than 15 IU/mL (IgM and IgA) and 20 UI/mL (FR IgG).

Anti-CCP was studied using "Quanta LiteTM CCP IgG ELISA", "Quanta LiteTM CCP3 IgG ELISA" and "Quanta LiteTM CCP3.1 IgG/IgA ELISA" (Inova Diagnostics CA, USA), according to manufacturer protocol. The serum of each patient was initially diluted at 1:100 in sample diluent. If the result of a sample had an optical density greater than 2.5, it was retested with dilutions at 1:500 and 1:2500, and the unit resulting value multiplied by the dilution factor. Results were expressed in units (U), with < 20 U considered negative, 20-39 U weak positive, 40-59 U moderate positive, and ≥ 60 U strong positive, for all tests.

The test for detection of anti-Sa was performed on the original plates developed by McGill University Autoimmune Research Laboratory.⁶ The results were calculated and expressed in units: < 20 U negative, 21-79 U doubtful, and \geq 80 U positive. The samples were processed for this purpose in the Division of Rheumatology, McGill University Health Center, Quebec, Canada.

Patients received standard treatment regimen used in the service, including the traditional disease modifying anti-

rheumatic drugs (DMARDs) or biological response modifier therapy, according to necessity.

Descriptive statistics was performed for all assessed variables. To detect differences between two means, the Student's t-test or paired t-test for samples of normal distribution was used, considering the average values and standard deviation.

For cases in which normality was rejected, we applied the nonparametric Wilcoxon test or Mann-Whitney test taking into account the median value and the interquartile range.

The study was approved by the Research Ethics Committee of *Faculdade de Medicina da Universidade de Brasília (CEP FM-UnB)*. Project registration: CEP-FM 028/2007.

RESULTS

Characteristics of study population

Initially, we evaluated 65 patients diagnosed with early RA, mean age of 45-64 years (\pm 14-51), ranging from 26 to 71 years. There was a predominance of female (56 patients, 86.15%) and white ethnic group (31 patients, 47.69%). The duration of joint symptoms before diagnosis was on average 32 weeks (\pm 15.41). Although the ACR criteria have not been considered for early RA definition in this study, all 65 patients met at least four criteria in the first assessment. Table 1 summarizes these population demographic and clinical characteristics.

Red cell counts

In the initial evaluation, the mean hemoglobin value of 65 patients was 12.73 g/dL (\pm 1.06). Twelve patients (18.46%) had laboratory diagnosis of anemia (hemoglobin < 12 g/dL), with mean hemoglobin value of 10.91 g/dL (\pm 1.21).

Evidence of inflammatory activity

As for VHS, 51 patients (78.46%) had values above those of reference test, with the average value of 40.43 mm in the first hour (\pm 16.97). The level of CRP showed higher values than the reference test (1.0 mg/dL) in 46 patients (70.76%), and the average value found was 2.46 mg/dL (\pm 1.72).

Autoantibodies

Rheumatoid factor

In the first assessment, among the 65 patients, 32 subjects (49.23%) were positive for at least one of the RF isotypes, and 28 patients (43.07%) were positive for RF IgA, 19 (29.23%) for IgG, and 32 (49.23%) for IgM RF, respectively. Among

Table 1

Patients with early rheumatoid arthritis assessed at *Hospital Universitário de Brasília* according to their general characteristics (baseline assessment, n: 65)

Characteristics		n (±) or n (%)
Age (years)		45.64 (± 14.51)
Sex	Male	9 (13.80%)
	Female	56 (86.15%)
Ethnic group	White	31 (47.69%)
	White/black	18 (27.69%)
	White/native	13 (20%)
	Black	1 (1.53%)
	Black/native	2 (3.07%)
Education (years)		8.3 (± 4.95)
Duration of disease (weeks)		32 (± 15.41)
Current or previous smoking		7 (10.76%)
DAS 28		6.79 (± 1.11)
HAQ		1.87 (± 0.81)
Radiographic erosion		31 (47.69%)

Variables are represented in mean absolute value (± standard deviation) or n (%). DAS 28: disease activity index; HAQ: Health Assessment Questionnaire.

those with positive serology for RF, the mean titers of IgA at baseline were 76 IU/dL (\pm 56.17), RF IgG 71 IU/mL (\pm 51.21), and RF IgM 105 IU/mL (\pm 73.13).

