

RESEARCH ARTICLE

Venous blood gases and electrolyte values of captive red foot tortoise (*Chelonoidis carbonarius*)

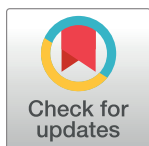
Sofia Silva La Rocca de Freitas^{1☯*}, Laís Velloso Garcia^{2‡}, Jairo Antonio Melo dos Santos^{3‡}, Líria Queiroz Luz Hirano^{4☯}

1 Resident of Wild Animals Medicine at Federal University of Uberlândia (UFU), Uberlândia, Minas Gerais, Brasil, **2** Veterinary at University at Brasília (UnB), Brasília, Brasil, **3** Chief Executive Officer at S.E.V.VO, Brasília, Brasil, **4** Professor of the Graduate Program in Animals Sciences at University of Brasília (UnB), Brasília, Brasil

☯ These authors contributed equally to this work.

‡ LVG and JAMS also contributed equally to this work.

* sofiaslarocca@gmail.com



OPEN ACCESS

Citation: Silva La Rocca de Freitas S, Garcia LV, dos Santos JAM, Luz Hirano LQ (2024) Venous blood gases and electrolyte values of captive red foot tortoise (*Chelonoidis carbonarius*). PLoS ONE 19(3): e0299451. <https://doi.org/10.1371/journal.pone.0299451>

Editor: Ulrike Gertrud Munderloh, University of Minnesota, UNITED STATES

Received: May 31, 2023

Accepted: February 10, 2024

Published: March 15, 2024

Copyright: © 2024 Silva La Rocca de Freitas et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The authors received no specific funding for this work.

Competing interests: The authors have declared that no competing interests exist.

Abstract

Blood gas analysis reflects the exchange of oxygen and carbon dioxide in the lungs. This test provides important information, since the relationship between these gases has a direct impact on the acid-basic balance in the body. Given the significance of blood gas analysis in Brazilian reptiles, this study set out to establish temperature-corrected and uncorrected reference intervals for venous blood gas measurements in *Chelonoidis carbonarius*, and to compare values between females and males. In this study, 19 animals were used, 8 males and 11 females. Blood samples were collected from the dorsal coccygeal vein, and the analyses were performed immediately after blood sample collection. The following parameters were measured: pH, PO₂, HCO₃⁻, TCO₂, BE_{ecf}, Na, K, I_{Ca}, and Glu, and were compared between females and males. Additionally, pH, pCO₂, and pO₂ values were compared with and without temperature correction. Oxygen saturation and Na levels were significantly higher (p<0.05) in males. Furthermore, it was possible to infer that the lower the body temperature relative to the environmental temperature, the larger the difference in pH following temperature correction.

Introduction

Blood gas analysis reflects the exchange of oxygen and carbon dioxide in the lungs. This test provides important information, since the relationship between these gases has a direct impact on the acid-basic balance in the body. Abnormal blood gas levels may indicate respiratory, metabolic or renal disorders. Venous blood gas values are similar to arterial blood values. The only exception is partial pressure of oxygen, which is significantly lower in venous blood due to oxygen consumption in metabolic processes [1].

Body temperature affects the concentration of most blood gases. Ceretta et al. [2] compared venous blood gas measurements with and without temperature correction in *Agkistrodon*

contortrix and *Pantherophis alleghaniensis*. Temperature correction yielded higher pH values, whereas remaining parameters tended to be lower when temperature correction was used.

According to Lewbart et al. [3], the establishment of species-specific parameters is particularly important in reptiles due to adaptive variations driven by environmental factors. Blood gas values have been reported in *Agkistrodon contortrix*, *Pantherophis alleghaniensis* [2], *Chelonia mydas* [4], *Caretta caretta* [5] and *Amblyrhynchus cristatus* [3]. However, despite the growing relevance of blood gas analysis in reptile medicine, there is a limited amount of data, especially on native Brazilian testudines.