Twenty-eight patients (43.07% of the total sample and 87.50% of those positive for at least one of the RF serotypes) were positive for more than one serotype. Seventeen patients (26.15% of the total sample and 53.12% of those positive for at least one of the RF serotypes) were positive for all three RF serotypes. Two patients (3.07% of the total sample and 6.25% were positive for at least one of the RF serotypes) were positive only for IgA RF, and six patients (9.23% of the total sample and 18.75 % of those positive for at least one of the RF serotypes) were positive only for IgM and negative for the other serotypes. No patient tested positive only for IgG. Five patients (7.69% of total sample and 15.62% of those positive for at least one RF serotype) were positive for RF IgA, IgM and negative for IgG, while two (3.07% of the total sample and 6.25% of those positive for at least one of the RF serotypes) were positive for IgG and IgM but negative for IgA RF. No patient was positive for IgA and IgG or negative for IgM.

Cyclic citrullinated peptide antibodies (anti-CCP)

As for anti-CCP antibodies, 34 patients (52.30% of the total) were positive for at least one of the techniques used in screening (CCP2,

CCP3, or CCP3.1). With the use of ELISA 2 (CCP2) technique, 33 patients (50.77% of the total population tested) were negative, five (7.69%) were weak positive, and 27 (41.54%) were strong positive. With ELISA 3 (CCP3) technique, 30 patients (46.15%) were negative, five (7.69%) were weak positive, two (3.08%) were moderate positive, and 28 (43.08%) were strong positive. With ELISA 3.1 (CCP3.1) technique, 31 patients (47.69%) were negative, two (3.08%) were weak positive, three (4.62%) were moderate positive, and 29 (44.62%) were strong positive.

Among those with positive serology for anti-CCP, the average values obtained by CCP2 technique at baseline were 568 IU/dL (\pm 833.28); by CCP3, 1,148 IU/mL (\pm 1,584.15); and by CCP3.1, 1,272 IU/mL (\pm 1,721.97). Titles obtained by the third generation techniques were not significantly higher than those obtained by the second generation technique (P > 0.05).

Thirty-two patients (49.23% of the total population and 94.11% of those with positive results for at least one of the techniques) were positive for anti-CCP in more than one technique, while 29 patients (44.61% of total population and 85.29% of those positive for at least one of the techniques) were positive for all three techniques.

Three patients (4.62% of the total population and 8.82% among the positives) were positive for anti-CCP3 and CCP3.1 and negative for anti-CCP2, and one patient (1.53% of total population and 2.94% among positives) was positive for CCP3 and negative for CCP2 and CCP3.1 (in all cases, positive results were weak positive). Two patients (3.08% of total population and 5.88% of the total number of positives) were positive for anti-CCP 3.1 and CCP 2 and negative for CCP3 (in both cases the result by CCP3.1 technique was weak positive). There was no statistical difference between positivity for anti-CCP analyzed by different techniques – CCP 2, CCP 3, and CCP 3.1 (P > 0.05).

Antivimentin citrullinated (anti-Sa)

At baseline assessment, of the 65 patients evaluated, 52 (80%) were anti-Sa negative, four (6.15%) had an uncertain outcome, and nine (13.85%) were positive. Among those with anti-Sa positive serology, the mean values obtained at baseline was 370.2 IU/dL (\pm 263.80). Table 2 summarizes the profile of positivity for RF, anti-CCP, and anti-Sa in 65 patients initially evaluated.

DISCUSSION

The interaction between multiethnic origins, colonial heritage, and immigration patterns in Latin America resulted in rather complex demographic characteristics and in a highly mixed population, varying between different countries in the region, with a wide variability of gene expression.²⁻⁴

Data on incidence, prevalence, and characteristics of RA in populations of Latin American countries are scarce.⁷ In analyzing the results of studies on AR performed in developing countries, one should bear in mind that the disease characteristics can be affected by socioeconomic, demographic, and health systems of these countries.⁸

The characteristics of patients in our cohort were compared with data from other cohorts, American and European, and with preliminary information from GLADAR,⁹ prospective, multicenter observational cohort study that evaluated 1,059 patients with early RA, allocated in 46 centers of 14 Latin American countries.^{10,11} The Rheumatology Service of HUB/ UnB participated in the GLADAR study with 30 patients other than those evaluated in this study.

Hemocytometer

Anemia is a relatively frequent extra-articular manifestation in early RA (6-25%), and seems to correlate with worse joint prognosis,¹² functional disability, need for orthopedic

Table 2

Baseline serum characteristics of patients with early rheumatoid arthritis (n: 65)

Autoantibody	n (%)/title (IU/dL) - mean (± SD)	
RFs (any isotype)	32 (49.23%)	
RF IgM	32 (49.23%)/ 105 (± 73.13)	
RF IgG	19 (29.23%) / 71 (± 51.21)	
RF IGA	28 (43.07%) / 76 (± 56.17)	
RF IgM+ IgG + IgA+	17 (26.15%)	
RF IgA+ IgM+IgG-	5 (7.69%)	
RF IgM + IgG- IgA-	6 (9.23%)	
RF IgA +IgM- IgG-	2 (3.07%)	
RF IgM+ IgG+ IgA -	2 (3.07%)	
Anti-CCP (any technique)	34 (52.3%)	
CCP2	32 (49.23%) / 568 (± 833.28)	
CCP3	35 (53.85%) / 1,148 (± 1,584.15)	
CCP3.1	34 (52.31%) / 1,272 UI/mL (± 1,721.97)	
CCP2 + CCP3 + CCP3.1 +	29 (44.61%)	
CCP2 - CCP3 + CCP3.1 +	3 (4.62%)	
CCP2 - CCP3 + CCP3.1 -	1 (1.53%)	
CCP2 - CCP3 - CCP3.1 +	2 (3.08%)	
Anti-Sa	9 (13.85%) / 370.2 (± 263.8)	

Variables are represented in mean absolute value (± standard deviation) or n (%) RF: rheumatoid factor; CCP: cyclic citrullinated peptide antibodies; anti-Sa: antivimentin citrullinated antibodies. intervention,¹³ and mortality.¹⁴ Decreased RBC count is more frequent among male, smokers, patients with high levels of evidence of inflammatory activity, presence of RF, ANA and shared epitope.¹⁴

Nikolaisen *et al.*¹⁵ investigated the prevalence of anemia in a cohort of 111 consecutive patients with early RA during 74 months of follow-up and found reduced levels of hemoglobin in 25% of patients during the first year of follow-up. In this study, the presence of anemia was associated with higher levels of ESR, CRP, and IL-6, but not the more aggressive joint disease or mortality.

Evidence of inflammatory activity

More than two thirds of patients in our cohort showed increased evidence of inflammatory activity (ESR and CRP) tested at baseline.

The evidence of inflammatory activity at baseline does not seem to discriminate RA from other early arthritis, and do not predict persistent disease (erosive).¹⁶⁻¹⁸ Tunn and Bacon reported that, among patients of an early RA clinic who developed persistent arthritis, levels significantly higher of ESR was found. However, in this study, ESR had low discriminatory power and poor contribution to patient's longterm prognosis.¹⁹

Autoantibodies

Rheumatoid factor

At baseline evaluation, about 50% of patients in our cohort were positive for at least one of the RF serotypes, a percentage lower than that reported for GLADAR cohort (76%).²⁰ It is important to highlight the different methods employed for RF research by GLADAR (Waaler-Rose, nephelometry, ELISA). RF positivity in our study was similar to other studies that used ELISA,²¹⁻²² including the results of Nishimura *et al.* meta-analysis.