In Brazil, there are two native tortoise species, *Chelonoidis carbonarius* Spix, 1824 and *C. denticulatus*. These omnivore reptiles found in warm climates have thick skin and scales to prevent dehydration, and are able to absorb water through the cloaca before excretion. In these oviparous animals, sexual dimorphism is evident in sexually mature individuals: males have a concave dip in their plastron, whereas the female plastron is flat [6].

Given the significance of blood gas analysis in Brazilian reptiles, this study set out to establish emperature-corrected and uncorrected reference intervals for venous blood gas measurements in *Chelonoidis carbonarius*, and to compare values between females and males.

Methods

This study was submitted to SISBIO (System of Approval and Biodiversity Information) and approved by the Ethics Committee of University of Brasilia (UnB). Nineteen healthy adult *Chelonoidis carbonarius* specimens (8 males and 11 females) obtained from the Wildlife Triage Center of the Federal District (CETAS-DF) were used. Animals were rescued between 2019 and 2020.

Animals were submitted to clinical evaluation for determination of hydration status, level of activity and consciousness, and body condition score. All specimens were considered healthy based on clinical evaluation findings. Plastron size measurements were made using a measuring tape. Mean plastron length was 20.95 ± 5.85 cm in females and 28.13 ± 3.44 cm in males. Although their exact age could not be determined, all specimens had a similar size and were considered sexually mature for the species, as per Barros et al. [7].

Tortoises were numbered 1 to 19 using adhesive tape. Cloacal temperature (CT) was measured using a digital thermometer (B-Max Tp101, temperature range between -50°C and 300°C , precision 0.1°C , accuracy $\pm 1\%$, Shenzhen, GD, China) with the probe inserted 50 mm into the cloaca. Venous blood samples were collected between 11 a.m. and 4 p.m. at a mean environmental temperature of $26 \pm 3.55^{\circ}\text{C}$. Environmental temperature was controlled using a digital thermohygrometer (Incoterm 1005, temperature range between -50°C and 70°C , precision 0.1°C , accuracy $\pm 1\%$, Porto Alegre, RS, Brazil).

Animals were restrained in the supine position and blood samples (0.5 mL) collected from the dorsal coccygeal vein into 1 mL heparinized syringes (sodium heparin 5,000 IU/L; Eurofarma, Ribeirão Preto—SP, Brazil). Blood gas analyses were performed immediately after blood sample collection using a blood gas analyzer with automatic calibration and temperature control (Abbot VETSCAN i-STAT; Abaxis Europe, Griesheim, Germany).

The following parameters were measured: partial pressure of carbon dioxide (pCO_2), potential of hydrogen (pH), partial pressure of oxygen (pO_2), base deficit (BE_{ecf}), bicarbonate (HCO_3^-), total carbon dioxide (TCO_2), oxygen saturation (sO_2), sodium (Na), potassium (K), ionised calcium (iCa) and glucose (Glu). Temperature-corrected and uncorrected, pH, pCO_2 and pO_2 values were estimated. Since tortoises did not receive supplemental oxygen, the fraction of inspired oxygen (FiO_2) was set at 21% (atmospheric oxygen concentration) [8].

Data were entered into Excel spreadsheets (Microsoft Excel 16) and submitted to statistical analysis using BioEstat 5.3 [9]. Normal data distribution was confirmed using the Shapiro-

Wilk test. Extreme values were determined based on deviations and outliers excluded. Analysis of variance (ANOVA) was used to compare means between males and females. The paired t-test was used to compare mean temperature-corrected and uncorrected pH, pCO₂ and pO₂ values. The level of significance was set at 5%.

Results

Blood gas values obtained in this study are shown in Table 1. Oxygen saturation and Na levels were significantly higher ($p < 0.05$) in males. Remaining parameters did not differ significantly between males and females.

Comparative analysis of pH, pO₂, and pCO₂ values revealed significant differences in pO₂ in males, with lower values obtained following temperature correction. In contrast, all three parameters differed significantly ($p < 0.0001$) in females. Temperature-correction tended to yield higher pH and lower pO₂ and pCO₂ values, regardless of gender (Table 2).