In our study, we chose to research RF isotypes IgA, IgG, and IgM. The validity of the RF isotypes research in the assessment of early RA remains questionable. For example, the existence of correlation between titers of different isotypes of RF and the diagnosis of RA is not defined, as well as the relation between the presence of some specific serotype (or more than one) and a worst radiologic prognostic, or the behavior of different RF isotypes over time.²³

In our population, we observed IgM RF in 50%, IgA in 42% and IgG in 30% of patients diagnosed with RA and symptoms lasting less than 12 months, similar rates to those

referred in other works, such as the work by Vittecoq *et al.*, ²⁴ which described the presence of IgM RF in 51%, IgA RF in 36%, and IgG RF in 32% of patients diagnosed with RA with less than two years duration.

IgM RF is a useful marker to discriminate patients with polyarthritis that will evolve or not to RA.²⁵⁻³⁰ In contrast, the diagnostic properties of RF IgA and IgG are questionable.³⁰⁻³² In our study, the research of RF serotypes IgA and IgG did not increase the frequency of RF positivity and, therefore, does not contribute to RA diagnosis.

Some published studies have evaluated, as in our cohort, the average titles of different RF serotypes in early RA, and our results are similar to those reported by other authors.^{23,33}

Cyclic citrullinated peptide antibodies (anti-CCP)

The percentage of positivity for anti-CCP in our study was similar to that reported by several other studies involving patients with early RA. Fifty percent of patients in our cohort were positive for at least one of the techniques used in the assessment (CCP2, CCP3, or CCP3.1), and most were strong positive by the all three techniques. In a systematic literature review, the combined analysis of publications referring to more than 2,000 patients with early undifferentiated arthritis showed a 23% prevalence of anti-CCP (ELISA 2nd generation). This prevalence increased to 51% in over 1,000 patients who met criteria for RA after a mean follow-up of 18 months.³⁴

In our cohort, the prevalence of RF and anti-CCP was approximately the same (considering CCP positive for any of the three techniques examined), which was similar to other studies on the subject-matter.³⁵⁻³⁶ As reported by several authors, CCP2 appears to be as sensitive as – and more specific than – IgM RF, but their advantage would be the detection of antibodies in approximately 15% of patients with RA who are negative for RF.³⁷⁻⁴⁵ Nishimura *et al.*,²³ in their meta-analysis of published studies on accuracy of anti-CCP and RF for RA, have concluded that positivity for anti-CCP alone is more specific than the isolated positivity for IgM RF in RA diagnosis.

In our cohort, there was no difference between the techniques considered for detection of anti-CCP (CCP2, CCP3, and CCP3.1), and the prevalence of anti-CCP antibodies was almost the same by all three techniques (40%). The difference in sensitivity, specificity, and cost-effective among the three techniques for detection of anti-CCP is still controversial in literature, and studies in different populations are needed.⁴⁶

In 2005, a third generation of anti-CCP (CCP3) became available for laboratory diagnosis of RA. It was reported that these tests would recognize additional citrullinated epitopes, which would not be identified by the second generation test (CCP2), with sensitivity 5% greater than CCP2, maintaining specificity.⁴⁷

CCP3 test was evaluated Santiago *et al.*⁴⁸ and Wu *et al.*⁴⁹ and found to be more sensitive than the CCP2 while maintaining specificity. Anjos *et al.*⁴⁶ reported in a population of 70 patients with RA in southern Brazil that both CCP2 and CCP3 showed good diagnostic performance, with CCP3 4.3% more sensitive than CCP2, while maintaining specificity. However, other authors have reported very similar diagnostic performance between CCP2 and CCP3 tests.⁵⁰⁻⁵¹

The CCP3.1 evaluated in our study (INOVA) uses a conjugate that detects antibodies IgA, as well as the usual IgG antibodies, which in theory would improve the method sensitivity, since some patients with RA have IgA antibodies against CCP3, in the absence of IgG antibodies.⁵² Bizzaro *et al.*,⁵³ however, comparing 11 different laboratory techniques for CCP detection, noted a slight difference in results between CCP2 and CCP3 INOVA (sensitivity 64% and 67%, respectively) and no difference between CCP3 and CCP3.1, suggesting that the combination of IgA and IgG would not improve test performance, similar to what was observed in our cohort.

In our cohort, the titles of CCP obtained by three different techniques were similar. Titles of CCP2 on average tended to be lower than the third generation techniques. We observed high values (on average > 500 IU/dL) with the three techniques, higher than those reported in publications by Lee *et al.*⁵⁴ and Papadopoulos *et al.*,⁵⁵ two of the few studies on absolute titles of anti-CCP and its correlation with disease progression.

Antivimentin citrullinated (anti-Sa)

Less than 15% of our patients' cohort presented with anti-Sa antibodies in the initial evaluation; this value is less than that reported by Boire *et al.*⁶ (28% of their cohort of 165 patients with early polyarthritis) and Vossenaar *et al.*⁵⁶ (40% of 87 sera from patients with established RA).

The mean titers of anti-Sa found in our cohort varied from 200 to 300 IU/dL, values similar to those found by other authors,^{6,57} although there are few publications on the subject.

CONCLUSIONS

Although the demographic and clinical characteristics of patients enrolled in this Brazilian cohort differ in several aspects from those of North American, European, and Latin American cohorts previously published, our laboratory findings, including the initial prevalence of autoantibodies, are similar to other populations.

The prevalence of RF and anti-CCP (50%) was similar to that reported in other cohorts of early RA. As the initial positivity for both autoantibodies was similar in our cohort, we infer that, in our specific population, anti-CCP did not aggregate value for the diagnosis of RA in its initial phase.

Additionally, there was no difference between the techniques considered for detection of anti-CCP (CCP2, CCP3, and CCP3.1), suggesting that the third-generation tests did not bring contribution to the diagnosis of early RA. Moreover, the research of anti-Sa was not useful for diagnosis of early RA, compared to RF and anti-CCP.

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REFERÊNCIAS

REFERENCES

- Lee DM, Weinblatt ME. Rheumatoid arthritis. Lancet 2001; 358:903-11.
- Callegari-Jacques SM, Grattapaglia D, Salzano FM, Salamoni SP, Cronetti SG, Ferreira ME *et al.* Historical genetics: spatiotemporal analysis of the formation of the Brazilian population. Am J Hum Biol 2003; 15:824-34.
- Lisker R, Ramirez E, Bricen o RP. Gene frequencies and admixture estimates in four Mexican urban centers. Hum Biol 1990; 62:791-801.
- Sans M, Salzano FM, Chakraborty R. Historical genetics in Uruguay: estimates of biological origins and their problems. Hum Biol 1997; 69:161-70.
- Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS *et al*. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988; 31:315-24.
- Boire G, Cossette P, de Brum-Fernandes AJ, Liang P, Niyonsenga T, Zhou ZJ *et al.* Anti-Sa antibodies and antibodies against cyclic citrullinated peptide are not equivalent as predictors of severe outcomes in patients with recent-onset polyarthritis. Arthritis Res Ther 2005; 7:R592-603.