Discussion

Studies addressing blood gas analysis in reptiles have been published by Harms et al. [5], Lewbart et al. [4], Lewbart et al. [3] and Ceretta et al. [2]. In those studies, blood samples were obtained from the subcarapacial venous sinus, jugular vein, coccygeal arch and ventral coccygeal vein respectively. No publications describing arterial blood gas analysis in reptiles were found, probably due to technical difficulties associated with arterial puncture in these animals.

Temperature-corrected pH and pCO₂ values measured in *C. carbonarius* in this study differed from values reported in *A. contortrix* (pH 0.18 and pCO₂ 1.2 mmHg) and *P. alleghaniensis* (pH 0.13 and pCO₂ 1.9 mmHg) [2]. In *Caretta caretta*, pH values of 0.13 and pCO₂ values of 28.7 have been described [5]. In this sample, pH values of 0.18 and 0.115 and pCO₂ values of 8.91 mmHg and 11.43 mmHg were obtained in females and males respectively. Higher temperature-corrected pH values have been reported in studies with other reptile species. In those studies, the lower the body temperature relative to the ambient temperature, the larger the difference in pH following temperature correction [2–5].

Studies comparing blood gas measurements in male and female reptiles have not been published to date. In this study, the values of sO₂ were significantly higher in males than in females. This finding may be explained by higher red blood cell counts and hematocrit in males in response to androgens. Erythropoiesis is inhibited by estrogens and stimulated by testosterone. Hence the higher number of red blood cells available for oxygen transport in the blood in males [10].

According to O'Malley [11], testudines are the vertebrates with the highest levels of bicarbonate in the blood, which helps to buffer lactic acid produced during anaerobic respiration. Findings of this study support that hypothesis. Bicarbonate levels of 8.1 and 16.9 mmol/L have been reported in *A. contortrix* and *P. alleghaniensis* respectively, compared to 22.47 mmol/L in male and 18.79 mmol/L in female *C. carbonarius* specimens in this sample [2]. Even higher blood bicarbonate levels (43.8 mmol/L) have been described in *C. mydas*, a marine turtle species which can survive prolonged apnea, suggesting aquatic testudines are able to neutralize higher amounts of lactic acid. Shorter apneic periods in *C. carbonarius* are consistent with the terrestrial habits of this species [4].

Ionised Ca, sodium, and pH values in *C. carbonarius* specimens in this study did not differ from *A. contortrix* or *P. alleghaniensis* [2]. However, lower iCa values have been reported in *C. mydas* [4]. Potassium values tend to be lower in tortoises than in snake species [2]. Differences in potassium levels may reflect the physiology of feeding and excretion in terrestrial reptile species. Reptiles excrete uric acid, an evolutionary adaptation to prevent water loss and

Table 1. Venous blood gas analysis data for male and female specimens of *Chelonoidis carbonarius*.