- Cardiel MH; Latin American Rheumatology Associations of the Pan-American League of Associations for Rheumatology (PANLAR); Grupo Latinoamericano de Estudio de Artritis Reumatoide (GLADAR). First Latin American position paper on the pharmacological treatment of rheumatoid arthritis. Rheumatology (Oxford) 2006; 45:ii7-ii22.
- Mijiyawa M. Epidemiology and semiology of rheumatoid arthritis in Third World countries. Rev Rhum Engl Ed 1995; 62:121-6.
- Grupo Latino Americano de Estudio de Artritis Reumatoide. Disponível em http://www.gladar.org/>. Acesso em: 07 abr. 2009.
- Rheumatoid arthritis in Latin America. Disponível em http://www.sochire.cl/Dr_Pons_Estell.pdf>. Acesso em: 27. abr. 2009.
- Estel BAP, Massardo L, Wojdyla D, Acevedo E, Laurindo IMM, Guibert ZM *et al.* Is there something we can learn from rheumatoid arthritis in Latin America? A descriptive report on an inception Cohort of 1093 patients Ann Rheum Dis 2008; 67:336 (Abstract).
- 12. Dixey J, Solymossy C, Young A. Early RA Study, Is it possible to predict radiological damage in early rheumatoid arthritis (RA)? A report on the occurrence, progression, and prognostic factors of radiological erosions over the first 3 years in 866 patients from the Early RA Study (ERAS). J Rheumatol Suppl 2004; 69:48-54.
- James D, Young A, Kulinskaya E, Knight E, Tomson W, Oliver W *et al.* Orthopaedic intervention on early rheumatoid arthritis. Occurrences and predictive factors in an inception cohort of 1064 patients followed for 5 years. Rheumatology 2004; 43:369-76.
- Young A, Koduri G. Extra-articular manifestations and complications of rheumatoid arthritis. Best Pract Res Clin Rheumatol 2007; 21:907-27.
- Nikolaisen C, Figenschau Y, Nossent JC. Anemia in early rheumatoid arthritis is associated with interleukin 6-mediated bone marrow suppression, but has no effect on disease course or mortality. J Rheumatol 2008; 35:380-6.
- 16. Visser H, le Cessie S, Vos K, Breedveld FC, Hazes JM. How to diagnose rhematoid arthritis early. A prediction model for persistent (erosive) arthritis, Arthritis Rheum 2002; 46:357-65.
- Green M, Marzo-Ortega H, McGonagle D, Wakefield R, Proudman S, Conaghan P *et al*. Persistence of mild, early inflammatory arthritis: the importance of disease duration, rheumatoid factor, and the shared epitope. Arthritis Rheum 1999; 42:2184-8.
- van der Heijde DM, van RP, van RM van de Putte LB. Influence of prognostic features on the final outcome in rheumatoid arthritis: a review of the literature. Semin Arthritis Rheum 1988; 17:284-92.
- Tunn EJ, Bacon PA. Differentiating persistent from self-limiting symmetrical synovitis in an early arthritis clinic. Br J Rheumatol 1993; 32:97-103.
- Siegel DM. Chronic arthritis in adolescence. Adolesc Med State Art Rev 2007; 18:47-61.
- Nell-Duxneuner V, Machold K, Stamm T, Eberl G, Heinzl H, Hoefler E *et al*. Autoantibody profiling in patients with very early rheumatoid arthritis - a follow-up study. Ann Rheum Dis. 2010; 69(1):169-74.
- Tedesco A, D'Agostino D, Soriente I, Amato P, Piccoli R, Sabatini P. A new strategy for the early diagnosis of rheumatoid arthritis: a combined approach. Autoimmun Rev 2009; 8:233-7.
- Nishimura K, Sugiyama D, Kogata Y, Tsuji G, Nakazawa T, Kawano S *et al*. Meta-analysis: diagnostic accuracy of anti-cyclic citrullinated peptide antibody and rheumatoid factor for rheumatoid arthritis. Ann Intern Med 2007; 146:797-808.

- Vittecoq O, Pouplin S, Krzanowska K, Jouen-Beades F, Menard JF, Gayet A *et al.* Rheumatoid factor is the strongest predictor of radiological progression of rheumatoid arthritis in a three-year prospective study in community-recruited patients. Rheumatology (Oxford) 2003; 42: 939-46.
- 25. Vallbracht I, Rieber J, Oppermann M, Förger F, Siebert U, Helmke K. Diagnostic and clinical value of anti-cyclic citrullinated peptide antibodies compared with rheumatoid factor isotypes in rheumatoid arthritis. Ann Rheum Dis 2004; 63:1079-84.
- Greiner A, Plischke H, Kellner H, Gruber R. Association of anticyclic citrullinated peptide antibodies, anti-citrullin antibodies, and IgM and IgA rheumatoid factors with serological parameters of disease activity in rheumatoid arthritis. Ann N Y Acad Sci 2005; 1050:295-303.
- Wolfe F, Cathey MA, Roberts FK. The latex test revised rheumatoid factor testing in 8,287 rheumatic disease patients. Arthritis Rheum 1991; 34:951-60.
- Schellekens GA, Visser H, de Jong BAW, van den Hoogen FH, Hazes JM, Breedveld FC *et al*: The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. Arthritis Rheum 2000; 43:115-63.
- 29. Visser H. Early diagnosis of rheumatoid arthritis. Best Pract & Res Clin Rheum 2005; 19:55-72.
- Saraux A, Berthelot JM, Chalès G, Le Henaff C, Mary JY, Thorel JB *et al.* Value of laboratory tests in early prediction of rheumatoid arthritis. Arthritis Rheum 2002; 47:155-65.
- Vittecoq O, Pouplin S, Krzanowska K, Jouen-Beades F, Menard JF, Gayet A *et al.* Rheumatoid factor is the strongest predictor of radiological progression of rheumatoid arthritis in a three-year prospective study in community-recruited patients. Rheumatology (Oxford) 2003; 42:939-46.
- 32. Procaccia S, Gasparini A, Colucci A, Lanzanova D, Bianchi M, Forcellini P *et al.* ELISA determined IgM, IgG and IgA rheumatoid factors in rheumatoid arthritis and in other connective tissue diseases. Clin Exp Rheumatol 1987; 5:335-42.
- Westwood OM, Nelson PN, Hay FC. Rheumatoid factor: what's new? Rheumatology Oxford 2006; 45:379-85.
- Avouac J, Gossec L, Dougados M. Diagnostic and predictive value for anti-cyclic citrullinated protein antibodies in rheumatoid arthritis: a systematic literature review. Ann Rheum Dis 2006; 65:845-51.
- Dorner T, Egerer K, Feist E, Burmester GR. Rheumatoid factor revisited. Curr Opin Rheumatol 2004; 16:246-53.
- Goldbach-Mansky R, Lee J, McCoy A, Hoxworth J, Yarboro C, Smolen JS *et al*. Rheumatoid arthritis associated autoantibodies in patients with synovitis of recent onset. Arthritis Res 2002; 3:236-43.
- Saraux A, Berthelot JM, Devauchelle V, Bendaoud B, Chalès G, Le Henaff C *et al.* Value of antibodies to citrulline-containing peptides for diagnosing early rheumatoid arthritis. J Rheumatol 2004; 30:2535-9.
- Sauerland U, Becker H, Seide M, Schotte H, Willeke P, Schorat A et al. Clinical utility of the anti-CCP assay: experiences with 700 patients. Ann NY Acad Sci 2005; 1050:314-8.
- Dubrous P, Gardet V, Hugard L. Value of anti-cyclic citrullinated peptides antibodies in comparison with rheumatoid factor for rheumatoid arthritis diagnosis. Pathol Biol 2005; 53:63-7.