		MALES						
		MEAN	MEDIAN	STANDARD DEVIATION	STANDARD ERROR	MINIMUM	MAXIMUM	p M x F*
pH		7.43	7.41	4.54	1.6	7.39	7.48	0.2335
pH+TC (mmHg)		7.54	7.56.	1.61	5.69	7.27	7.76	0.213
PCO ₂ (mmHg)		33.39	33.60	12.41	4.39	11.1	55.9	0.1069
PCO ₂ +TC (mmHg)		21.96	22.1	9.73	3.44	7.2	38.5	0.1072
PO ₂ (mmHg)		66.25	66.50	12.01	4.24	45	80	0.6755
PO ₂ +TC (mmHg)		33.14	36	10.49	3.71	13	46	0.6021
BEecf (mmol/L)		-4.25	-2	7.13	2.52	-18	4	0.9947
HCO ₃ ⁻ (mmol/L)		22.47	23.1	3.38	1.2	17.1	27.3	0.1388
TCO ₂ (mmol/L)		23.57	24	3.26	1.15	18	28	0.1135
sO ₂ (%)		91.87	93.5	4.76	16.80	84	96	< 0.0001
Na (mmol/L)		126.71	127	1.7	0.60	124	129	0.0249
K (mmol/L)		3.34	3	0.81	0.29	2.4	4.5	0.3253
iCa (mmol/L)		1.17	0.95	0.73	0.26	0.33	2.44	0.8428
Glu (mg/dL)		45.57	48	7.57	2.67	34	56	0.6931
CT (°C)		26.24	26.7	3.55	1.25	19.2	30.5	0.6825
		FEMALES						
		MEAN	MEDIAN	STANDARD DEVIATION	STANDARD ERROR	MINIMUM	MAXIMUM	
pH		7.51	7.47	1.94	0.58	7.26	7.86	
pH+TC		7.69	7.59	1.96	0.59	7.52	8.03	
PCO ₂ (mmHg)		24.45	23.3	10.46	3.15	11.1	39.2	
PCO ₂ +TC (mmHg)		15.54	16.10	6.33	1.91	7.3	25.2	
PO ₂ (mmHg)		70.3	64.5	26.73	8.05	35	113	
PO ₂ +TC (mmHg)		37.30	29.00	18.57	5.59	15	67	
BEecf (mmol/L)		-4.27	-4	7.34	2.21	-14	10	
HCO ₃ ⁻ (mmol/L)		18.79	19.1	5.6	1.69	9.3	27.6	
TCO ₂ (mmol/L)		19.64	20	5.61	1.69	10	28	
sO ₂ (%)		90.3	93.5	10.15	3.05	68	100	
Na (mmol/L)		123.43	123	2.94	0.88	120	128	
K (mmol/L)		2.98	2.9	0.62	0.19	2.3	3.7	
iCa (mmol/L)		1.1	0.86	0.71	0.21	0.25	2.24	
Glu (mg/dL)		53.6	48	21.77	6.56	30	92	
CT (°C)		26.81	27.4	2.46	0.74	22.3	30.8	

*p M x F = p value in the mean comparison between males and females according to the paired t test, with a significance of 5%.

+TC = with temperature correction; BEecf = Base excess in extracellular fluid; Glu = Glucose; HCO₃⁻ = Bicarbonate; iCa = Calcium ion; K = Potassium; Na = Sodium; PCO₂ = Partial pressure of carbon dioxide; pH = Hydrogen potential; PO₂ = Partial pressure of oxygen; sO₂ = Oxygen saturation; CT = Cloacal temperature; TCO₂ = Total carbon dioxide.

<https://doi.org/10.1371/journal.pone.0299451.t001>

Table 2. Temperature-corrected (+TC) and uncorrected (-TC) pH, pO₂, and pCO₂ values obtained in male and female specimens of *Chelonoidis carbonarius* submitted to venous blood gas analysis.

	MALES			FEMALES		
	MEAN (-TC)	MEAN (+TC)	p*	MEAN (-TC)	MEAN (+TC)	p*
pH (mmHg)	7.43	7.54	0.0814	7.51	7.69	< 0.0001
PCO ₂ (mmHg)	33.39	21.96	0.1198	24.45	15.54	< 0.0001
PO ₂ (mmHg)	66.25	33.14	< 0.0001	70.3	37.30	< 0.0001

*p = p value in the mean comparison between males and females according to the paired t test, with a significance of 5%.

-TC = without temperature correction; +TC = with temperature correction.

<https://doi.org/10.1371/journal.pone.0299451.t002>

dehydration, and urate salts are composed of potassium in herbivorous species [12]. Although tortoises are omnivorous, they have a predilection for vegetarian diets (i.e., tend to be herbivores). Since urate salts consist primarily of potassium, serum potassium levels may be lower in tortoises when kidney function is normal [12, 13].

Conclusions

This study provides reference intervals for venous blood gas analysis in *Chelonoidis carbonarius* and reveals significant gender-related differences in oxygen saturation and sodium levels, with higher values in males. It was also possible to infer that the lower the body temperature relative to the environmental temperature, the larger the difference in pH following temperature correction.