- 40. Silveira IG, Burlingame RW, von Mühlen CA, Bender AL, Staub HL. Anti-CCP antibodies have more diagnostic impact than rheumatoid factor (RF) in a population tested for RF. Clin Rheumatol 2007; 26:1883-9.
- 41. Vallbracht I, Helmke K. Additional diagnostic and clinical value of anti-cyclic citrullinated peptide antibodies compared with rheumatoid factor isotypes in rheumatoid arthritis. Autoimmun Rev 2005; 4:389-94.
- 42. Solanki K, Spellerberg M, Chapman P, Moller P, O'Donnell J. Anticyclic citrullinated antibodies: complementary to IgM rheumatoid factor in the early diagnosis of rheumatoid arthritis. J N Z Med 2004; 117:1097.
- Araki C, Hayashi N, Moriyama M, Morinobu S, Mukai M, Koshiba M *et al.* Usefulness of anticyclic citrullinated peptide antibodies (anti-CCP) for the diagnosis of rheumatoid arthritis. Rinsho Byori 2004; 52:966-72.
- 44. van Dongen H, van Aken J, Lard LR, Visser K, Ronday HK, Hulsmans HM *et al.* Efficacy of methotrexate treatment in patients with probable rheumatoid arthritis: a double-blind, randomized, placebo-controlled trial. Arthritis Rheum 2007; 56:1424-32.
- 45. Quinn MA, Gough AKS, Green MJ, Devlin J, Henrsor EMA, Greenstein A *et al.* Anti-CCP antibodies measured at disease onset help identify soronegative rheumatoid arthritis and predict radiological and functional outcome. Rheumatology 2006; 45:478-80.
- 46. Anjos LME, Pereira IA, d'Orsi E, Seaman A, Burlingame RW, Morato EF. A comparative study of IgG second and third generation anti-cyclic citrullinated peptide (CCP) ELISAs and their combination with IA third generation ELISA for the diagnosis of RA. Clin Reumatol 2009; 28:153-8.
- 47. Vieira LMEA, d'Orse E, Pereira IA, Morato EF, Burlingame R. Rheumatoid arthritis diagnosis: a comparative study of second and third generation anti-cyclic citrullinated peptide (CCP) antibody ELISAS. INOVA Newsletter 2007; 2:8-9.
- 48. Santiago M, Baron M, Miyachi K, Fritzler MJ, Abu-Hakima M, Leclercq *et al.* A comparison of the frequency of antibodies to cyclic citrullinated peptides using a third generation anti-CCP assay (CCP3) in systemic sclerosis, primary biliary cirrhosis and rheumatoid arthritis. Clin Rheumatol 2008; 27:77-83.
- 49. Wu R, Shovman O, Zhang Y, Gilburd B, Zandman-Goddard G, Shoenfeld Y (2007) Increased prevalence of anti-third generation cyclic citrullinated peptide antibodies in patients with rheumatoid arthritis and CREST syndrome. Clin Rev Allergy Immunol 2007; 32:47-56.
- Caro-Oleas JL, Fernandez-Suarez A, Reneses-Casteros S, Porrino C, Nunes-Roldan A, Wichmann-Schlipf I. Diagnostic usefulness of a third generation anti-cyclic citrulline antibody test in patients with recent-onset polyarthritis. Clin Chem Lab Med 2007; 45:1396-401.
- Lutteri L, Malaise M, Chapelle JP. Comparison of second- and third-generation anti-cyclic citrullinated peptide antibodies assays for detecting rheumatoid arthritis. Clin Chim Acta 2007; 386:76-81.
- 52. Szekanecz Z, Burlingame R. The INOVA CCP 3.1 IgA/IgG ELISA represents significant improvement in the laboratory diagnosis of rheumatoid arthritis. INOVA Newsletter 2007; 2:6-7.
- Bizzaro N, Tonutti E, Tozzoli R, Villalta D. Analytical and Diagnostic Characteristics of 11 2nd- and 3rd-Generation Immunoenzymatic Methods for the Detection of Antibodies to Citrullinated Proteins. Clin Chem 2007; 53:1527-33.

- Lee DM, Phillps R, Hagan EM, Chibnik LB, Costenbader KH, Schur PH. Quantifying anti-cyclic citrullinated peptide titres: clinical utility and association with tobacco exposure in patients with rheumatoid arthritis. Ann Rheum Dis 2009; 68:201-8.
- 55. Papadooulos NG, Tsiasousis GZ, avlitou-Tsioontsi A, Giannakon A, Galanopoulou VK. Does the presence of anti-CC autoantivodies and their serum levels influence the severity and activity in rheumatoid arthritis patients? Cli Rev Allergy Immunol 2008; 34:11-5.
- Vossenaar ER, Després N, Lapointe E, van de Heijden A, Lora M, Senshu T *et al.* Rheumatoid arthritis specific anti-Sa antibodies target citrulliated vimentin. Arthritis Res Ther 2004; 6:142-50.
- 57. Innala L, Kokkonen H, Ericsson C, Jidell E, Berglin E, Rantapää-Dahlqvist S. Antibodies against mutated citrullinated vimentin are a better predictor of disease activity at 24 months in early rheumatoid arthritis than antibodies against cyclic citrullinated peptides. J Rheumatol 2008; 35:1002-8.