Author Contributions

Data curation: Laís Velloso Garcia, Jairo Antonio Melo dos Santos.

Supervision: Líría Queiroz Luz Hirano.

Writing – original draft: Sofia Silva La Rocca de Freitas, Líría Queiroz Luz Hirano.

Writing – review & editing: Sofia Silva La Rocca de Freitas, Líría Queiroz Luz Hirano.

References

1. Teixeira Neto FJ. Equilíbrio Ácido-base e Eletrolítico em Anestesiologia. In: Massone F (ed) Anestesiologia Veterinária: Farmacologia e Técnicas. Guanabara Koogan, São Paulo. 2011; 215–226.
2. Ceretta AJ, Cannizzo SA, Smith DC, Minter LJ. Venous hematology, biochemistry, and blood gas analysis of free-ranging Eastern Copperheads (*Agkistrodon contortrix*) and Eastern Ratsnakes (*Pantherophis alleghaniensis*). *Plus One*. 2020, 15(2):1–15. <https://doi.org/10.1371/journal.pone.0229102>.
3. Lewbart GA, Hirschfeld M, Brothers JR, Muñoz-Pérez JP, Denking J, Vinuesa L, et al. Blood gases, biochemistry and haematology of Galápagos marine iguanas (*Amblyrhynchus cristatus*). *Conserv Physiol*. 2015, 3(1):1–7. <https://doi.org/10.1093/conphys/cov034>.
4. Lewbart GA, Hirschfeld M, Denking J, Vasco K, Guevara N, Garcia J, et al. Blood Gases, biochemistry, and hematology of galapagos green turtles (*Chelonia mydas*). *Plos One*. 2014, 9(5):1–7. <https://doi.org/10.1371/journal.pone.0096487>.
5. Harms CA, Mallo KM, Ross PM, Segars A. Venous blood gases and lactates of wild loggerhead sea turtles (*Caretta caretta*) following two capture techniques. *J Wildl Dis*. 2003, 39(2):366–374. <https://doi.org/10.7589/0090-3558-39.2.366>.
6. Guix JC, Fedullo DL, Molina FB. Masculinization of captive females of *Chelonoidis carbonaria* (Testudinidae). *Rev Esp Herp*. 2001, 15(1):65–75.
7. Barros MS, Resende LC, Silva AG, Ferreira Junior PD. Morphological variations and sexual dimorphism in *Chelonoidis carbonaria* (Spix, 1824) and *Chelonoidis denticulata* (Linnaeus, 1766) (Testudinidae). *Braz J Biol*. 2012, 21(1):153–161. <https://doi.org/10.1590/S1519-69842012000100018>.
8. Silveira A. Influência da umidade atmosférica sobre o mecanismo de transferência de gases através da interface água-atmosfera. 2004; Available from: <http://www.teses.usp.br/teses/disponiveis/18/18138/tde-29082016-091551/pt-br.php>.
9. Ayres M, Ayres Júnior M, Ayres DL, Santos AA. Aplicações estatísticas nas áreas das ciências biomédicas. Ong Mamiraua, Belém. 2007.
10. Schmidt SEM. Patologia Clínica em Aves. In: Cubas ZS, Silva JCR, Catão-Dias JL (eds) Tratado de Animais Selvagens, 2nd ed. Roca, Rio de Janeiro. 2014, 1577–1596.
11. O'Malley B. Clinical Anatomy and Physiology of Exotic Species. Editora Elsevier Saunders, London, 2005, 17–93.
12. Bentley P. J. Osmoregulation. In Gans C. & Dawson W. R. (eds.), *Biology of the reptilia*. Vol. 5, Physiology A. London: Academic Press. 1976, 356–408.
13. Dantzer W. H. Renal Function (with special emphasis on nitrogen excretion). In Gans C. & Dawson W. R. (eds.), *Biology of the reptilia*. Vol. 5, Physiology A. London: Academic Press. 1976, 447–496.